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# AlphaFold 2

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## **Protein folding at DeepMind**

- DeepMind is on a long-term mission to advance scientific progress
- We're interested in solving fundamental scientific problems using AI
- Protein folding is such an important fundamental problem that is well-suited for Al
- We're thankful that CASP is providing such an ideal experimental setup to evaluate progress





## Presenting the work of the AlphaFold team



+ Jonas Adler, Trevor Back, Stig Petersen, David Reiman, Martin Steinegger, Michalina Pacholska, David Silver, Oriol Vinyals, Koray Kavukcuoglu, Pushmeet Kohli, Demis Hassabis & with help from many others from across DeepMind

### Protein example: T1064 (ORF8)



**T1064 / 7jtl** 87.0 GDT (ORF8, SARS-CoV-2)

Ground truth Prediction

7JTL: Flower, T.G., et al. (2020) Structure of SARS-CoV-2 ORF8, a rapidly evolving coronavirus protein implicated in immune evasion. Biorxiv.



## **Protein example: T1044 (RNA Polymerase)**



6VR4: Leiman, P.G., et al. Virion-packaged DNA-dependent RNA polymerase of crAss-like phage phi14:2 (CASP target). (To be published.)

Ground truth **Prediction** 



Long-chain-trained model trained after the submission

#### **Individual domains**



T1041



T1042

T1043



## **Inductive Bias for Deep Learning Models**



### Convolutional Networks (e.g. computer vision)

- data in regular grid
- information flow to local neighbours



### Recurrent Networks (e.g. language)

- data in ordered sequence
- information flow sequentially



Graph Networks (e.g. recommender systems or molecules)

- data in fixed graph structure
- information flow along fixed edges



### Attention Module (e.g. language)

- data in unordered set
- information flow dynamically controlled by the network (via keys and queries)



## Putting our protein knowledge into the model

- Physical insights are built into the network structure, not just a process around it
- End-to-end system directly producing a structure instead of inter-residue distances
- Inductive biases reflect our knowledge of protein physics and geometry
  - The positions of residues in the sequence are de-emphasized
  - Instead residues that are close in the folded protein need to communicate
  - The network iteratively learns a graph of which residues are close, while reasoning over this implicit graph as it is being built







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# System Design

### Inputs

### Sequence databases

- UniRef90<sup>6</sup> (JackHMMER<sup>3</sup>)
- $\rightarrow$  BFD<sup>5</sup> (HHblits<sup>4</sup>)
- MGnify clusters<sup>2</sup> (JackHMMER<sup>3</sup>)

### **Structural databases**

- PDB<sup>1</sup> (training)
- PDB70 clustering (hhsearch<sup>4</sup>)

### All publicly available data.

[1] Berman et al., Nature Structural Biology (2003) doi:10.1038/nsb1203-980
[2] Mitchell et al., Nucleic Acids Research (2019) doi:10.1093/nar/gkz1035
[3] Potter et al., Nucleic Acids Research (2018) doi:10.1093/nar/gky448
[4] Steinegger et al., BMC Bioinformatics (2019) doi:10.1186/s12859-019-3019-7
[5] Steinegger et al., Nature Methods (2019) doi:10.1038/s41592-019-0437-4
[6] Suzek et al., Bioinformatics (2015) doi:10.1093/bioinformatics/btu739

Visualisations:

The PyMOL Molecular Graphics System, Version 2.0 Schrödinger, LLC. AS Rose, et al., Bioinformatics (2018) doi:10.1093/bioinformatics/bty419









### Embedding

templates

### Trunk

### Heads

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MSA picture inspired by: Riesselman, A.J., Ingraham, J.B. & Marks, D.S., Nature Methods (2018) doi:10.1038/s41592-018-0138-4

## **Template embedding**

- 4 templates used (from PDB70 clusters, searched with HHsearch<sup>1,2</sup>)
- Input features are sequences, side chains, and distograms
- Templates are processed in the same way as the residue-residue representation





Remmert, M., Biegert, A., Hauser, A., & Söding, J. (2012). HHblits: lightning-fast iterative protein sequence searching by HMM-HMM alignment. Nature Methods, 9(2), 173-175.
 Steinegger, M. et al. (2019). HH-suite3 for fast remote homology detection and deep protein annotation. BMC Bioinformatics, 20(1), 1-15.





 $\rightarrow$ 

End-to-end folding instead of gradient descent

Protein backbone = gas of 3-D rigid bodies (chain is learned!)



Image: Dcrjsr, vectorised Adam Rędzikowski (CC BY 3.0, Wikipedia)

 3-D equivariant transformer architecture updates the rigid bodies / backbone
 Also builds the side chains



Target: T1041



 $\rightarrow$ 

End-to-end folding instead of gradient descent

Protein backbone = gas of 3-D rigid bodies (chain is learned!)



Image: Dcrjsr, vectorised Adam Rędzikowski (CC BY 3.0, Wikipedia)

**3-D equivariant transformer architecture** updates the rigid bodies / backbone

• Also builds the side chains





 $\rightarrow$ 

End-to-end folding instead of gradient descent

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Image: Dcrjsr, vectorised Adam Rędzikowski (CC BY 3.0, Wikipedia)

 3-D equivariant transformer architecture updates the rigid bodies / backbone
 Also builds the side chains

**Iteration 3** 

Target: T1041





 $\rightarrow$ 

End-to-end folding instead of gradient descent

Protein backbone = gas of 3-D rigid bodies (chain is learned!)



Image: Dcrjsr, vectorised Adam Rędzikowski (CC BY 3.0, Wikipedia)





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End-to-end folding instead of gradient descent

Protein backbone = gas of 3-D rigid bodies (chain is learned!)



Image: Dcrjsr, vectorised Adam Rędzikowski (CC BY 3.0, Wikipedia)



## **Refinement in structure module**

 $\rightarrow$ 

Improves both accuracy and stereochemical quality





Target: T1041

6

### Relaxation

- The end result of iterative refinement is not guaranteed to obey all stereochemical constraints
- Violations of these constraints are resolved with coordinate-restrained gradient descent
- We use the Amber ff99SB force field<sup>1</sup> with OpenMM<sup>2</sup>



Orange: pre-relax Blue: post-relax

[1] Hornak, V. et al. (2006). Comparison of multiple Amber force fields and development of improved protein backbone parameters. Proteins: Structure, Function, and Bioinformatics, 65(3), 712-725.
[2] Eastman, P. et al. (2017). OpenMM 7: Rapid development of high performance algorithms for molecular dynamics. PLoS Computational Biology, 13(7), e1005659.



## Knowing where we are right

## $\mathsf{IDDT}\text{-}\mathsf{C}\alpha$ prediction from the last layer of the structure module



### Confidence calibration on CASP14 chains Median absolute error: 3.3 LDDT-Cα



Five models per chain, coloured by chain Excluding T1044 domains, T1088



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# How AlphaFold understands proteins



## **Biological context**

- > Computational structure prediction is typically underspecified
  - Oligomeric state, ligands, DNA-binding, experimental conditions, multiple conformations etc.
- Our networks implicitly models the missing context
- Ises a variety of physical and evolutionary information (e.g. profile-only is still pretty accurate)



AlphaFold / Experiment

TBM-hard, 98.2 GDT



AlphaFold (monomer prediction x3)

FM/TBM, 85.9 GDT

**Experimental structure** 

### **Interrogating the Network**

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T1038

6YA2: Bahat, Y., et al. First structure of a glycoprotein from enveloped plant virus. (To be published.)





T1038

6YA2: Bahat, Y., et al. First structure of a glycoprotein from enveloped plant virus. (To be published.)







6YA2: Bahat, Y., et al. First structure of a glycoprotein from enveloped plant virus. (To be published.)















T1080: Not yet in PDB





T1080







T1061: Not yet in PDB 3 copies of monomer prediction overlaid on crystal





T1061



T1061: Not yet in PDB 3 copies of monomer prediction overlaid on crystal







T1061: Not yet in PDB 3 copies of monomer prediction overlaid on crystal





6VR4: Leiman, P.G., et al. Virion-packaged DNA-dependent RNA polymerase of crAss-like phage phi14:2 (CASP target). (To be published.)







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### **Manual interventions**

### We learned a lot during CASP14!



### Domains arising from H1O44 (RNA polymerase):

- Genetics search of full chain but folded in 4 parts
- Resulting pieces were used as templates to build the full chain
- Afterward, we fine-tuned our models to handle very long chains
- Can now obtain this accuracy in a fully-automated way

### > T1064 (ORF8)

- Five additional sequences were added to the MSA using NCBI Protein BLAST
- Tried more models to find a confident one

### T1024 (Multidrug transporter)

• Clustered templates into different classes to get diversity of opening angle

### Additional targets:

- Often the model diversity is low despite the error scores saying that there is error
- We would try to put older models in later positions to increase diversity



## What went badly

- Hanual work required to get a very high-quality Orf8 prediction
- Genetics search works much better on full sequences than individual domains
- Final relaxation required to remove stereochemical violations



### What went well

- Building the full pipeline as a single end-to-end deep learning system
- Building physical and geometric notions into the architecture instead of a search process
- Hodels that predict their own accuracy can be used for model-ranking
- Using model uncertainty as a signal to improve our methods (e.g. training new models to eliminate problems with long chains)



## Wrap up & future outlook

- We have built a system that confidently predicts accurate structures for most proteins and knows when it is wrong
- $\rightarrow$  As for CASP13<sup>1,2</sup>, we'll publish a peer-reviewed paper
- > We're also working on providing broad access to our work
- Demis Hassabis will be giving a keynote on Friday about Using AI to accelerate scientific discovery
- Lots of exciting work ahead for the field: Complexes, conformational change etc.
- Thanks again to the CASP organizers, experimentalists and everyone on whose work we're building

[1] Senior, A. W., et al. "Improved protein structure prediction using potentials from deep learning." Nature 577.7792 (2020): 706-710.
[2] Senior, A. W., et al. "Protein structure prediction using multiple deep neural networks in the 13th Critical Assessment of Protein Structure Prediction (CASP13)." Proteins 87.12 (2019): 1141-1148.









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