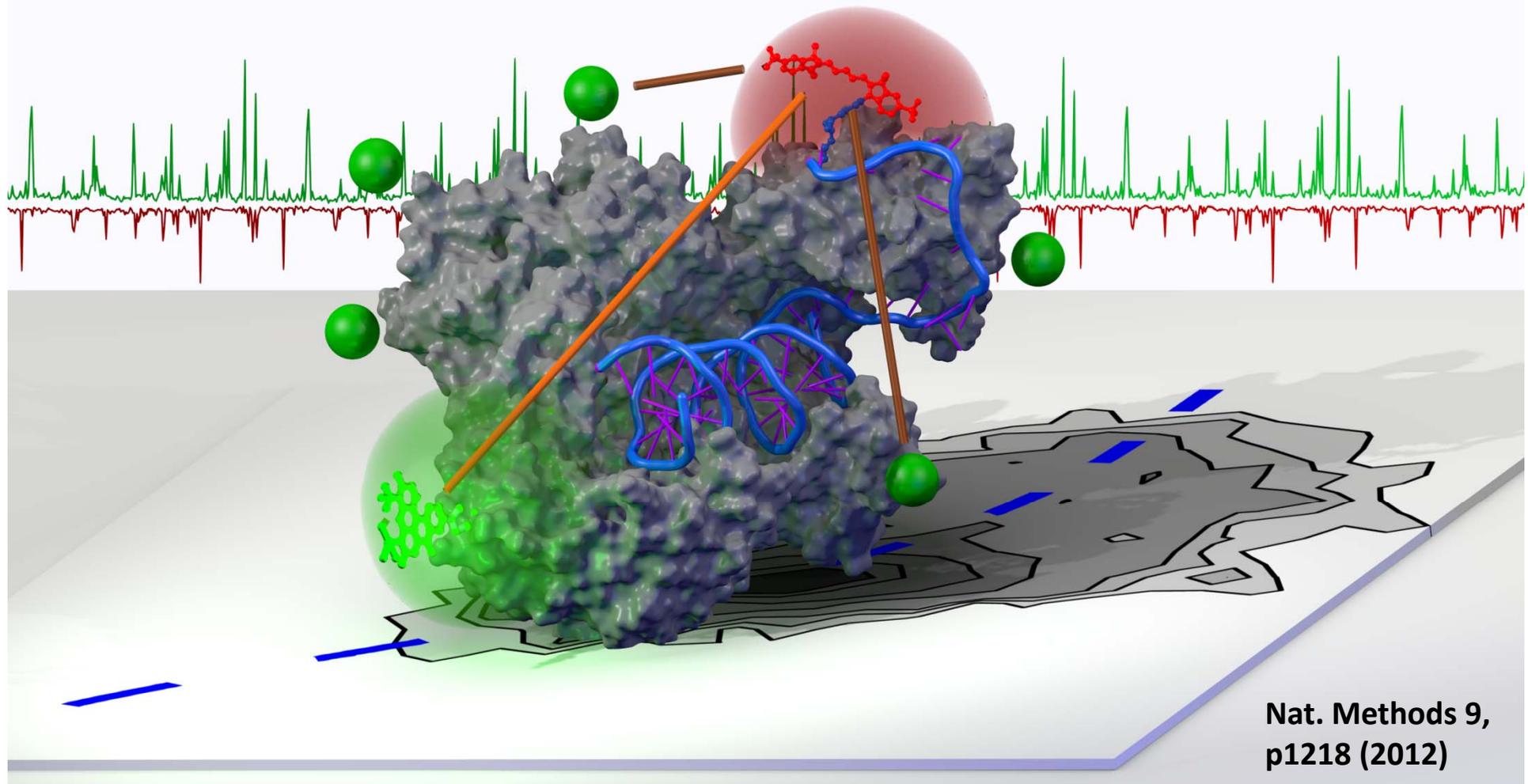


# Integrative dynamic structural biology with fluorescence spectroscopy

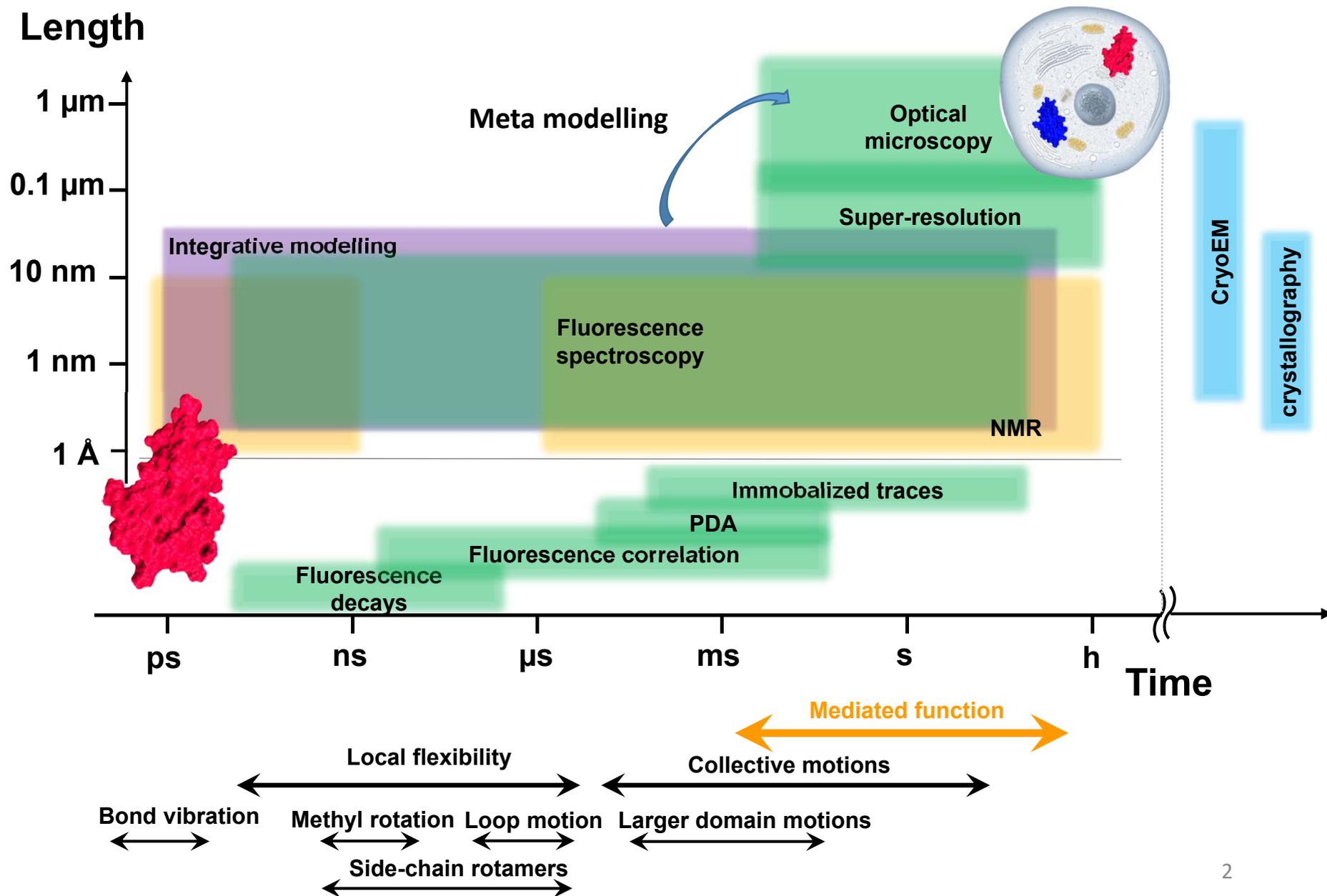
**Claus A. M. Seidel**

*Chair for molecular physical chemistry, Heinrich-Heine-Universität Düsseldorf; cseidel@hhu.de*



**Nat. Methods 9,  
p1218 (2012)**

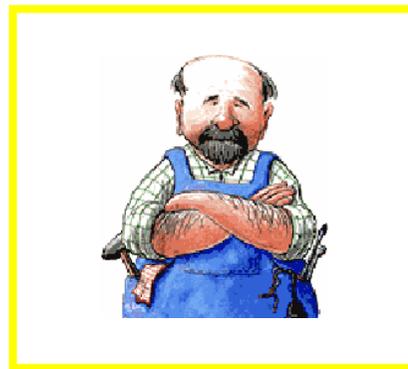
# Spatiotemporal models of living systems



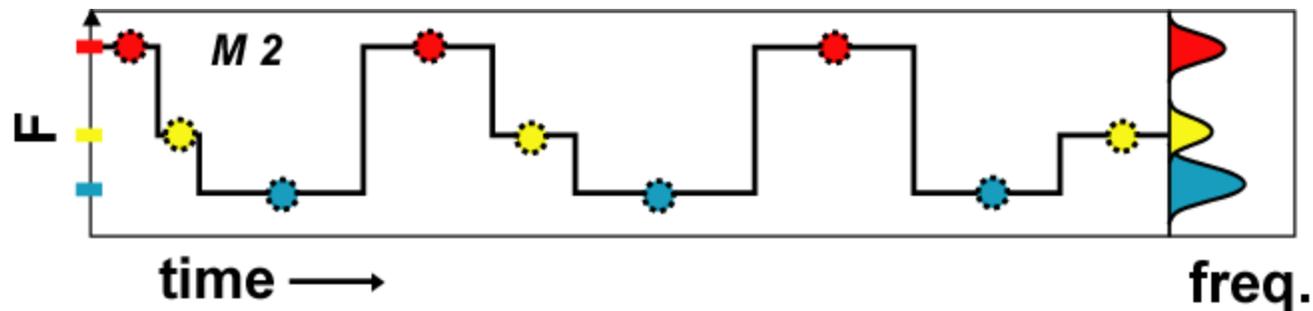
# Fluorescence dimensions and protein information

## Experiment types:

- single-molecule (variety of methods) to probe:
  - heterogeneity
  - structural information: proximities and distances
  - dynamics (from picoseconds to minutes): state connectivities



**Chemistry:**  
Purification..., but  
dynamic changes  
in time.

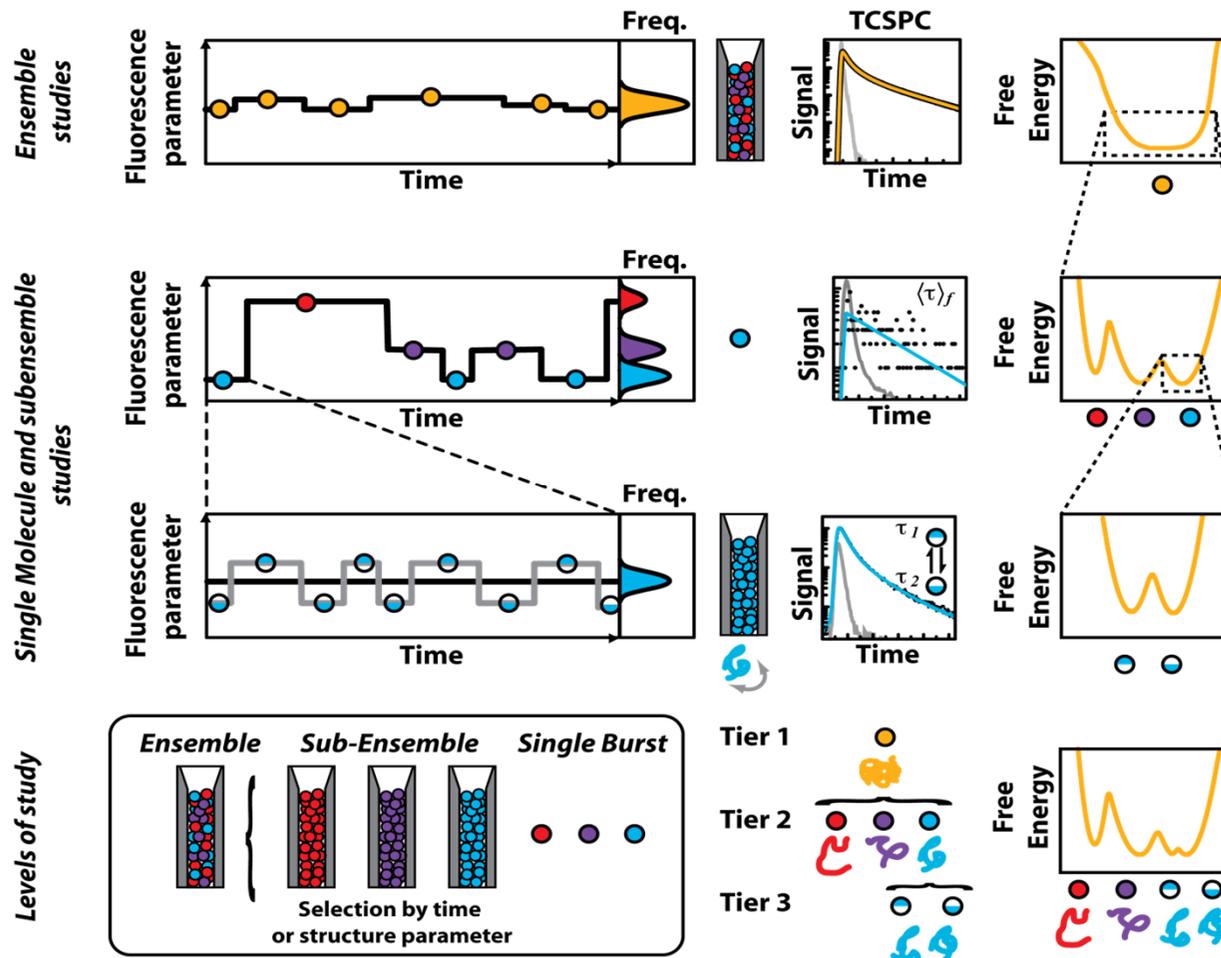


**Physics:**  
Isolation in time  
via SMD.

# Fluorescence dimensions and protein information

## Experiment types (single photon counting (Poisson statistics!!)):

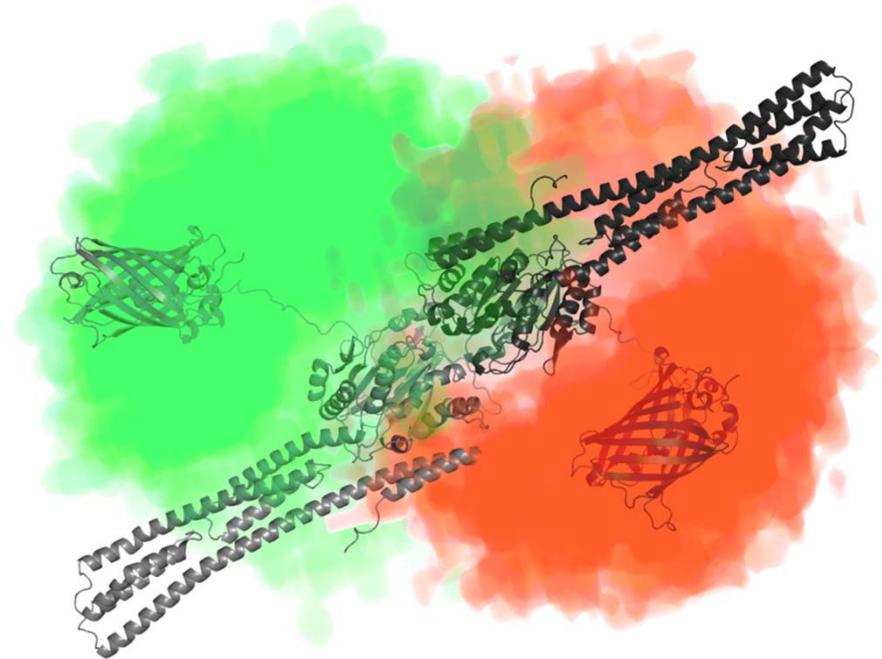
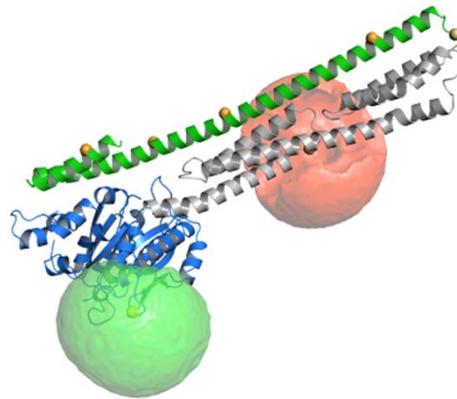
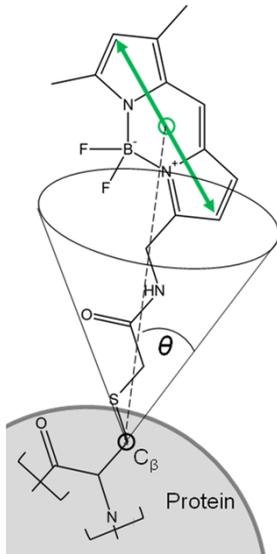
- single-molecule (variety of methods)
- sub-ensemble (like particle averaging in EM)
- ensemble (ultimate precision): 60 million photons



# Fluorescence dimensions and protein information

## Probes:

- Intrinsic, extrinsic, genetic:
  - aromatic amino acids (TRP, TYR)
  - selective labeling by small organic dyes,
  - fluorescent proteins



**Consider linkage:** Influence on spatial dye distribution

# Fluorescence dimensions and protein information

- Local structure

Spectral changes (polarity probes, ratiometric probes)	Localization of the segment
Quenching	Proximity of other segments, accessibility of sites
Polarization, anisotropy	Order parameters (local flexibility) [1]

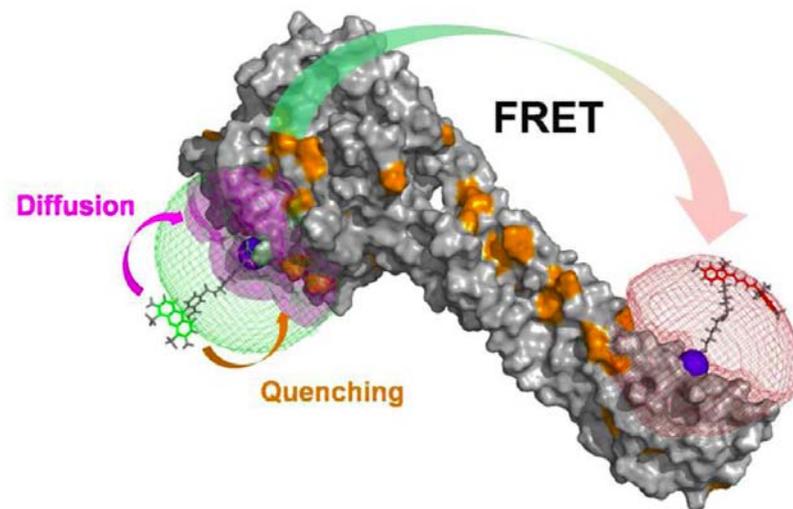
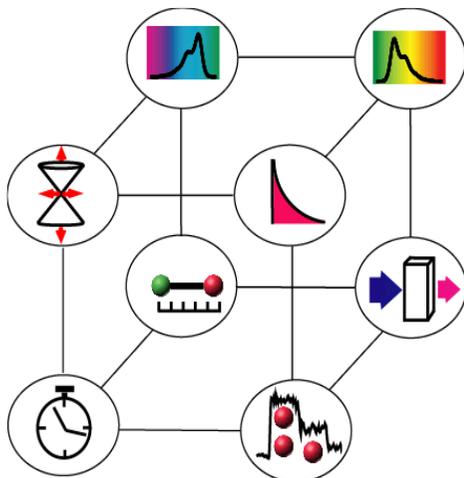
- Global structure

Förster energy transfer (FRET)	Inter-dye distances [2]
Diffusion (translation and rotation)	Shape

- Dynamic exchange

Time-resolved detection (ps-ms/ min)	Kinetic exchange networks
--------------------------------------	---------------------------

**A N-dimensional vector with all observables characterizes each protein state**



[1] Möckel et. al.  
J. Phys. Chem. B 122,  
in press (2018)

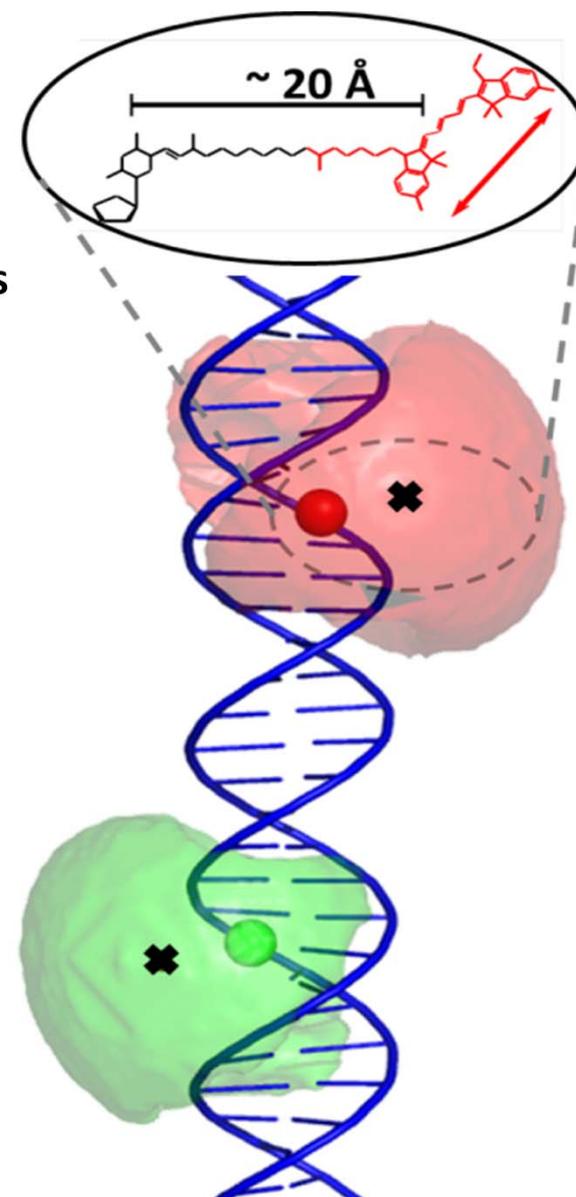
[2] Peulen et. al.  
J. Phys. Chem. B 121,  
8211 (2017) 6

# Quantitative high-precision FRET (hpFRET) analysis

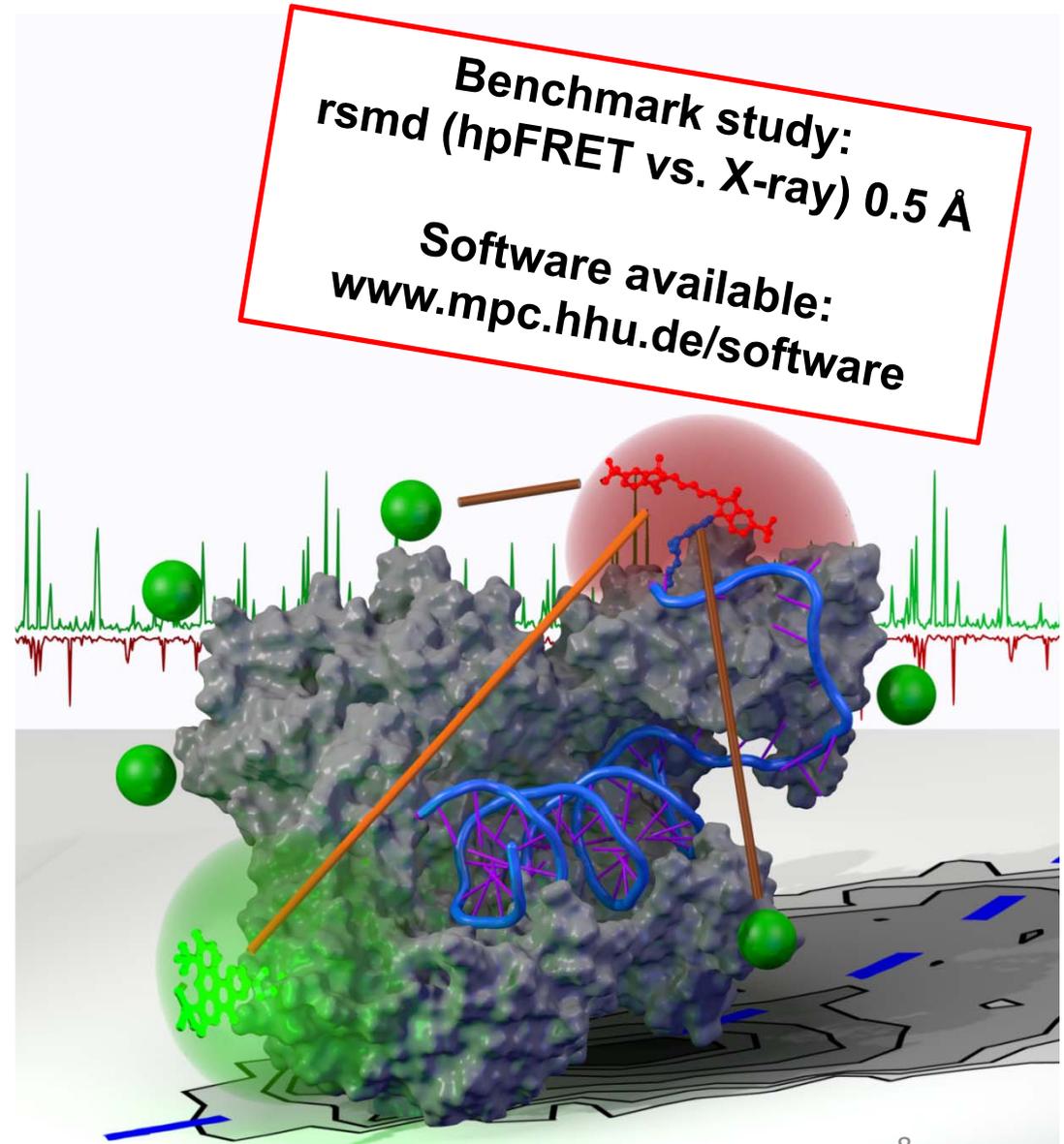
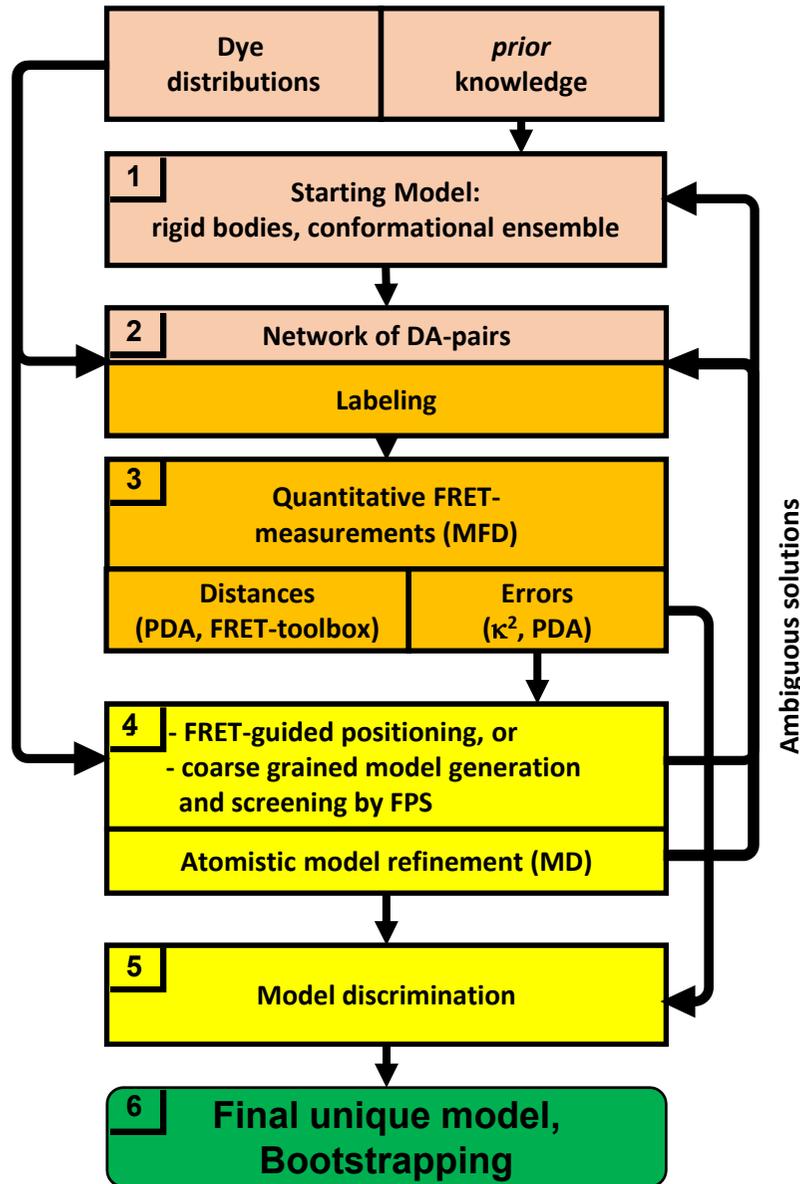
- Multiparameter Fluorescence Detection (MFD)
- Detect dynamic averaging
- Rigorous error analysis
- Dye properties: **Accessible volume (AV) simulations**
- Hybrid approach: **Combination with computer simulations**

## Structural models in data bases (wwPDB)

- Precision
- Accuracy
- Confidence levels
- **Dynamic / heterogeneous systems:**  
Use Single-molecule advantage:
  - multiple structures in parallel
  - kinetics: no need for synchronization
- Kinetics: reaction pathways
- Solution conditions, room temperature
- Large systems possible
- Combination with microscopy: *in vivo* option



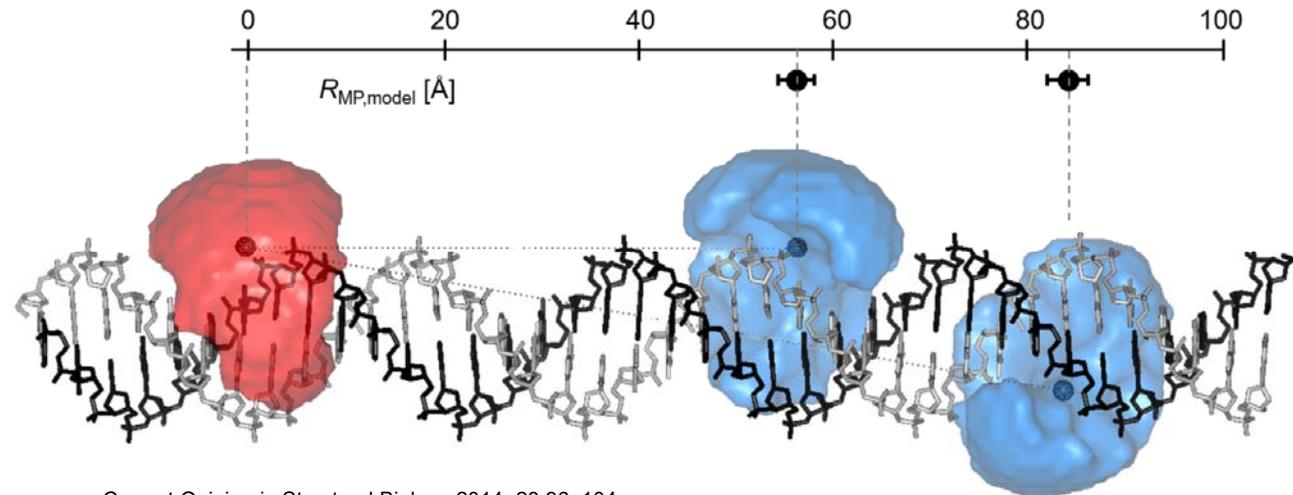
# High-precision FRET (hpFRET): 6 steps to FRET-restrained structural biology



# Multi-laboratory challenge demonstrates the accuracy and precision of FRET

## Blind study:

10 distinct DNA rulers were measured in 20 laboratories in the world

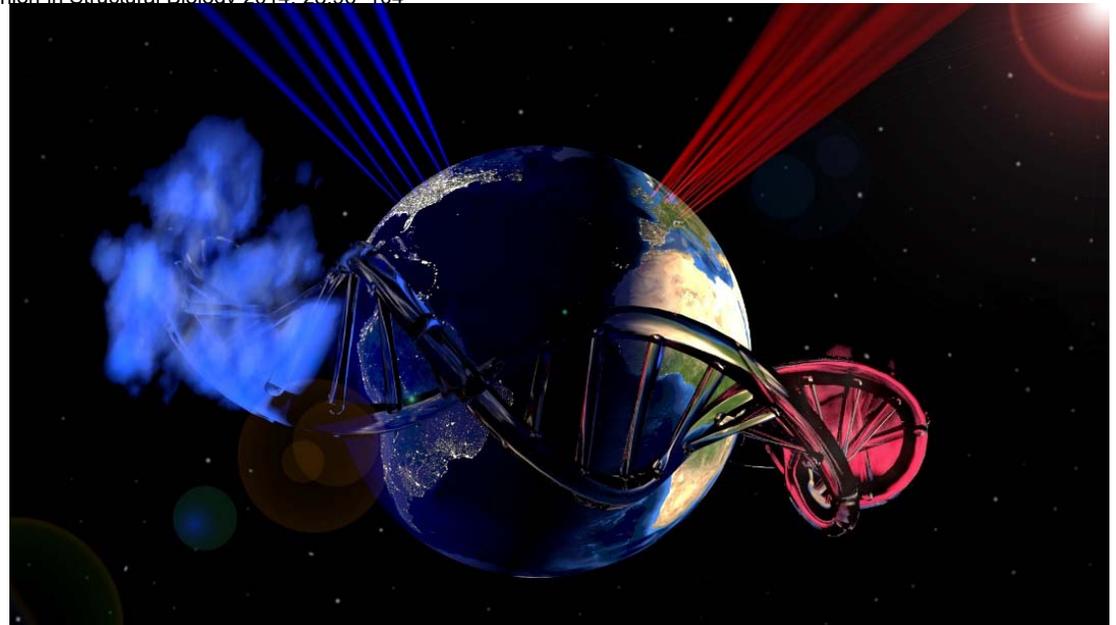


Current Opinion in Structural Biology 2014, 28:96–104

Standard deviation between the exp. FRET efficiencies:  
 **$\Delta E = \pm 0.02$  and  $\pm 0.05$**

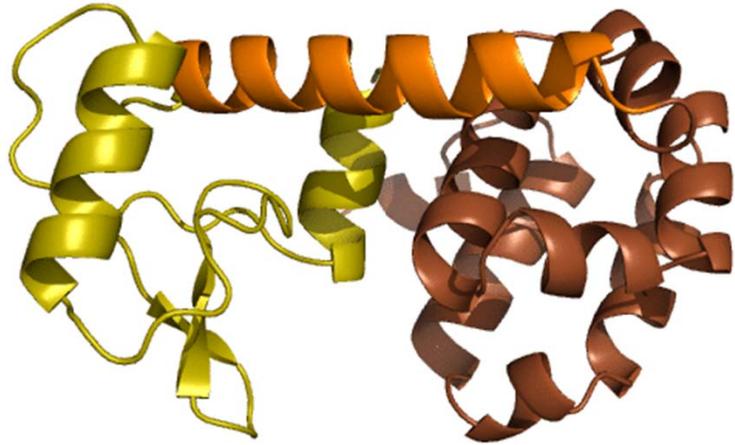
Rel. deviations between mean exp. and model distances  $\langle R \rangle$ :  
 **$\Delta R/R_{mod} = 0$  and  $\pm 0.05$**

**→ well within the expected error**



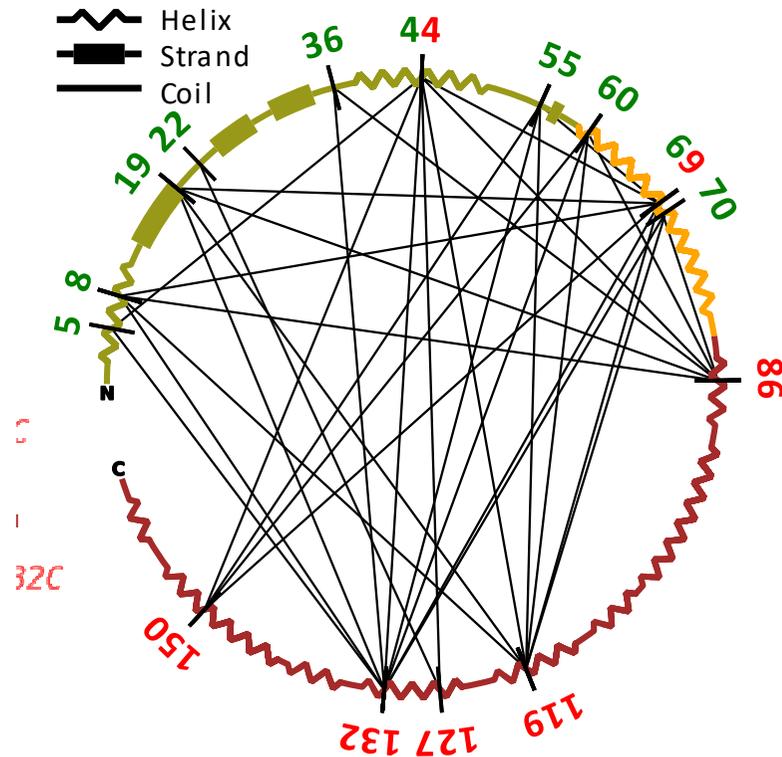
# Proteins with more than one conformation

---



## New challenge for CASP community:

- Predict and describe proteins in distinct conformations
- Data-assisted modelling using fluorescence information

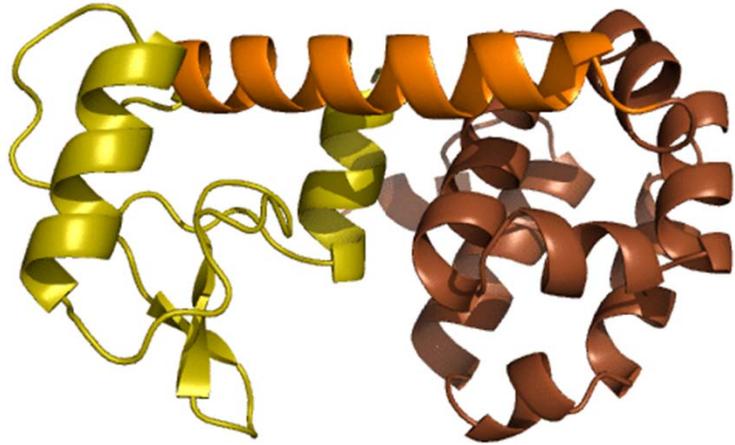


## FRET measurements

- Sparse data- combination with computer simulations
- Euclidean distance information between two points with upper and lower bound

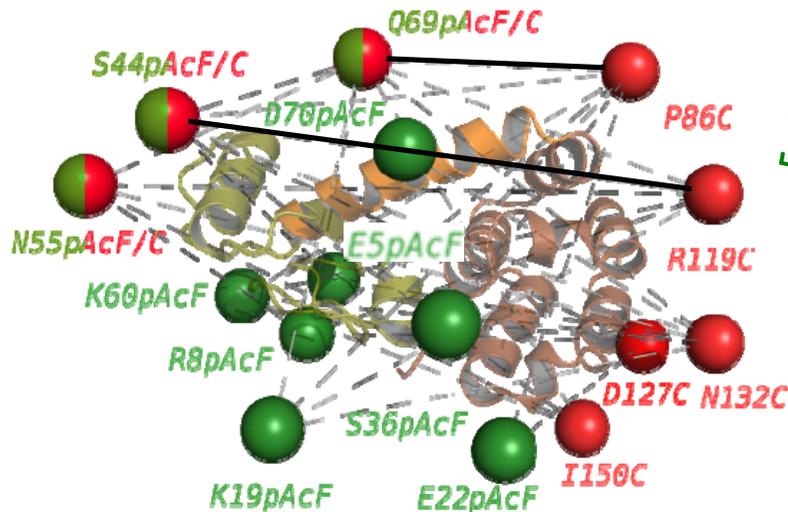
# Proteins with more than one conformation

---



## New challenge for CASP community:

- Predict and describe proteins in distinct conformations
- Data-assisted modelling using fluorescence information



## FRET measurements

- Sparse data- combination with computer simulations
- Euclidean distance information between two points with upper and lower bound
- Dynamics

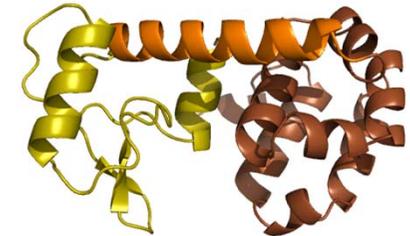
# Multiple structures and complexes: characterized on the fly

---

**Systems**                      **States**   **Relaxation times**

## Proteins with 2 domains

SNARE Protein Syntaxin 1	2	(700 $\mu$ s)
PGK	>2	(fast)
T4 Lysozyme	>3	(3, 200 $\mu$ s)
IF3 (ribosomal initiation factor)	>3 (7)	(2 $\mu$ s, 300 $\mu$ s)



## NA binding proteins

HIV-1 RT Primer/Template complexes:	3	(slow)
KlenTaq Polymerase	>3	(slow)
Klenow Fragment DNA Pol1	>5	(< 100 $\mu$ s)
BsoBI (restriction enzyme)	2-3	(10 $\mu$ s)
MutS (mismatch repair)	2	(slow)
Mononucleosomes	>3	(slow)

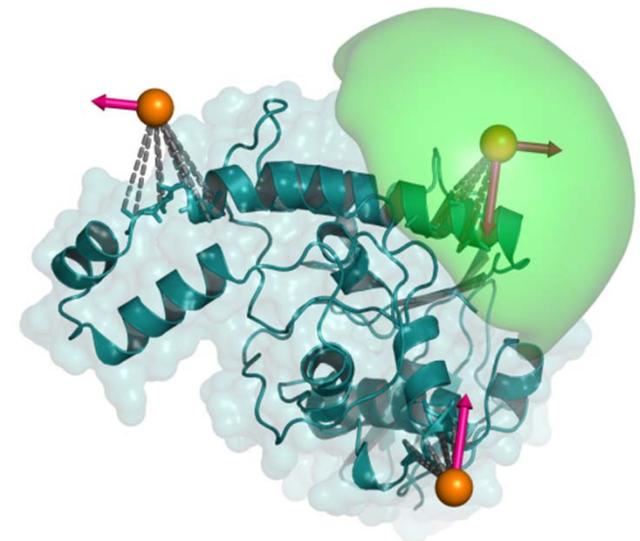
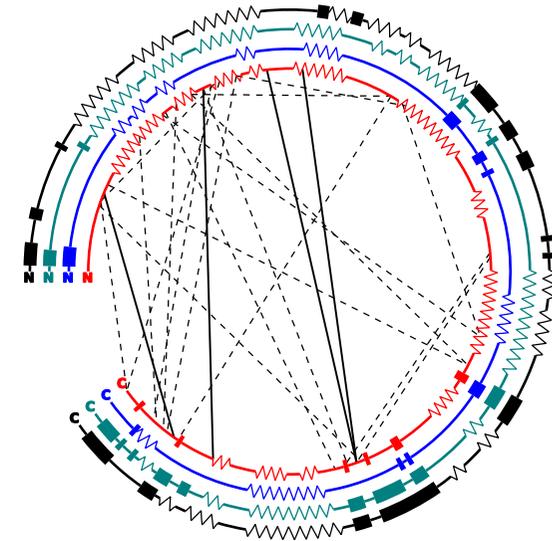
## Large GTPases

hGBP1	>2	(2, 20, 300 $\mu$ s)
Holliday junction (DNA 4W-Junction)	2(4)	(100 $\mu$ s)
Hairpin ribozyme (RNA 4W-Junction)	4	(100 $\mu$ s)

Intrinsically disordered proteins                      .....

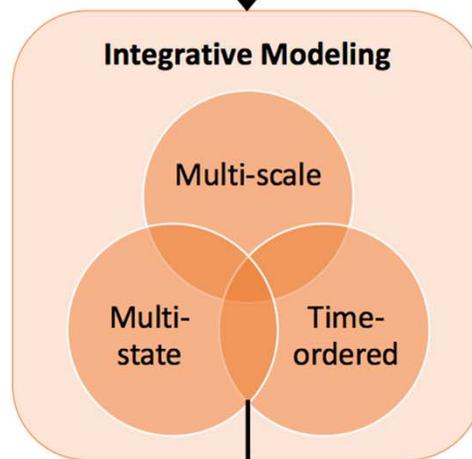
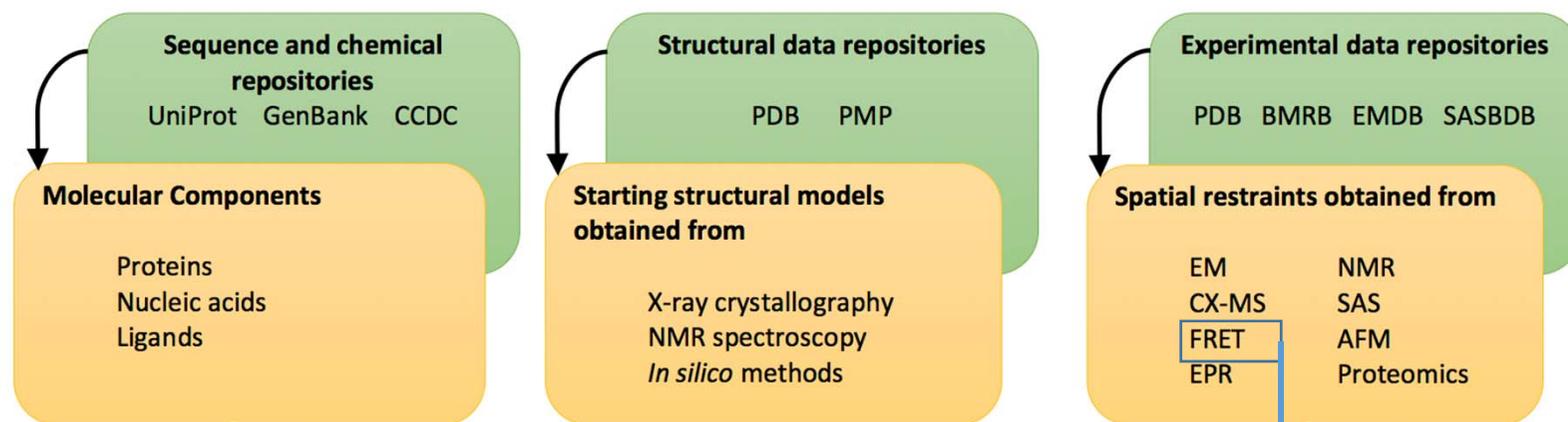
# Next generation of FRET toolkit: FPS 2.0 (tbd @ Poster)

1. **Finding most informative FRET-pairs:**  
FRET networks for least experimental work
2. **FRET-restrained optimization of structural models:**  
Beyond rigid body docking and simple model selection  
-> Targeted structural sampling by FRET guiding
3. **Estimation of accuracy:**  
Can we trust the FRET-restrained structural model?  
-> a crossvalidation approach analog to X-ray:  
 $R_{\text{free}}$  and
4. **Archiving of the I/H models to share information:**  
Generation of documentation: Fluorescence dictionary



-> deposition in PDB-Dev

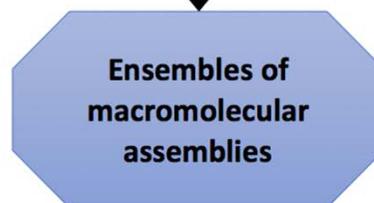
# PDB-Dev: New archive of structural models obtained through integrative/hybrid (I/H) methods



**Collaboration with PDB:**  
Brinda Vallat, Cathy Lawson,  
John Westbrook and Helen Berman.

Since 25.5.2018:

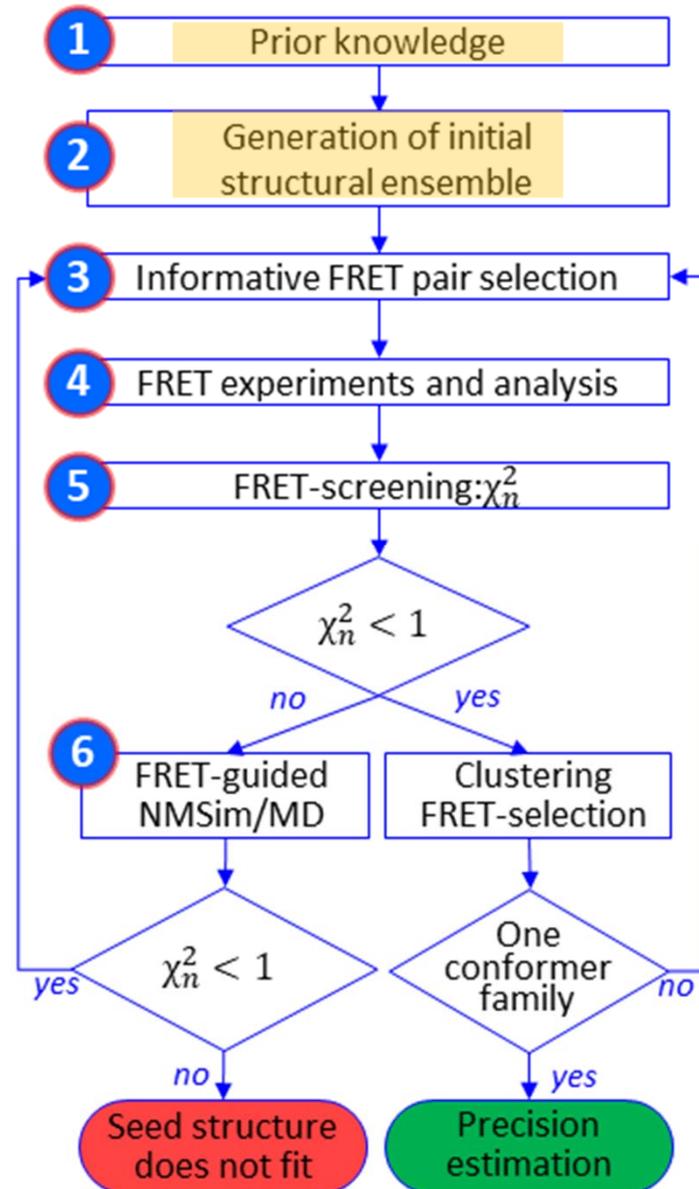
**First version of the fluorescence dictionary**  
<https://github.com/ihmwg/FLR-dictionary>



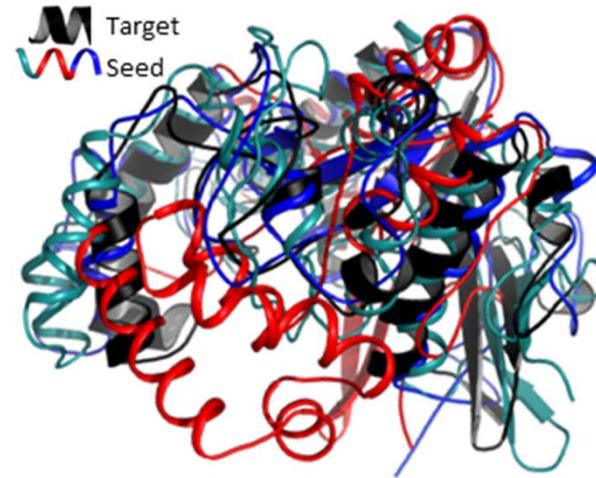
[https://github.com/ihmwg/IHM-dictionary/blob/master/dictionary\\_documentation/figures/IHM\\_dictionary\\_overview.png](https://github.com/ihmwg/IHM-dictionary/blob/master/dictionary_documentation/figures/IHM_dictionary_overview.png)

# Selection of the most informative FRET pairs

## a FRET-assisted modelling workflow

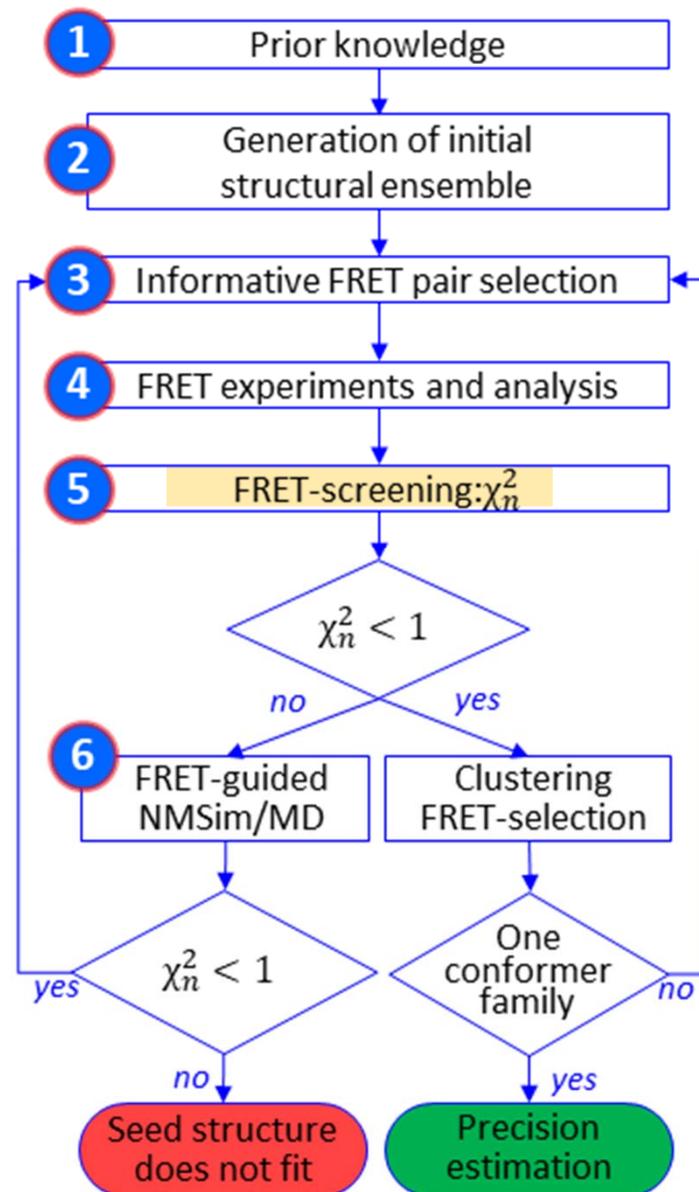


## CASP11 target T0806: YaaA (PDB ID 5CAJ)

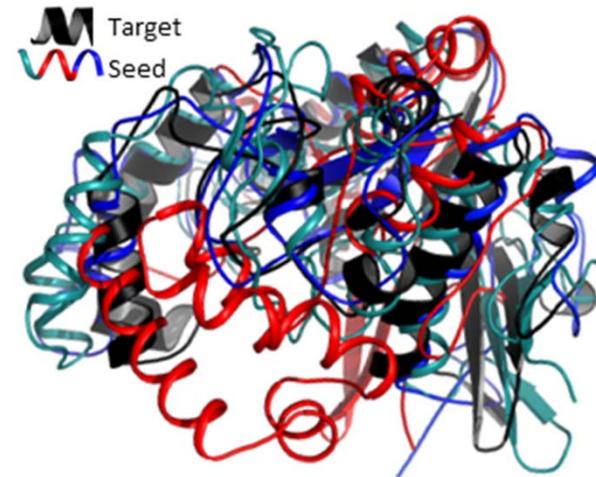


# Probing the accuracy of FRET-restrained structural models

## a FRET-assisted modelling workflow



## CASP11 target T0806: YaaA (PDB ID 5CAJ)



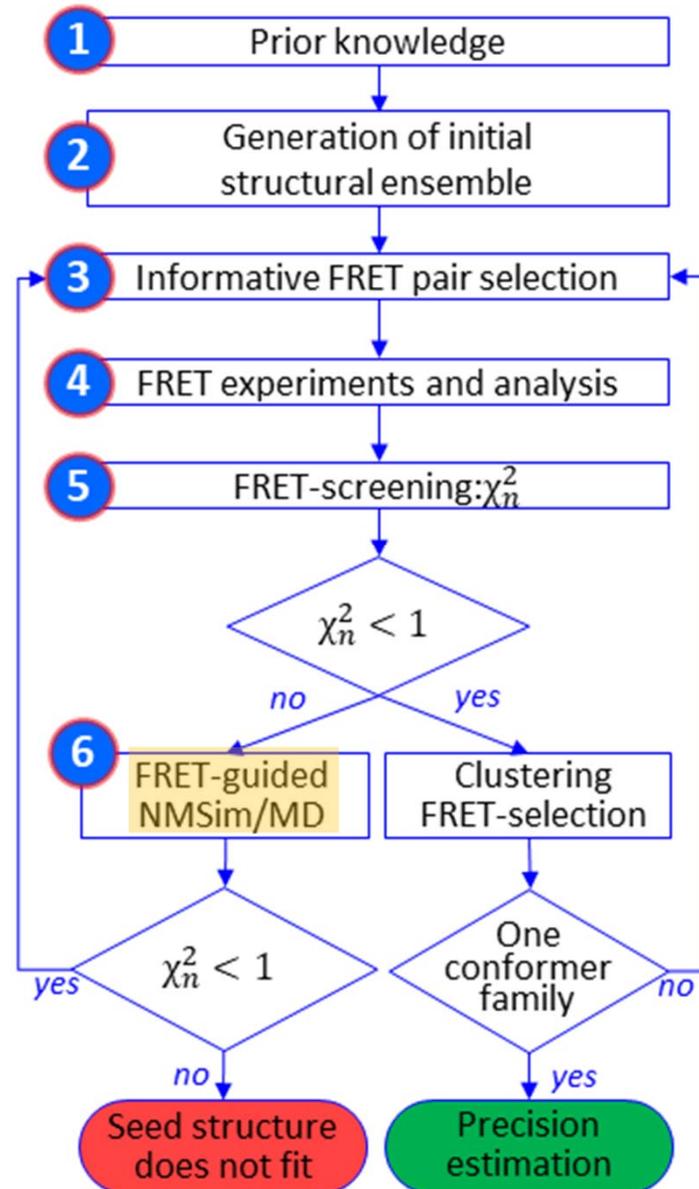
Quality parameter by cross validation:  $\chi_n^2$

$$\chi_n^2 = \chi^2 / \chi_p^2 = 68\%$$

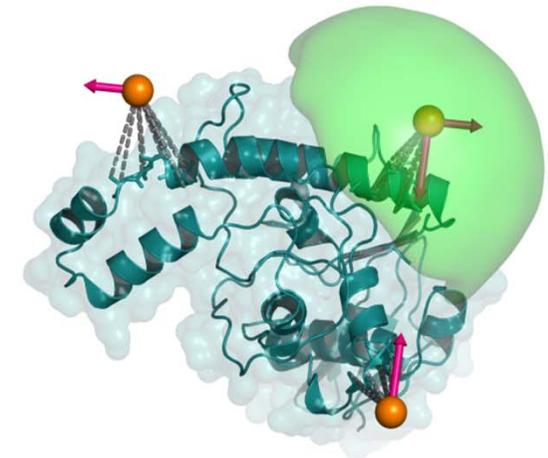
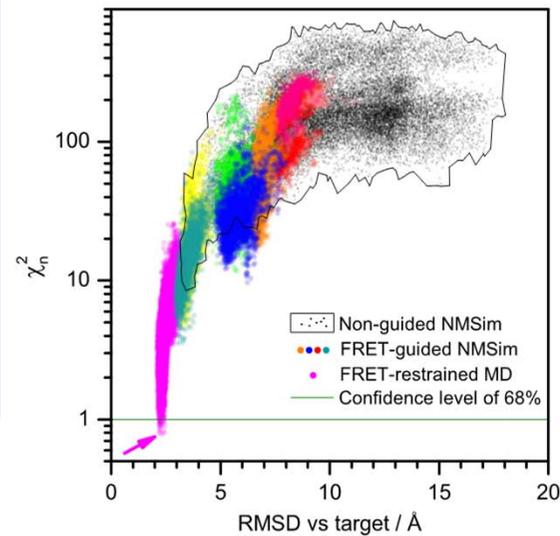
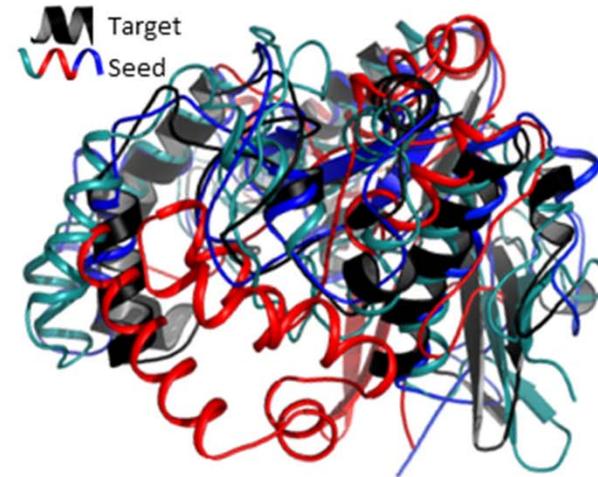
$$\begin{aligned} \chi_p^2 &= \text{Inv. } \chi^2(p, N_{dof}) \\ &= \frac{2^{-N_{dof}/2}}{\Gamma(N_{dof}/2)} p^{-N_{dof}/2-1} e^{-1/(2p)} \end{aligned}$$

# Accuracy of FRET-guided structural models

## a FRET-assisted modelling workflow

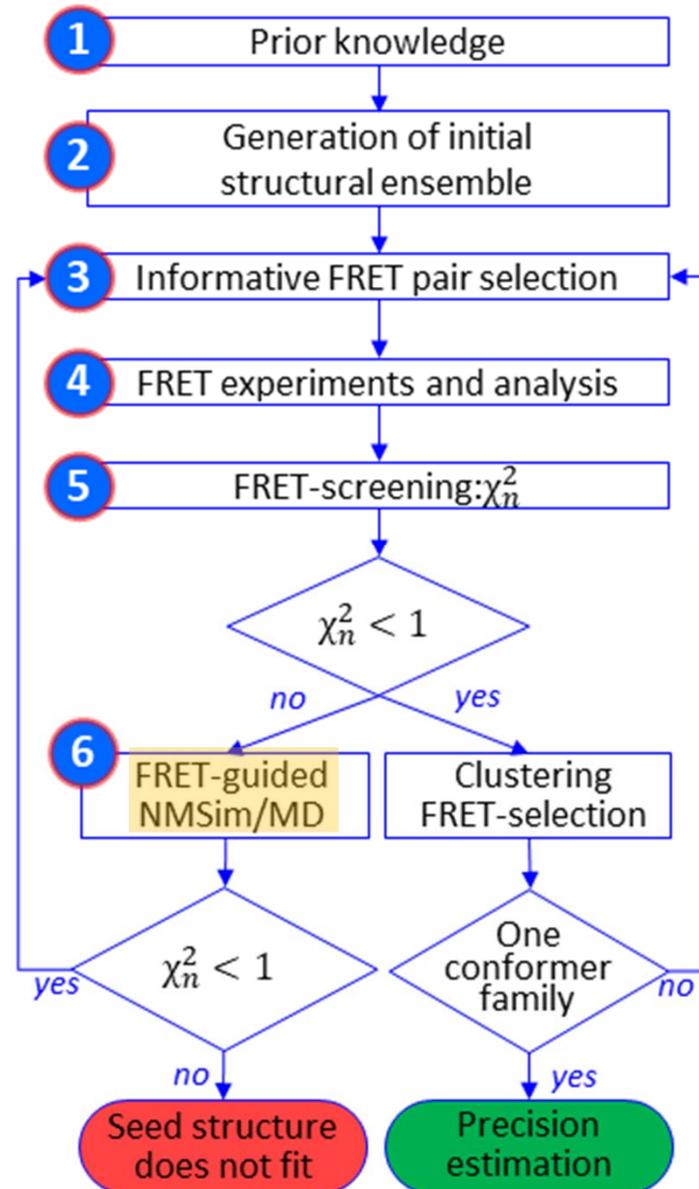


## CASP11 target T0806: YaaA (PDB ID 5CAJ)

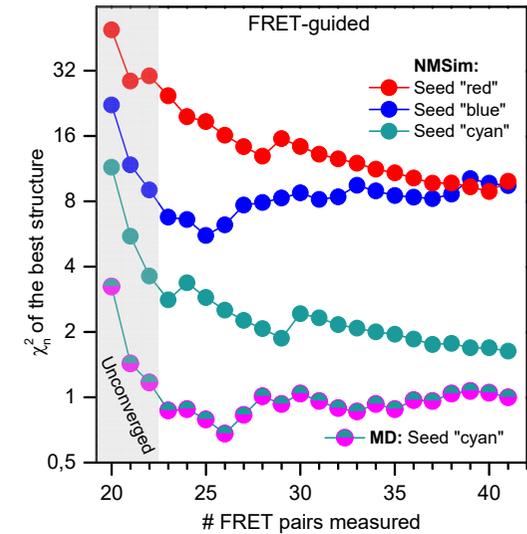
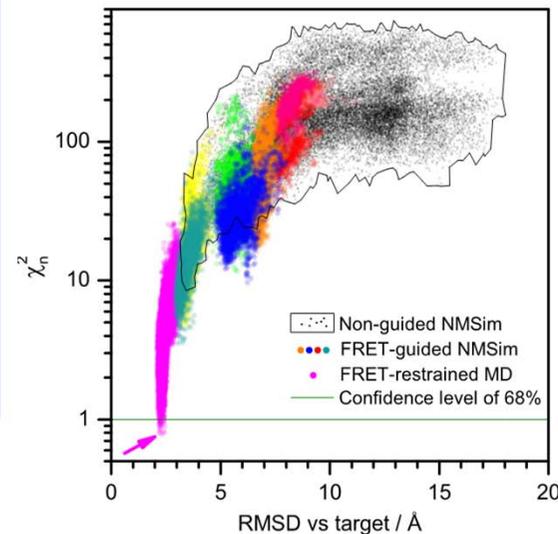
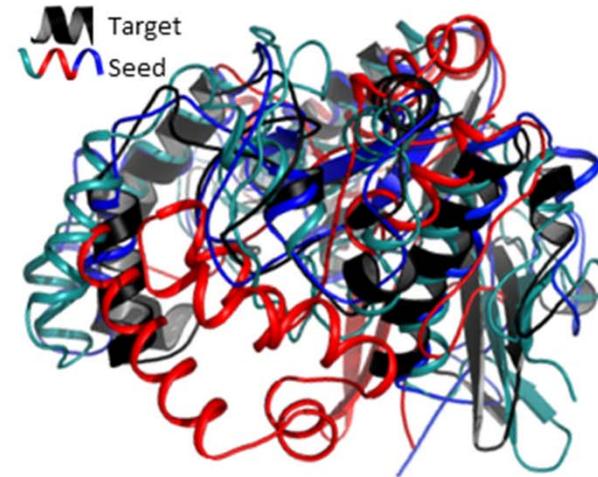


# Accuracy of FRET-guided structural models

## a FRET-assisted modelling workflow

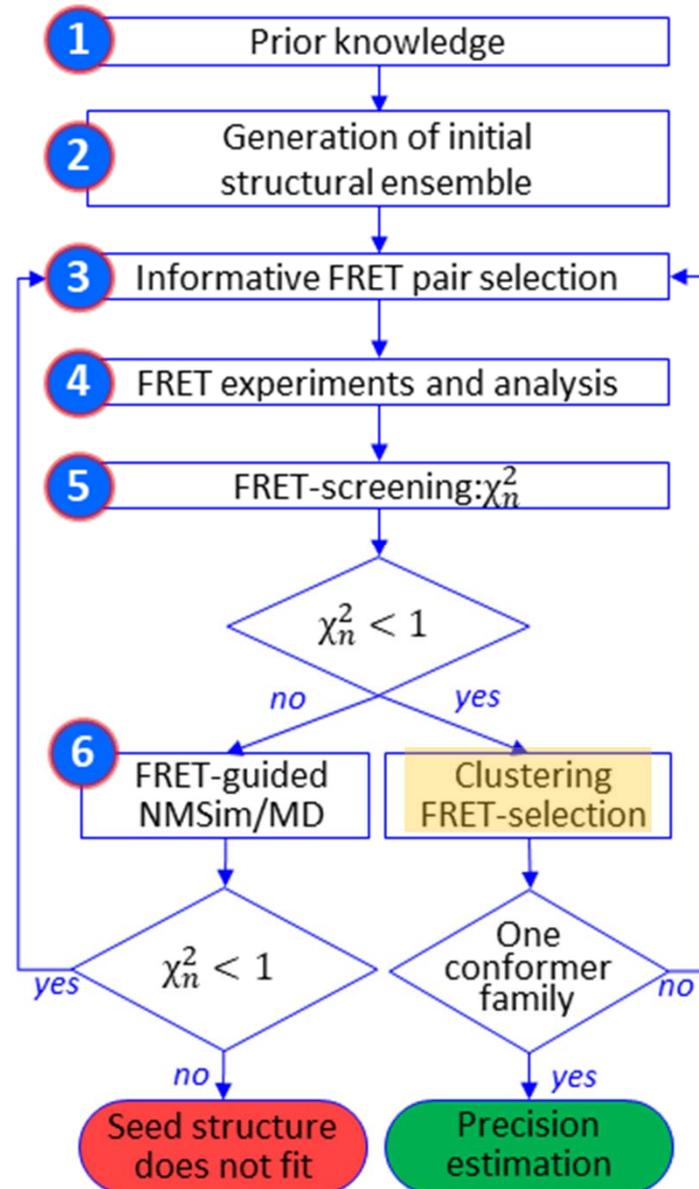


## CASP11 target T0806: YaaA (PDB ID 5CAJ)

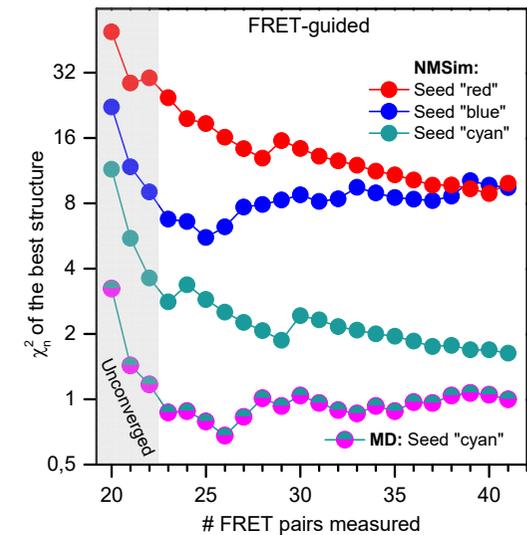
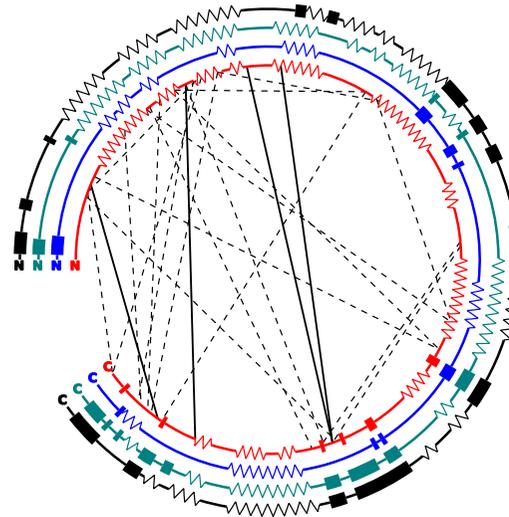
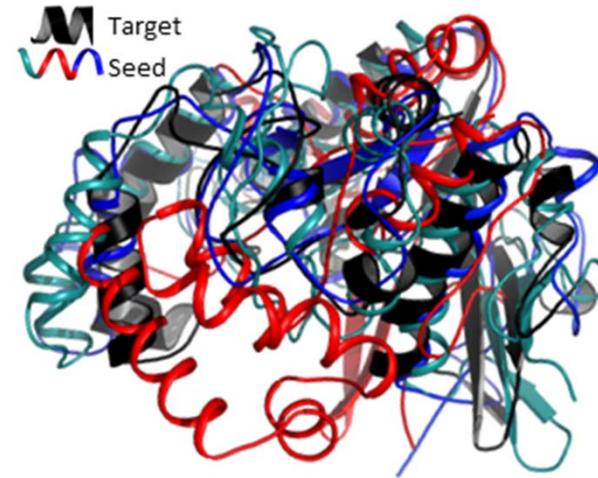


# Accuracy of FRET-guided structural models

## a FRET-assisted modelling workflow



## CASP11 target T0806: YaaA (PDB ID 5CAJ)



# Acknowledgements

## hybrid-FRET team:

Mykola Dimura, Thomas-Otavio Peulen,  
Christian Hanke.

## Soft- and Hardware:

[www.mpc.hhu.de/software](http://www.mpc.hhu.de/software)

<https://github.com/Fluorescence-Tools>

## Examples:

Curr. Opin. Struct. Biol. 40, 163–185 (2016)

Model Archive: DOI: 10.5452/ma-a2hbq

CASP13 webpage: [ProteinDynamics-FRET webinar](#)

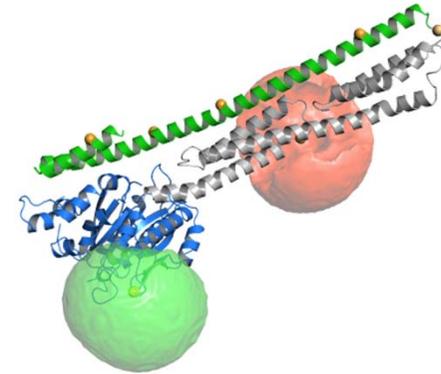


European Research Council

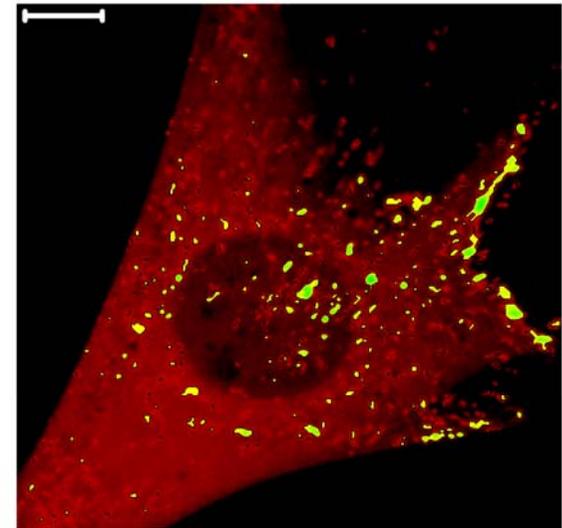
Established by the European Commission

hybridFRET :ERC Advanced Grant 2014

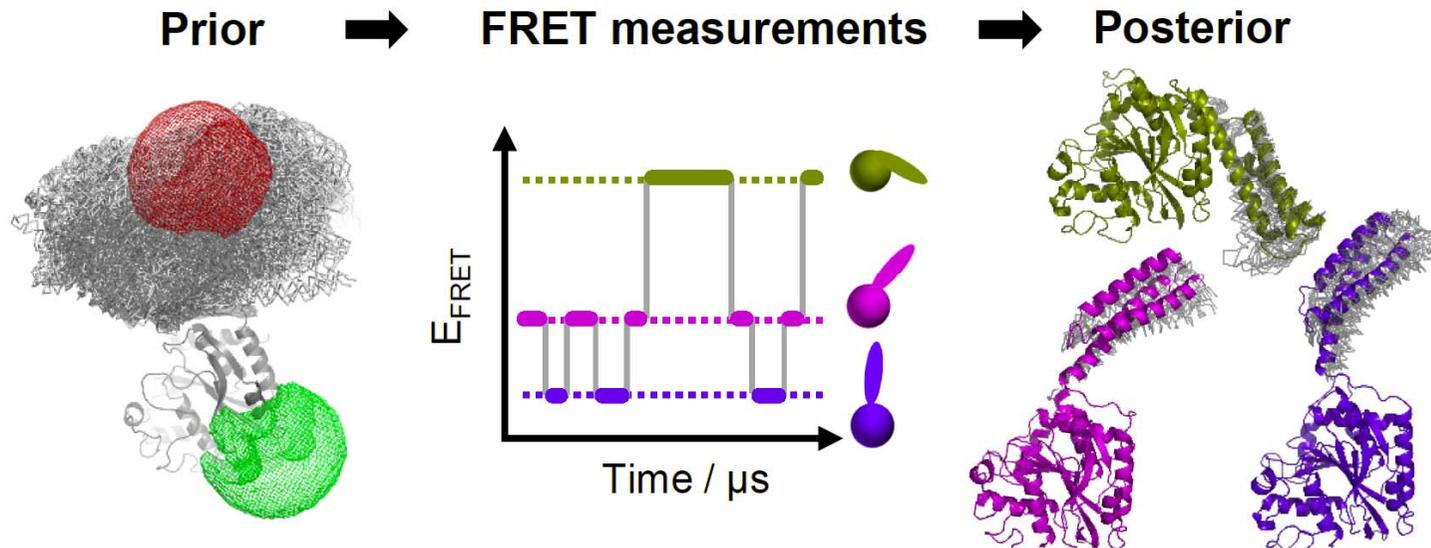
Watching ensembles



Live cell studies



# Exploring the capabilities of hybrid-FRET modeling: An *in silico* experiment:



**Curr. Opinion in Structural Biology** 40, 163–185 (2016)

and

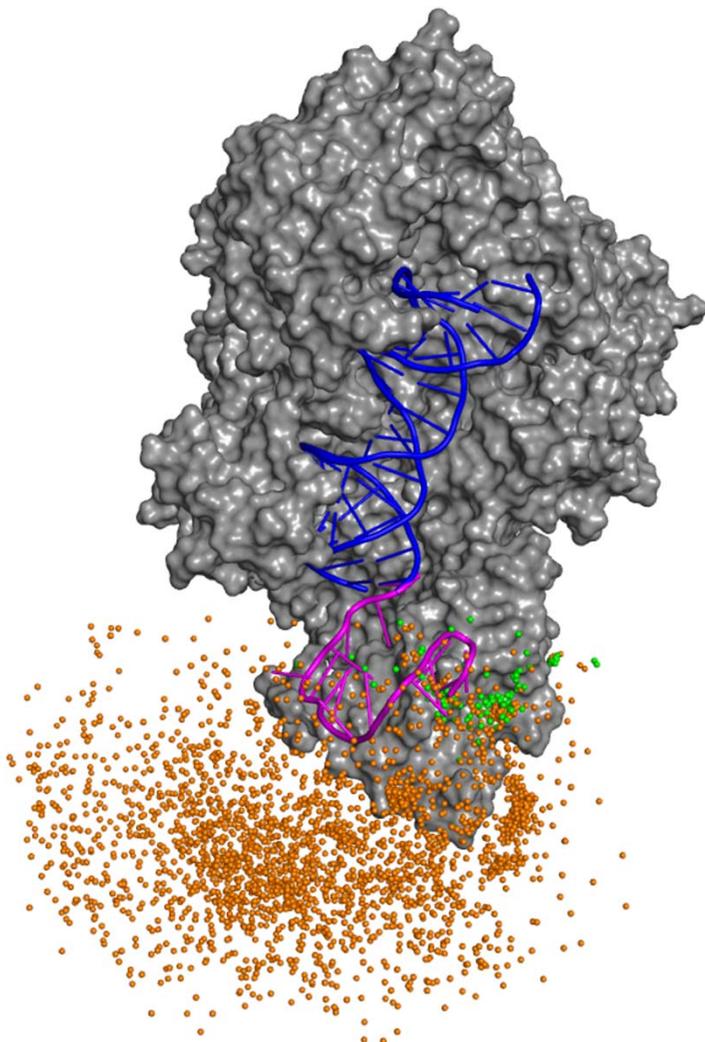
**Model Archive:** DOI: 10.5452/ma-a2hbq

with a comprehensive description, all tools and data

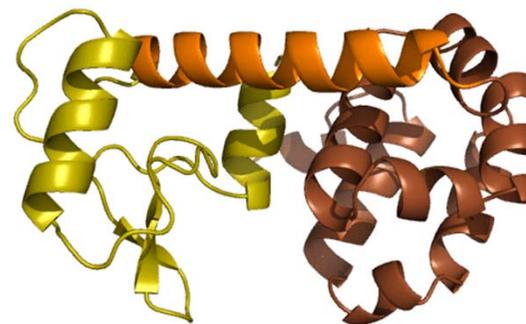
# Benchmark systems: Proteins

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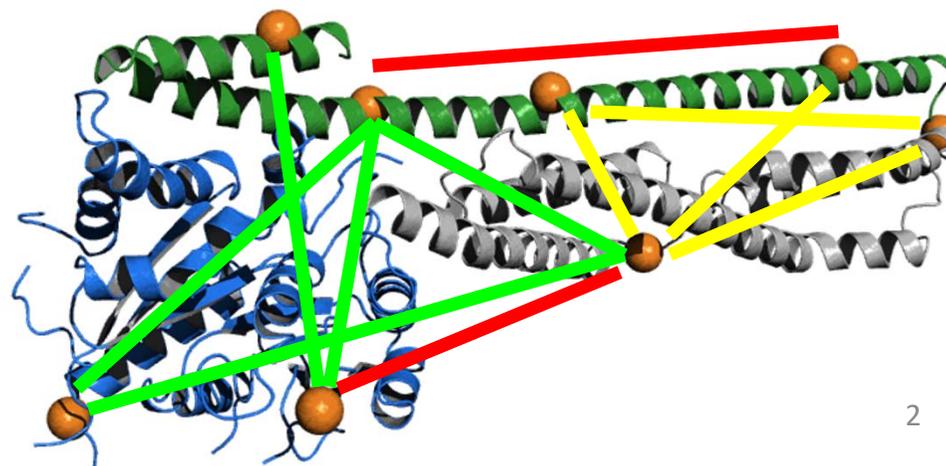
**HIV- RT : p/t**  
(*Nat. Methods* 9 p.1218 (2012))



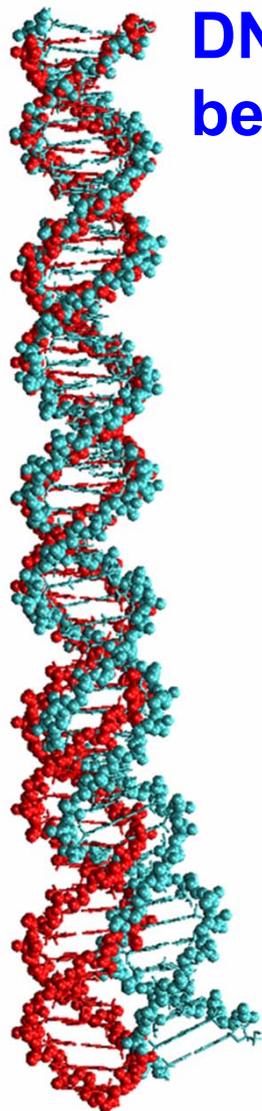
**T4 Lysozyme first structure of  
a new hidden state (200  $\mu$ s)**



**hGBP1 (new conformational state)**



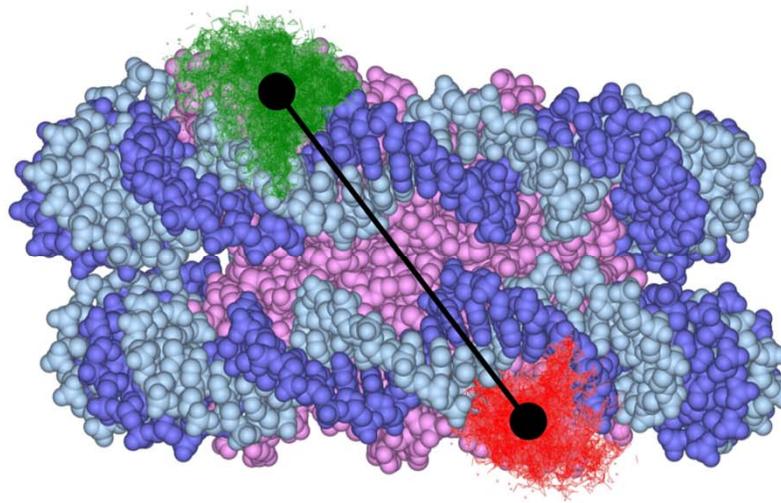
# Benchmark systems: Nucleic acids



**DNA:**  
bends ( $16^\circ$ ) and kinks

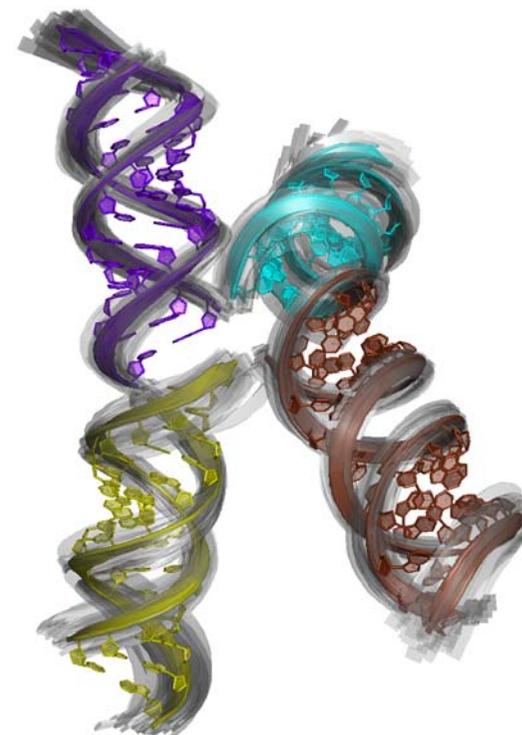
PNAS 105 p18773 (2008)

**Mononucleosomes:**  
disassembly pathway



PNAS 106 p15308 (2009)

**DNA + RNA: 3W**  
**+ 4W-junctions**



**in preparation:**

- > 260 FRET pairs
- 3 structures solved in parallel
- Precision  $2 - 4 \text{ \AA}$



# CASP target T0964 (CBM56) listed as [F0964](#)

- Protein name: CBM56
- Carbohydrate binding module from a  $\beta$ -1,3-glucanase (Bacillus circulans)
- Number of amino acids: 184 (694 - 877)
- Molecular weight: 18.936 kDa

```

710          720          730          740          750          760          770          780          790          694  700
TVNPTTAQVE VQGSVQLNAS VAPSNATNKQ VTWSVSGSSI ASVSPNGLVT GLAQGTTTVT ATTADGNKAA SATITVAPAP STVIVIGDEV KGLKKIGDDL
      810      820      830      840      850      860      870      877
LFYVNGATFA DLHYKVNNGG QLVNMAPTG  NGNYTYPVHN LKHGDTVEYF FTYNPGQGAL DTPWQTYVHG VTQGTPE
```





CASP target T0964 (CBM56) listed as [F0964](#)

## Data-assisted modeling

- Protein name: CBM56
- Carbohydrate binding module from a  $\beta$ -1,3-glucanase (Bacillus circulans)
- Number of amino acids: 184 (694 - 877)
- Molecular weight: 18.936 kDa



12 of 14 variants worked

Efficiency <E>	<RDA> / Å	$\pm \Delta<RDA>$ (Meta-analysis) / Å	$\pm \Delta<RDA>$ / Å	$\sigma_{DA}$ (total) / Å	$\pm \Delta\sigma_{DA}$ (total) / Å	$\sigma_{DA}$ (protein) / Å

# Dictionary for structural biologists

NMR	EPR	Fluorescence
2D NMR etc $\tau$ (NOESY mixing time)	double resonator	multi-parameter detection
T1 time, $1/\rho$	T1 time	fluorescence lifetime, $\tau$
order parameter S2	line shape analysis (High field EPR)	fluorescence anisotropy, $r$ rotational correlation time $\rho$ and corresponding amplitudes
distance information: <b>NOE</b> : short range <b>PRE</b> : Paramagnetic Relaxation Enhancement (PRE) long-range	PELDOR/ DEER (distance $r$ )	FRET (distance $R$ )
<b>Problems + advantages with the label</b> + Selectivity - Labelling strategies - Label position: (AV, rotamer libraries, MD simulations) - Orientation factor ( $\kappa^2$ )		
Line width analysis	line width analysis: ns-dynamics	PDA (Photon distribution analysis)
Relaxation dispersion analysis (transverse spin-relaxation, CPMG relaxation dispersion NMR experiments)	(DEER: frozen sample)	dynamic PDA
Correlation methods	(DEER: frozen sample)	FCS (Fluorescence correlation spectroscopy) <b>no gaps in the time axis over 10 orders of magnitude</b> SCCF (Species cross correlation function) in the MFD space