

# Joint CASP14-CAPRI50 assembly prediction round of 2020

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Structural Biology Group, VUB-VIB, Brussels, Belgium

Izmir Biomedicine and Genome Center, Izmir, Turkey

# CAPRI

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## CAPRI

Since 2002

Critical Assessment of  
PRedicted Interactions

Joint prediction rounds since 2014:

<b>25</b> Targets	Round 30
<b>10</b> Targets	Round 37
<b>21</b> Targets	Round 46
<b>12</b> Targets	Round 50

Prediction rounds on a “rolling” basis

Fits with publication schedule  
3 to 4 weeks per prediction round

## CASP

Since 1994

Structure PRedictions

CASP11	2014
CASP12	2016
CASP13	2018
CASP14	2020

Prediction season

# CAPRI

## CAPRI

Since 2002

Critical Assessment of  
PRedicted Interactions

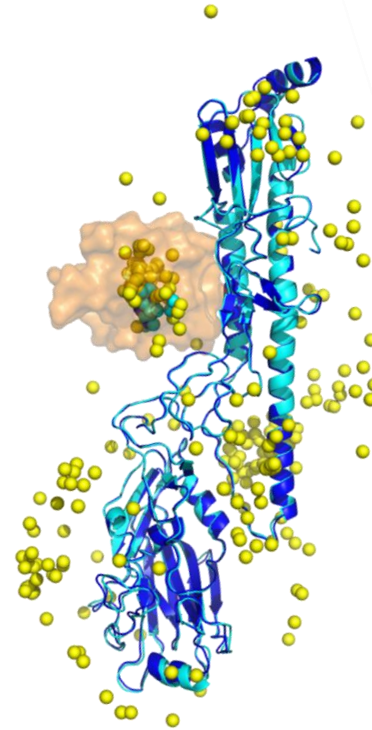
Dynamic experiment

Docking different molecules

Docking experiment

Scoring experiment

The targets are not the same



Proteins, Nucleic acids,  
Polysaccharides, Water,  
Peptides, Interfaces,  
Assemblies, SAXS,  
Binding affinities,  
Multi-domain  
organization

Marc



Shoshana



Guillaume



Sameer



Nurul



assessment, organization, website, operations, infrastructure

# CAPRI

Prioritized from  
**Gordon et al (2020)**

## CAPRI

Since 2002

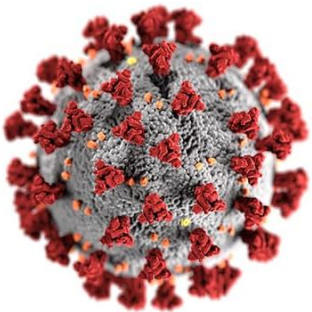
Critical Assessment of  
PRedicted Interactions

Dynamic experiment

Docking different molecules

Docking experiment

Scoring experiment



## COVID-19 Open Science Initiative

Round 51 Sep 15, 2020 – Jan 4, 2021

**Targets:** 3x SARS-CoV-2 with human host  
1x multi-component complex SARS-CoV-2 + RNA

All models (5000+) available for download:

<http://cb.iri.univ-lille1.fr/capri/covid.html>

3D-BioInfo community-wide initiative

February – July 2021

Benchmarking tools to discriminate between  
physiological/non-physiological protein-protein  
interfaces

Standard benchmark dataset (PDB) derived with  
QAlign and ProtCID – R. Dunbrack, E. Levy

Contact: [shoshana.wodak@gmail.com](mailto:shoshana.wodak@gmail.com)

# CAPRI

## CAPRI

Since 2002

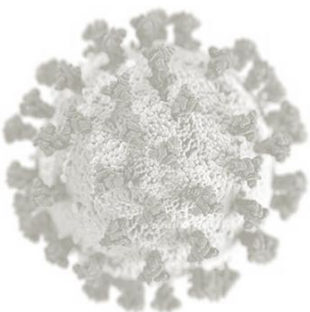
Critical Assessment of  
PRedicted Interactions

Dynamic experiment

Docking different molecules

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# Website

## CAPRI

Since 2002

**Critical Assessment of  
PRedicted Interactions**

<https://www.pdbe.org/capri>

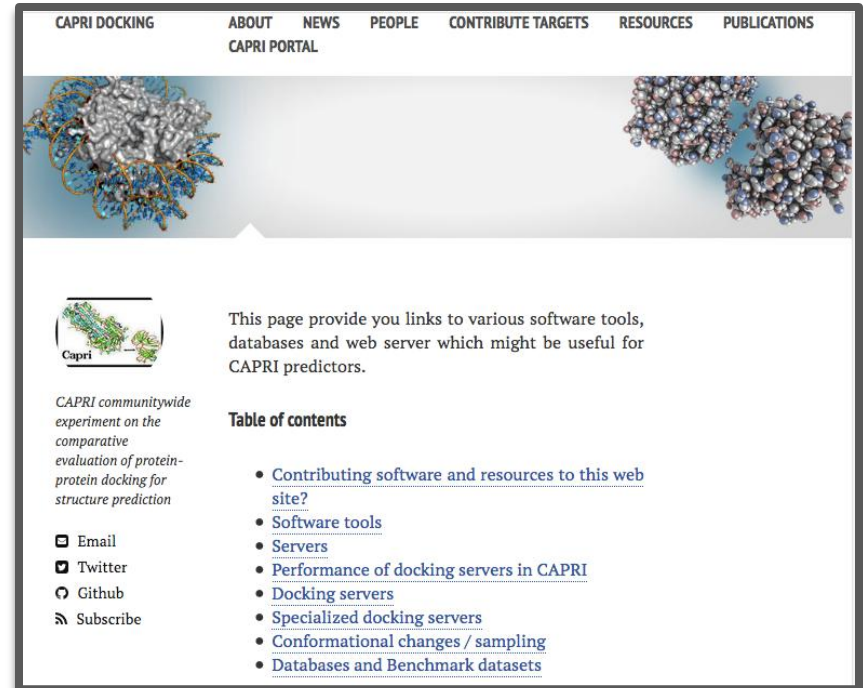
(for prediction submission)

<https://www.capri-docking.org/>

(community exchange portal)

 @CAPRIDOCK

<https://www.ebi.ac.uk/pdbe/complex-pred/capri/covid-capri/>



The screenshot shows the CAPRI website homepage. At the top, there is a navigation menu with links for CAPRI DOCKING, ABOUT, NEWS, PEOPLE, CONTRIBUTE TARGETS, RESOURCES, and PUBLICATIONS. Below the menu is a banner image featuring a 3D molecular model of a protein complex. The main content area includes a section titled 'Table of contents' with a list of links: 'Contributing software and resources to this web site?', 'Software tools', 'Servers', 'Performance of docking servers in CAPRI', 'Docking servers', 'Specialized docking servers', 'Conformational changes / sampling', and 'Databases and Benchmark datasets'. There is also a section for 'CAPRI communitywide experiment on the comparative evaluation of protein-protein docking for structure prediction' with social media links for Email, Twitter, Github, and Subscribe.

# Meetings

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## CAPRI

Since 2002

## Critical Assessment of PRedicted Interactions

Dynamic experiment

Docking different molecules

Docking experiment

Scoring experiment

Assessment meetings

La Londe-des-Maures	France	2002
Gaeta	Italy	2004
Toronto	Canada	2007
Barcelona	Spain	2009
Utrecht	The Netherlands	2013
Tel Aviv	Israel	2016
EBI Hinxton	UK	2019

# CAPRI evaluation meetings



To date: 50 rounds, 181 targets. 7 Evaluation meetings + CASP11/12/13/14  
7 Special Issues of Proteins dedicated to CAPRI, 2003 – 2020



# Management committee

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## CAPRI

Since 2002

Critical Assessment of  
PRedicted Interactions

Dynamic experiment

Docking different molecules

Docking experiment

Scoring experiment

Assessment meetings

Management committee

Alexandre Bonvin The Netherlands

Marc Lensink France

Michael Sternberg UK

Sandor Vajda USA

Ilya Vakser USA

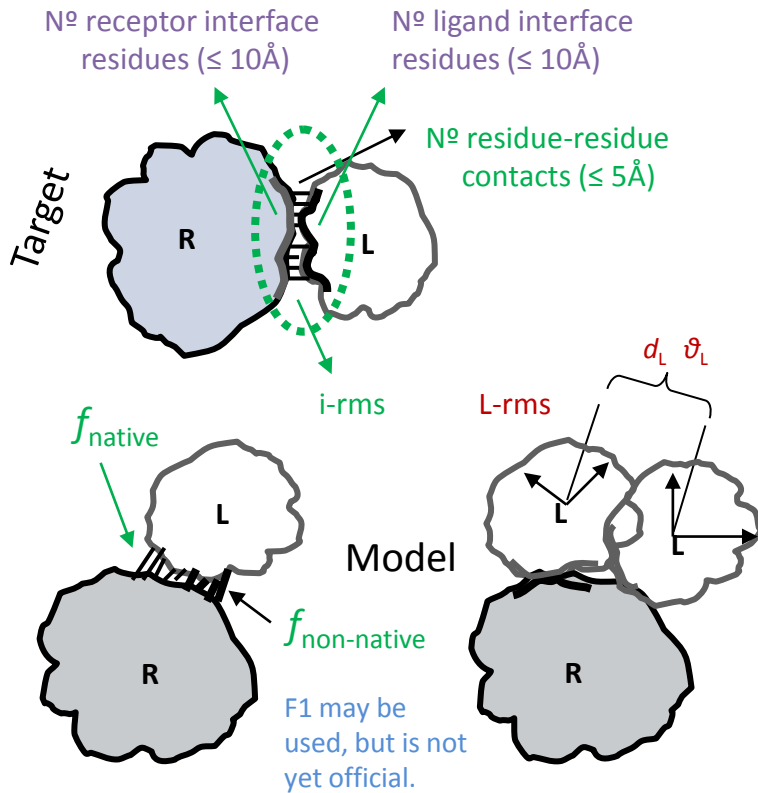
Sameer Velankar UK

Shoshana Wodak Belgium

Joel Janin France

# CAPRI Assessment

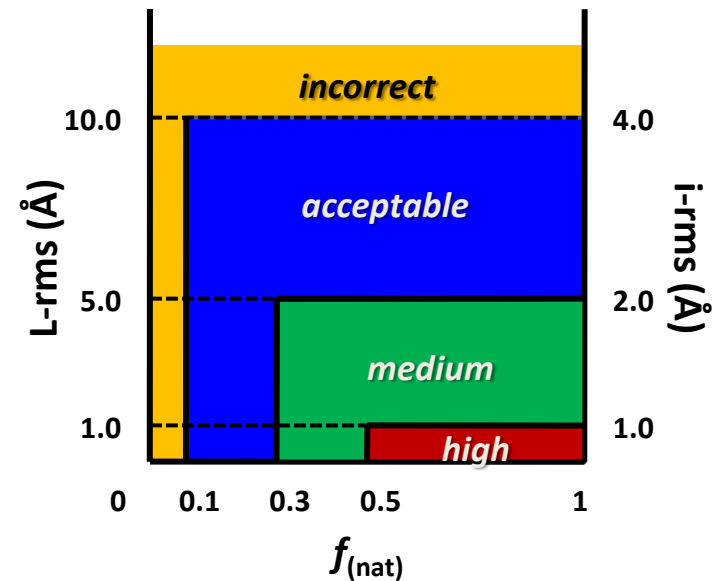
- Assessment criteria established as community consensus



$f_{\text{native}}$	residue-residue	5 Å
<b>i-rms</b>	Interface backbone	10 Å
<b>S-rms</b>	Interface side-chain	10 Å
<b>L-rms</b>	ligand backbone	
$n_{\text{clashes}}$	atom-atom	3 Å
$d_L$		
$\vartheta_L$		

Only **L-rms**, **i-rms** and  $f_{\text{nat}}$  are used to classify protein-protein interaction models in CAPRI.

Additional quantities are being calculated, such as **S-rms**, which are useful quality measures for protein-peptide interaction models.



- Evaluation of 5 submitted models
- Focusing on individual interfaces of interaction

# CAPRI Assessment

1. CAPRI assessment is (a) receptor/ligand and (b) interface based
  - fraction of native contacts  $f(\text{nat})$
  - i-rms
  - L-rms
2. Four categories
  - incorrect, acceptable, medium, high
3. For multimeric targets, each interface is assessed separately
4. Final target score (set of interfaces) is a combination of individual interface scores
  - Either an AverageOf or BestOf
5. Final predictor score is the sum of these scores

$$\text{Score} = \omega_1 \cdot N_{\text{ACC}} + \omega_2 \cdot N_{\text{MED}} + \omega_3 \cdot N_{\text{HIGH}}$$

$$\omega_1 = 1; \omega_2 = 2; \omega_3 = 3$$

# Targets & Participation

CAPRI predictor groups that submitted more than 10 models



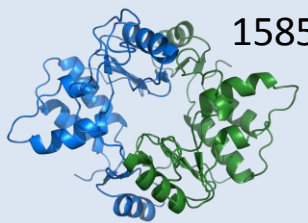
CAPRI ID	CASP ID	Stoichiometry	Difficulty	# interfaces	Predictors	Uploaders (>10)	CASP
<b>T164</b>	T1032	A2	Easy	1	28	18	29
<b>T165</b>	H1036	A:HL	Easy	1	27	12	32
<b>T166</b>	H1045	AB	Easy	1	24	16	36
T167	T1050			Cancelled	25	15	29
<b>T168</b>	T1052	A3	Easy	1	24	11	29
<b>T169</b>	T1054	A2	Difficult	1	27	16	32
<b>T170</b>	H1060	A6B12C3D6	Difficult	9	23	12	22
T171	T1063			No structure	26	13	31
T172	H1066			No structure	27	17	37
T173	H1069			No structure	27	17	37
<b>T174</b>	T1070	A3	Difficult	1	24	13	31
T175	T1073	A4/A2		Cancelled	25	13	31
<b>T176</b>	T1078	A2	Difficult	1	27	16	33
<b>T177</b>	H1081	A10:A10	Easy	3	24	12	27
<b>T178</b>	T1083	A2	Difficult	1	26	13	33
<b>T179</b>	T1087	A2	Difficult	1	25	13	32
<b>T180</b>	T1099	A2:A2	Easy	2	25	11	30
T181	H1103			No structure	27	15	34

Easy targets

Difficult targets

12 Targets

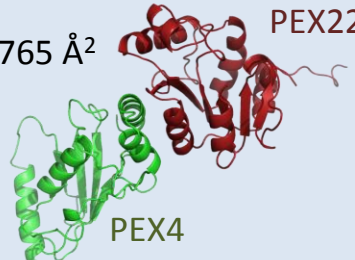
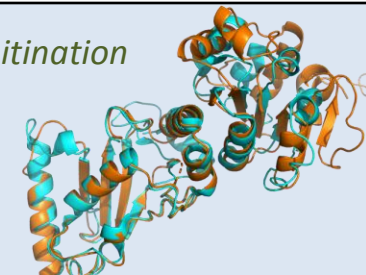
# The targets: 2 easy dimers 1/2

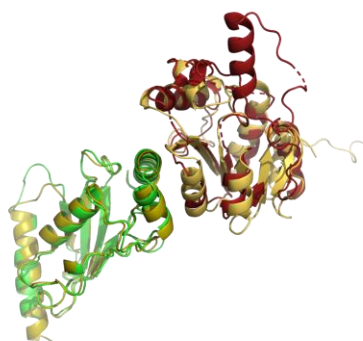
T164	 <p>1585 Å<sup>2</sup> but 2 contact regions</p>	<i>Structural maintenance of chromosomes (SMC) Flexible hinge domain-containing protein 1</i>				1gwk	acceptable
T1032						f(nat)	0.5575
6n64						L-rms	5.84 Å
		<b>Template</b>	<b>RMSD</b>	<b>LGA</b>	<b>Seq.ID</b>		
		1gwk (PPI3D)	2.8 Å (complex)	50.5	30%	i-rms	3.84 Å

**EASY**

Quality	Group
**	Gray, Seok, <b>MDOCKPP</b>
*	Kihara, Chang, Bates, Venclovas, Zou, Huang, Shen, Pierce, Kozakov/Vajda, Nakamura
	<b>SWARMDOCK, HDOCK, HAWKDOCK, CLUSPRO, GALAXYPPDOCK, LZERD</b>
	<i>Baker, CoDock, DATE, Elofsson, Risoluto, htjcadd, Takeda-Shitaka, Seok-naïve-assembly</i>

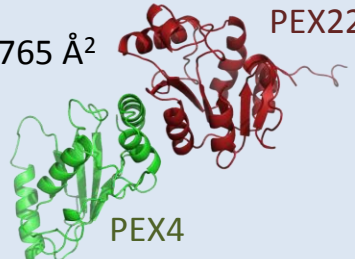
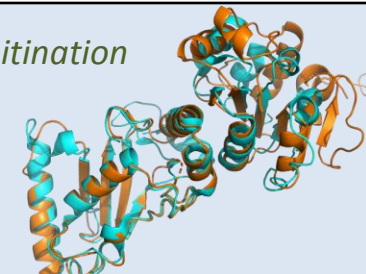
# The targets: 2 easy dimers 2/2

T166	765 Å <sup>2</sup>		<p>PEX4 – protein peroxin-4 – may be involved in ubiquitination</p> <p>PEX22 – peroxisome biogenesis protein</p>	
H1045			heterodimer	
n/a				<p>“[...] ubiquitin conjugating enzyme Pex4p in complex with [...] Pex22p”</p>



	Individual templates		Template for the complex		
	<i>Ubiquitin-conjugating enzyme</i>	<i>Deoxyribonucleotidase</i>			
Template	3ptf	3bvv	5nkz	PEX-4	PEX22
RMSD	0.5 Å	1.6 Å	RMSD	0.6 Å	2.6 Å
LGA	94.8	56.0	LGA	94.4	N/A*
Seq.ID	39%	21%	Seq.ID	39%	37%
* not listed in the first 25 hits					

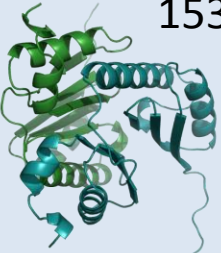
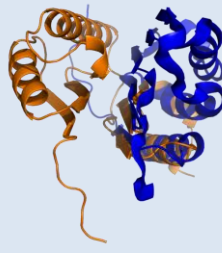
# The targets: 2 easy dimers 2/2

T166	765 Å <sup>2</sup>		<p><i>PEX4 – protein peroxin-4 – may be involved in ubiquitination</i></p> <p><i>PEX22 – peroxisome biogenesis protein</i></p>	
H1045			heterodimer	<p><i>“[...] ubiquitin conjugating enzyme Pex4p in complex with [...] Pex22p”</i></p>
n/a				

**EASY**

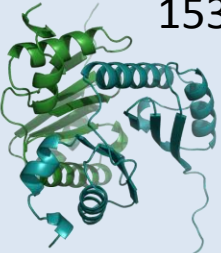

Quality	Group
***	Venclovas, Takeda-Shitaka
**	Huang, Vakser, Kozakov/Vajda, Chang, Shen, Fernandez-Recio, Bates, Pierce, Zou, Kihara, Seok, Nakamura
	<i>AILON, Lamoureux, CoDock, DATE</i>
*	<b>MDOCKPP, SWARMDOCK, GALAXYPPDOCK, HDOCK, DellaCorte</b>

# The targets: 4 difficult dimers 1/4


T169		1530 Å <sup>2</sup> <i>Uncharacterized protein from Acinetobacter baumannii</i>		<b>Template</b>	<b>2kgs</b>
T1054				RMSD (half)	2.8 Å
n/a				RMSD (whole)	8.8 Å
		No templates for the assembly		LGA	44.7



# The targets: 4 difficult dimers 1/4

T169		1530 Å <sup>2</sup> <i>Uncharacterized protein from Acinetobacter baumannii</i>		<b>Template</b>	<b>2kgs</b>
T1054				RMSD (half)	2.8 Å
n/a				RMSD (whole)	8.8 Å
		No templates for the assembly No acceptable submissions		LGA	44.7

# The targets: 4 difficult dimers 2/4

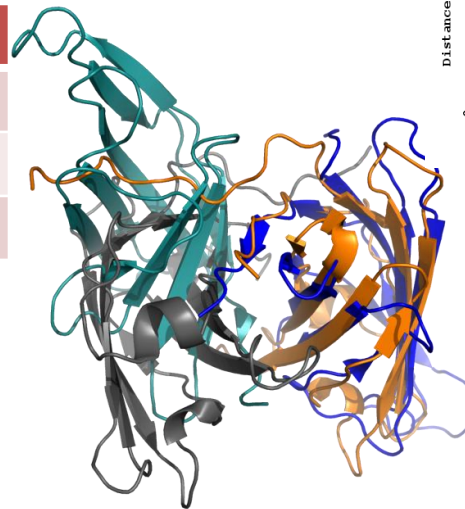
T176	 <p>1480 Å<sup>2</sup></p>	<p><i>Tsp1 protein from Trichoderma virens, Cysteine rich protein</i>  <i>Plays an important role in fungi-host interactions</i></p>
T1078		
n/a		

## DIFFICULT

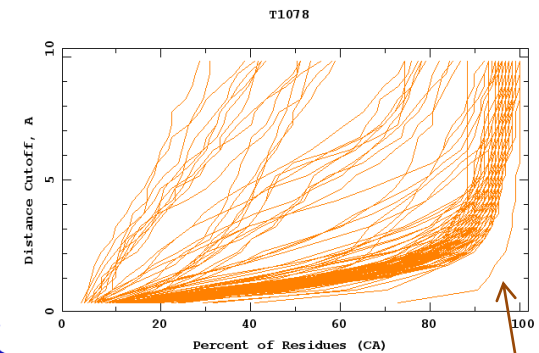
Major allergen from *Alternaria alternata*  
 Hypersensitive response-inducing protein from *Verticillium dahliae*

Template	3v0r	4aud	5xmz
RMSD	3.8 Å	4.5 Å	6.9 Å
LGA	53.9	52.2	52.6
Seq.ID	11.0%	16.8%	No alignment

**4aud** onto target  
 Single-chain: 4.5 Å  
 Double-chain: 10.5 Å



CASP14



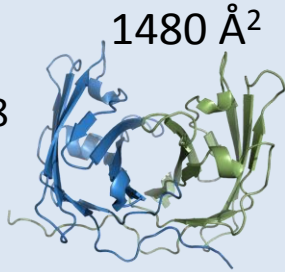
AlphaFold2

# The targets: 4 difficult dimers 2/4

T176 1480 Å<sup>2</sup>

T1078

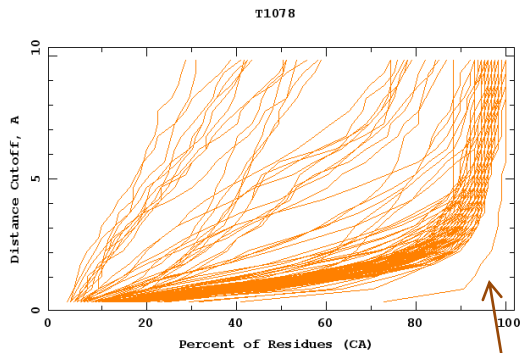
n/a



*Tsp1 protein from Trichoderma virens, Cysteine rich protein  
Plays an important role in fungi-host interactions*

## DIFFICULT

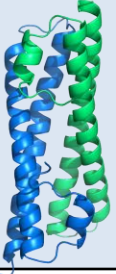
Quality	Group
*	MDOCKPP, Seok, Zou, Elofsson



AlphaFold2

# The targets: 4 difficult dimers 3&4

T178  
T1083  
n/a



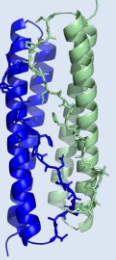
*Protein segment from Nitrosococcus oceani*

1650 Å<sup>2</sup>

*Alpha-pore forming toxin*

Template	6h2f	6grj	6r1j
RMSD	5.7 Å	4.7 Å	5.2 Å
Seq.ID	7%	7%	7%

T179  
T1087  
n/a

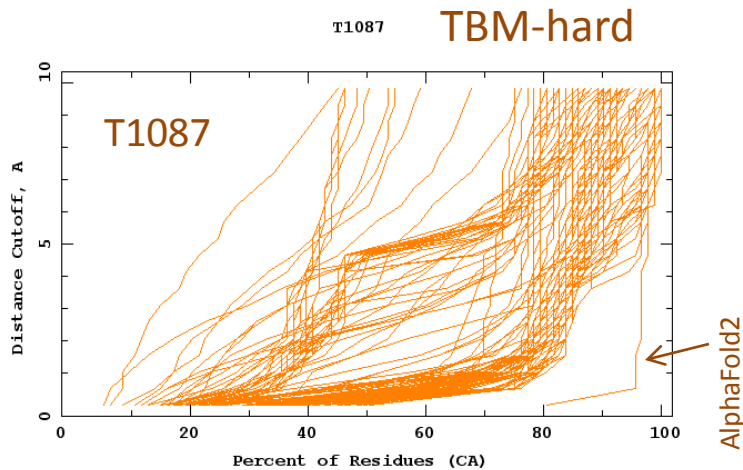


*Protein segment from Methylobacter tundripaludum*

1620 Å<sup>2</sup>

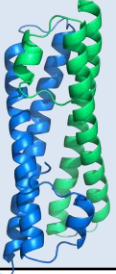
*Bacillus thuringiensis Cry6aa2 prototoxin*

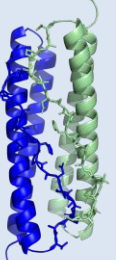
Template	5kuc	5ghe
RMSD	5.4 Å	5.4 Å
Seq.ID	7.1%	7.4%



**DIFFICULT**

# The targets: 4 difficult dimers 3&4

T178 T1083 n/a		<i>Protein segment from Nitrosococcus oceanii</i>  1650 Å <sup>2</sup>	<i>Alpha-pore forming toxin</i>	<table border="1"> <thead> <tr> <th>Template</th> <th>6h2f</th> <th>6grj</th> <th>6r1j</th> </tr> </thead> <tbody> <tr> <td>RMSD</td> <td>5.7 Å</td> <td>4.7 Å</td> <td>5.2 Å</td> </tr> <tr> <td>Seq.ID</td> <td>7%</td> <td>7%</td> <td>7%</td> </tr> </tbody> </table>	Template	6h2f	6grj	6r1j	RMSD	5.7 Å	4.7 Å	5.2 Å	Seq.ID	7%	7%	7%
Template	6h2f	6grj	6r1j													
RMSD	5.7 Å	4.7 Å	5.2 Å													
Seq.ID	7%	7%	7%													

T179 T1087 n/a		<i>Protein segment from Methylobacter tundripaludum</i>  1620 Å <sup>2</sup>	<i>Bacillus thuringiensis Cry6aa2 prototoxin</i>	<table border="1"> <thead> <tr> <th>Template</th> <th>5kuc</th> <th>5ghe</th> </tr> </thead> <tbody> <tr> <td>RMSD</td> <td>5.4 Å</td> <td>5.4 Å</td> </tr> <tr> <td>Seq.ID</td> <td>7.1%</td> <td>7.4%</td> </tr> </tbody> </table>	Template	5kuc	5ghe	RMSD	5.4 Å	5.4 Å	Seq.ID	7.1%	7.4%
Template	5kuc	5ghe											
RMSD	5.4 Å	5.4 Å											
Seq.ID	7.1%	7.4%											

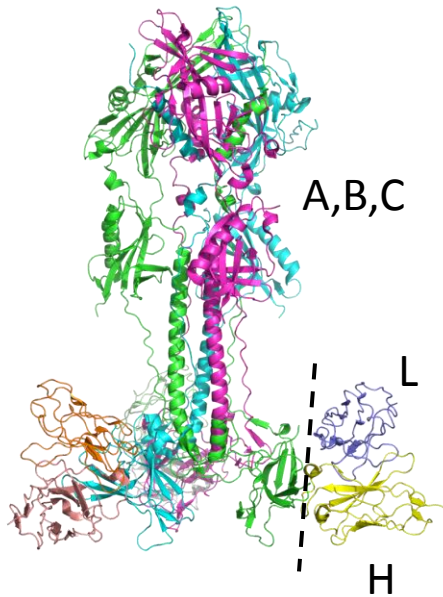
Quality	Group T178	Group T179
**	Venclovas	
*	Chang, Lubecka, Pierce, Seok, Kihara, Bates, Fernandez-Recio, Zou, Liwo	Zou, Venclovas, Liwo, Chang, Shen, Seok, Vakser, Pierce
	<b>LZERD, MDOCKPP, HAWKDOCK</b>	<b>LZERD, MDOCKPP</b>
	<i>Baker, CoDock, UNRES, McGuffin, htjcadd, UNRES-contact</i>	<i>Baker, CoDock, UNRES</i>

# The targets: 3 trimers

*Herpesvirus envelope glycoprotein*  
*Human monoclonal antibody*

*Tail fibre of the Salmonella*  
*virus epsilon15*

*Phage tail protein,*  
*attachment region*



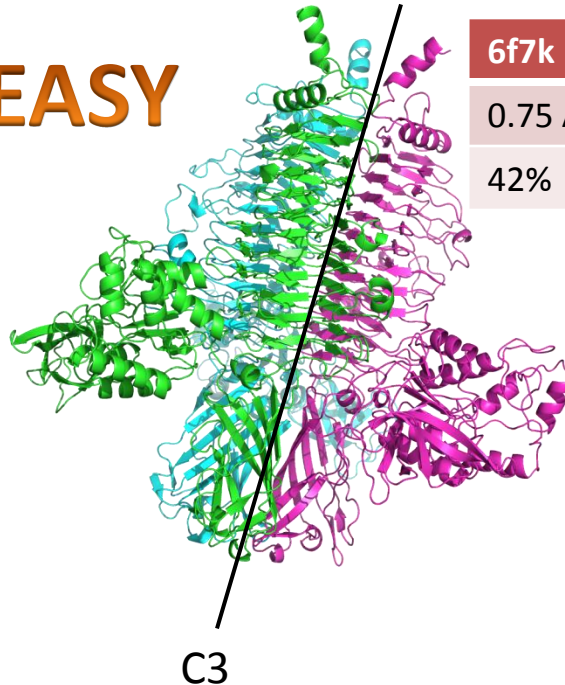
T165
H1036
6vn1

T168
T1052
N/A

T174
T1070
N/A

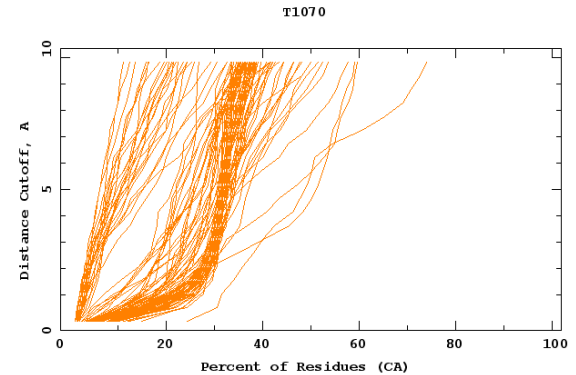
**EASY**

6f7k
0.75 Å
42%



**DIFFICULT**

6esc	4oaw
1.0 Å	1.2 Å
60%	75% (H)
	86% (L)



# The targets: 3 trimers

*Herpesvirus envelope glycoprotein*  
*Human monoclonal antibody*

*Tail fibre of the Salmonella*  
*virus epsilon15*

*Phage tail protein,*  
*attachment region*

T165  
H1036  
6vn1

T168  
T1052  
N/A

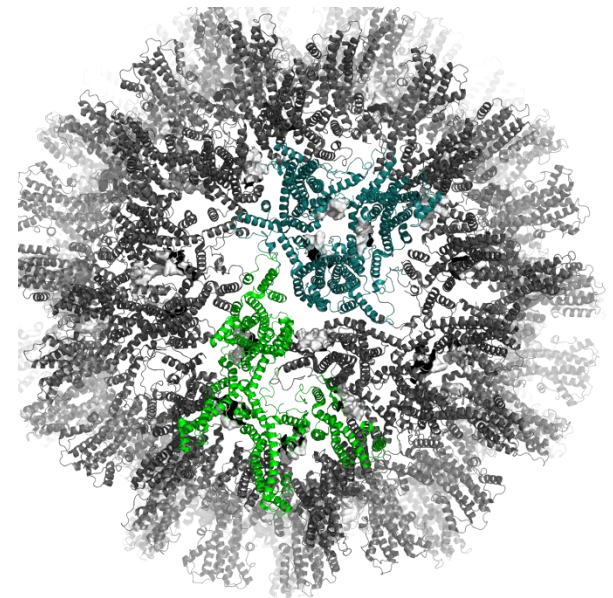
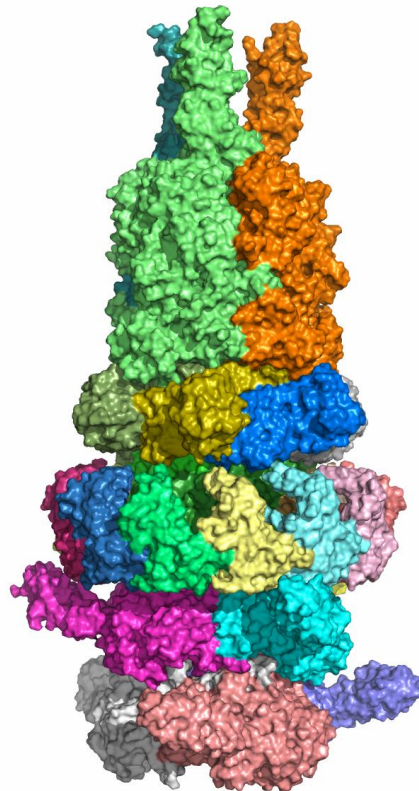
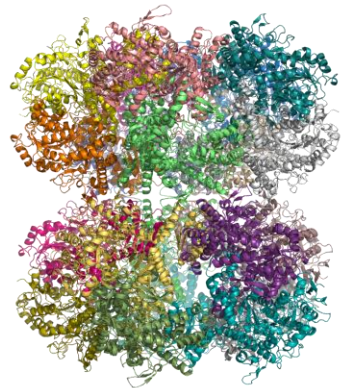
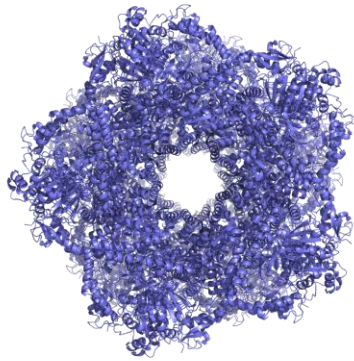
T174  
T1070  
N/A

No acceptable  
solutions for  
T165 and T174



Quality	Group T168
***	Zou, <b>GALAXYPPDOCK</b>
**	Kihara, Vakser, Fernandez-Recio, Chang, Pierce, Huang, Venclovas
	<b>LZERD</b>
*	<i>Baker, CoDock, Lamoureux, UNRES-template, Takeda-Shitaka</i>
	<i>Seok, Kozakov/Vajda, Czaplewski, UNRES</i>

# The remaining 3 targets





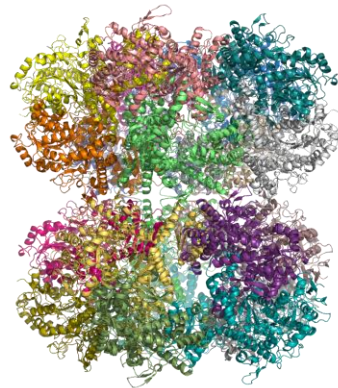
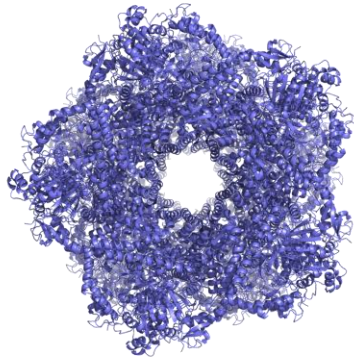
# The decarboxylase

T170

H1060

N/A

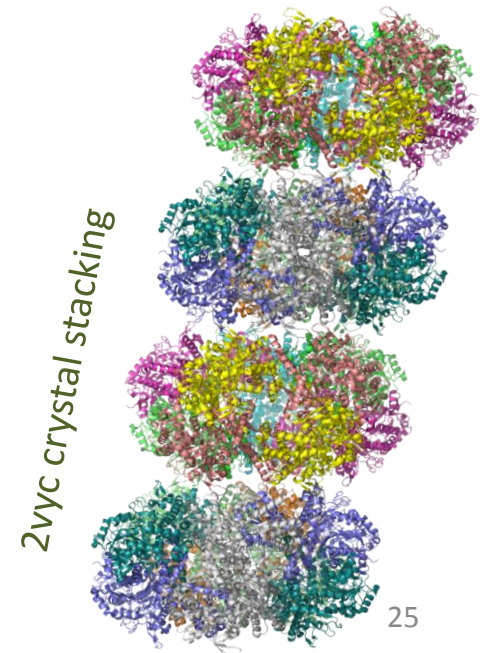
*Arginine decarboxylase involved in bacterial stress response; cryo-EM  
Stoichiometry: 2 stacked decamers, each adopting D5 symmetry. The  
main challenge for this target is to predict the stacking interface*



**EASY**

Template	Stoich.		RMSD	Seq.ID
2vyc	10		0.46 Å	71%
5fkz	1		1.55 Å	35%
3n75	5	10*	0.97 Å	33%
6q6i	1		1.36 Å	39%
1c4k	1	2*	1.38 Å	29%

*\*applying symmetry operations*

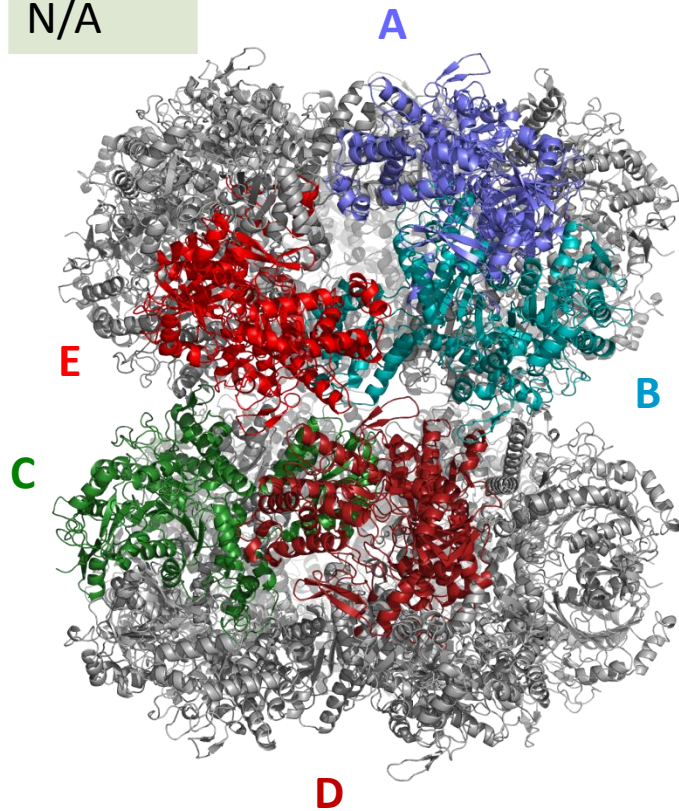


# The decarboxylase

T170

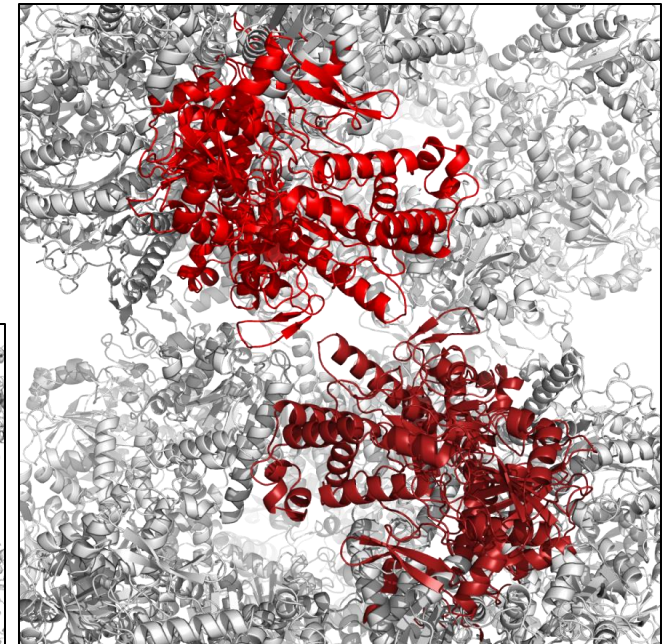
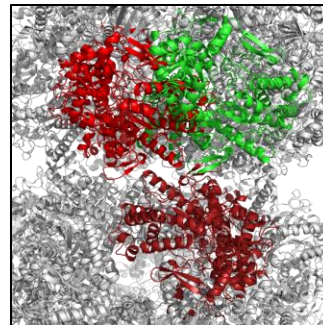
H1060

N/A



Interface		Area	Location	Occurrence
1	A:B	5000 Å <sup>2</sup>	Intra	10x
2	C:D	1250 Å <sup>2</sup>	Intra	20x
3	D:E	300 Å <sup>2</sup>	<b>Inter</b>	5x
		180 Å <sup>2</sup>	Intra	10x

*Green monomer is  
crystal packing  
from template 2vyc*

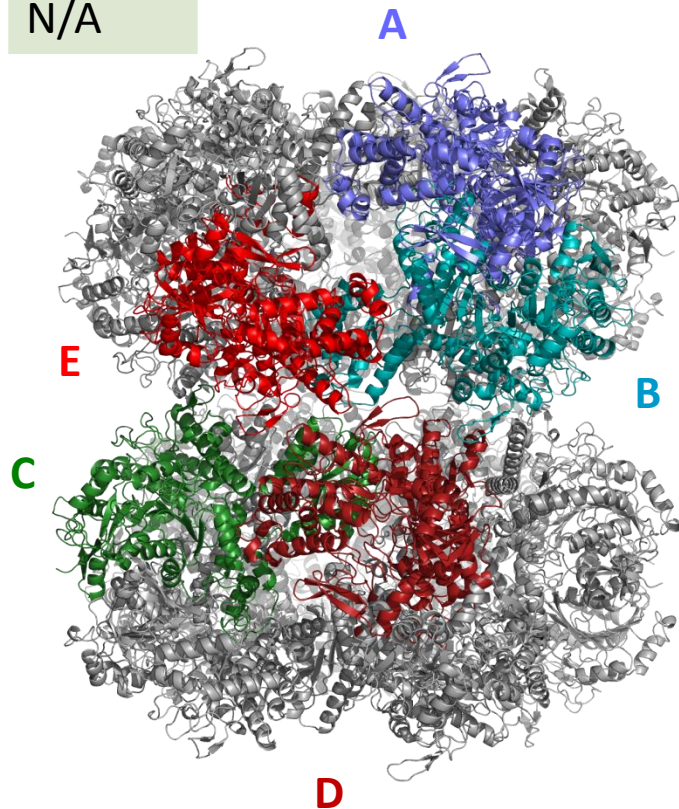


# The decarboxylase

T170

H1060

N/A



Interface		Area	Location	Occurrence
1	A:B	5000 Å <sup>2</sup>	Intra	10x
2	C:D	1250 Å <sup>2</sup>	Intra	20x
3	D:E	300 Å <sup>2</sup>	Inter	5x

\*\*\* predictions for interfaces 1 and 2, including the **top-1** of server predictions

Quality	Group T177 Interface 3
***	<b>MDOCKPP</b>
**	Venclovas, Zou, Grudin, Kozakov/Vajda, Pierce, CLUSPRO, VoroCNN-select, Ornate-select
*	Bates, Seok, Del Carpio, Huang, Nakamura
	<b>SWARMDOCK, HDOCK</b>

T180

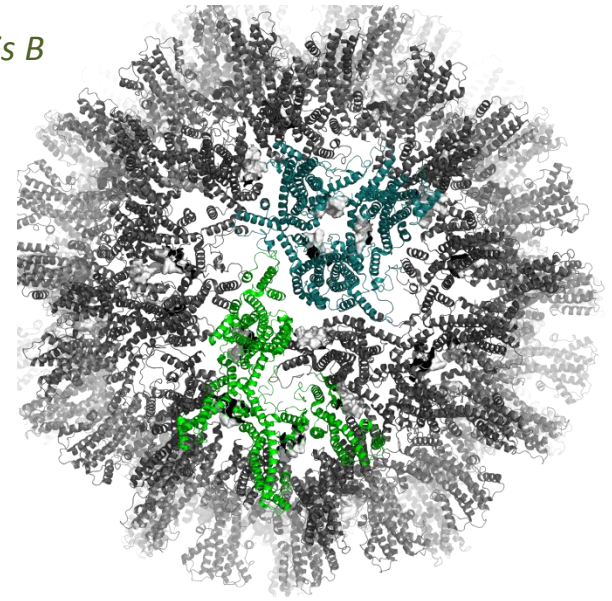
T1099

6ygh

# The viral capsid

*The target represents the cryo-EM structure of the capsid of the duck hepatitis B virus, which features T=4 icosahedral symmetry with a total of 240 subunits.*

*We invite predictors to submit models containing the minimum number of subunits necessary to define the unique interfaces.*



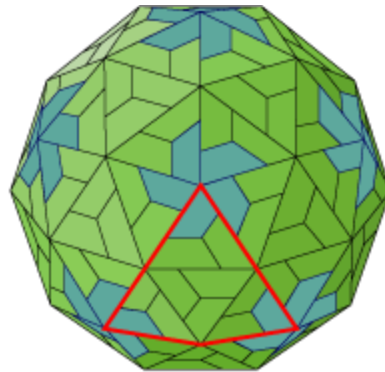
viralzone.expasy.org



x60



x20



T=4

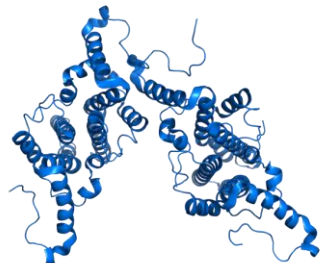
T=4 icosahedral  
asymmetric unit

Triangular facet

*The capsid is composed of 60 asymmetric units made of 4 proteins, for a total of 240 capsid proteins.*



Target



Hepatitis B virus core antigen  
Cryo-EM 3.5 Å  
19.5 % sequence identity

CASP14

**EASY**

# The viral capsid

*The target represents the cryo-EM structure of the capsid of the duck hepatitis B virus, which features  $T=4$  icosahedral symmetry with a total of 240 subunits.*

*We invite predictors to submit models containing the minimum number of subunits necessary to define the unique interfaces.*

#chains	2	3	<b>4</b>	6	7	8	9	12	16	20	52
#models	7	4	<b>42</b>	25	15	5	1	5	5	11	5

*The capsid is composed 60 asymmetric **units made of 4 proteins**, for a total of 240 capsid proteins.*



# The viral capsid

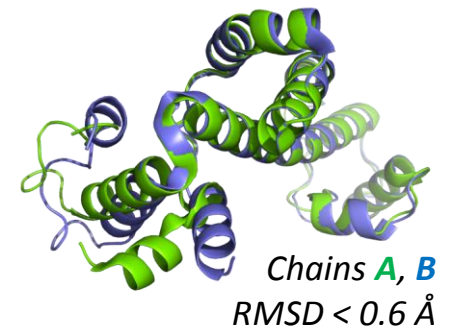
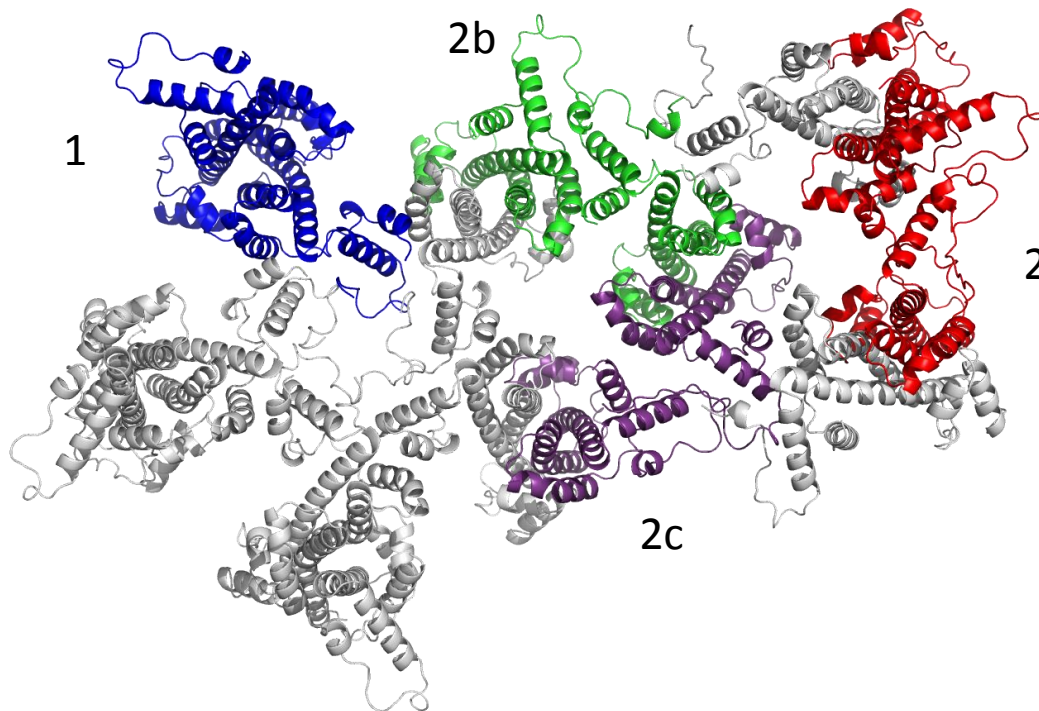
T180

T1099

6ygh



Interface	Area	Chains
1	1970 Å <sup>2</sup>	A:B
2	1110 Å <sup>2</sup>	B:C
2b		D:A'
2c		A:D'



*Pseudo-symmetry for interfaces 2/2b/2c*

(Å)	A	B	C	D
A	0	0.44	0.55	0.51
B		0	0.63	0.62
C			0	0.74
D				0

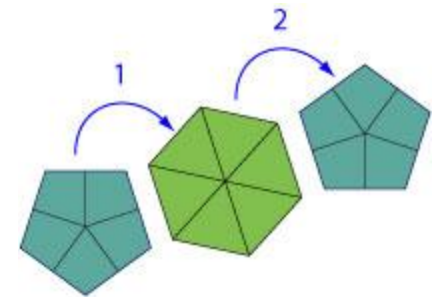
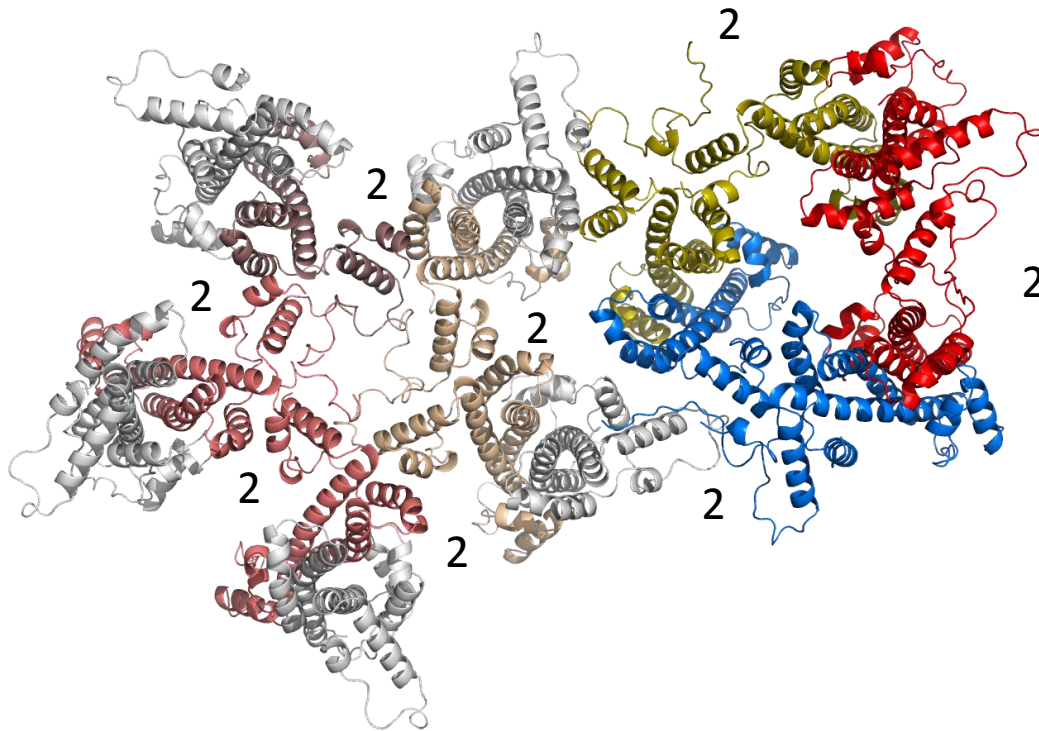
# The viral capsid

T180

T1099

6ygh

Interface	Area	Chains
1	1970 Å <sup>2</sup>	A:B
2	1110 Å <sup>2</sup>	B:C
2b		D:A'
2c		A:D'



$$h=2, k=0$$

$$T=(2)^2 + (2)(0) + (0)^2=4$$

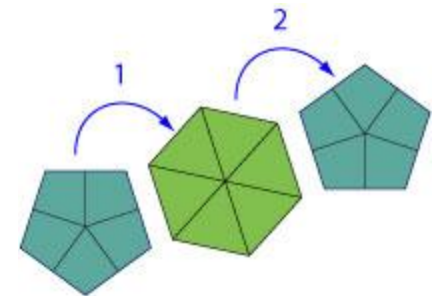
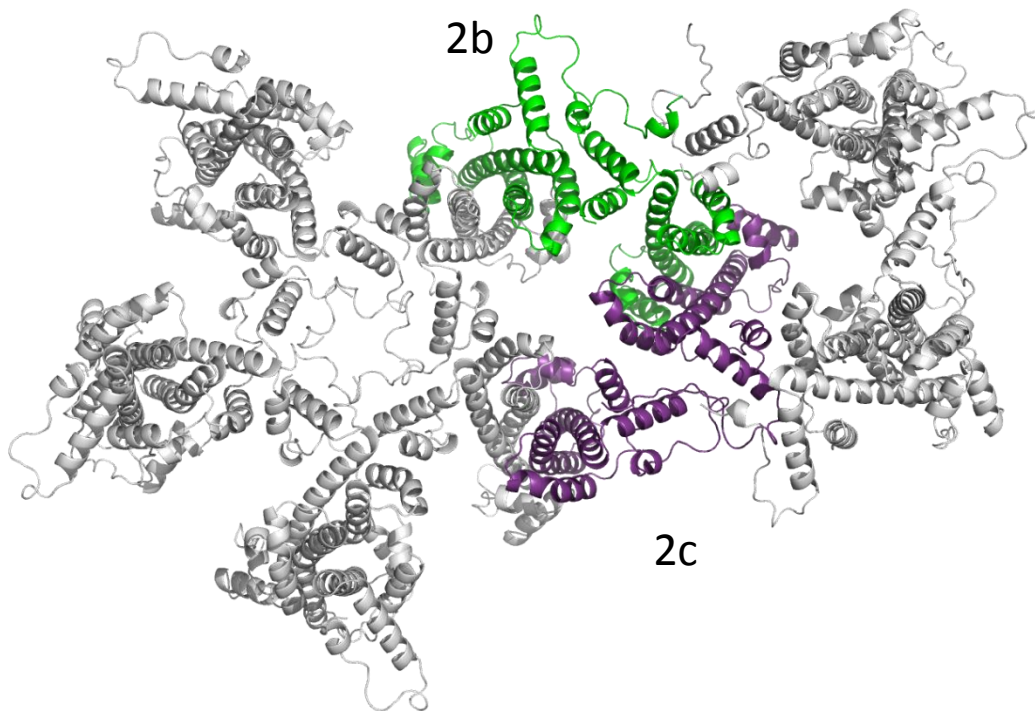
# The viral capsid

T180

T1099

6ygh

Interface	Area	Chains
1	1970 Å <sup>2</sup>	A:B
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2b		D:A'
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$$h=2, k=0$$

$$T=(2)^2 + (2)(0) + (0)^2=4$$

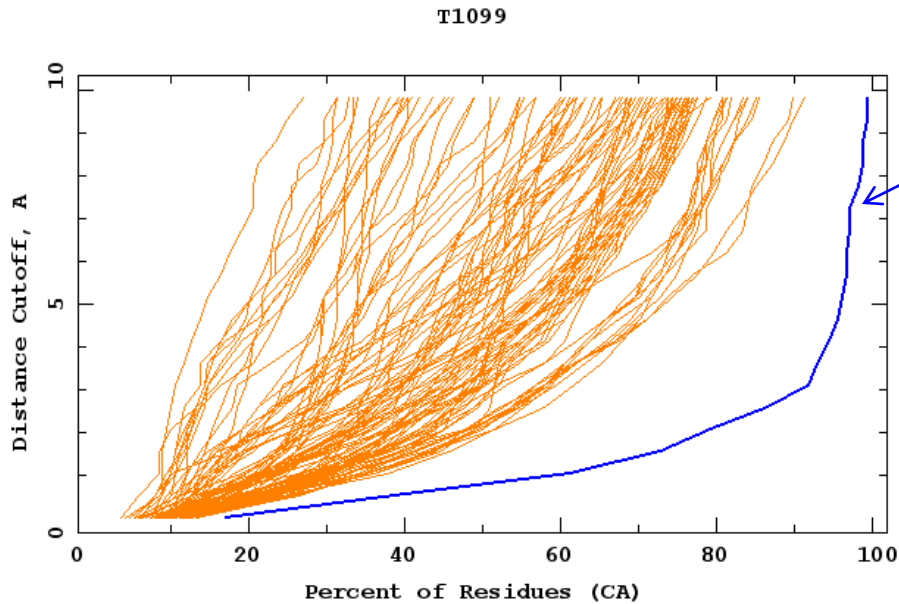


# The viral capsid

T180

T1099

6ygh

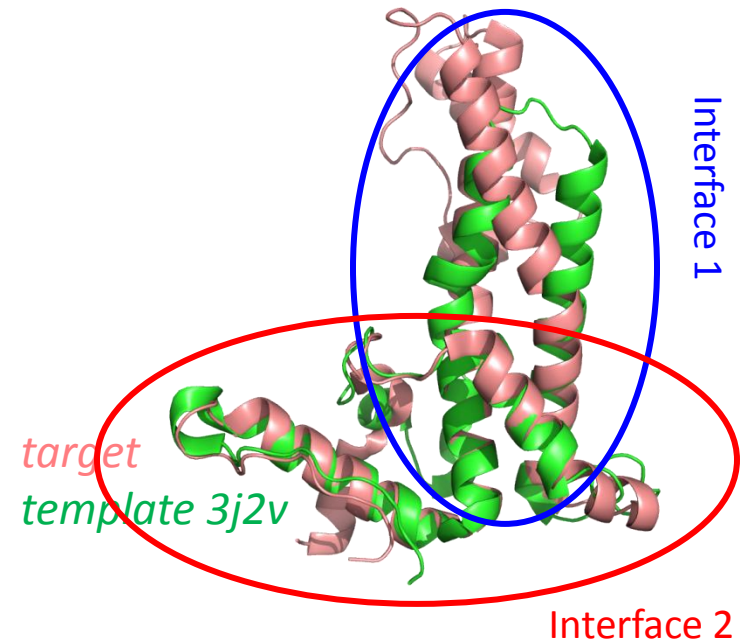


AlphaFold2

*Did not participate in assembly prediction*

Template	Seq.ID	RMSD	LGA
3j2v	19.5%	2.0 Å	48.6
3kxs	20.1%	3.8 Å	43.6
5t2p	19.5%	4.4 Å	48.2
6ecs	18.1%	5.8 Å	

*Identified by hhpred*

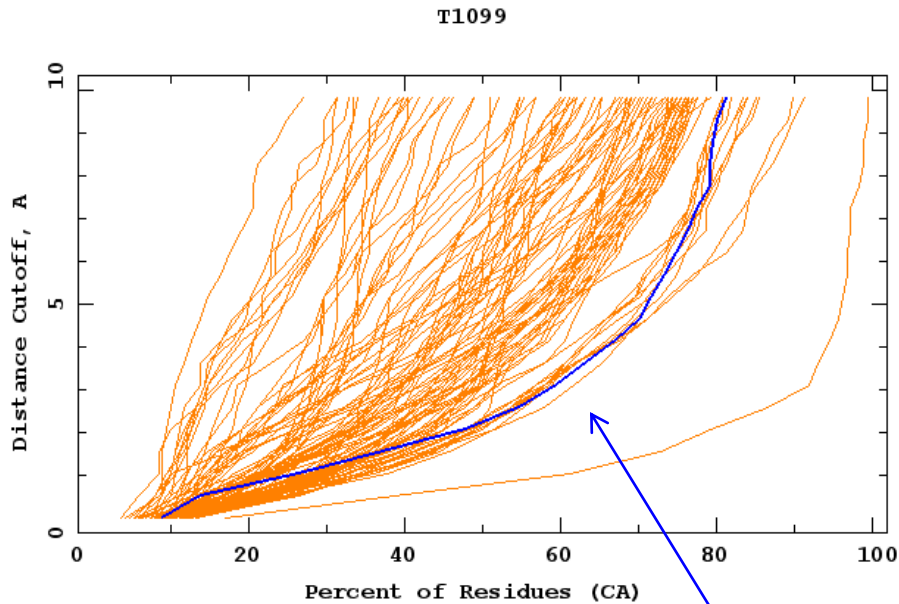


# The viral capsid

T180

T1099

6ygh



Interface	Area	Chains
1	1970 Å <sup>2</sup>	A:B
2	1110 Å <sup>2</sup>	B:C

12 groups have \*\* in top-1 for interface 2,  
Including **CLUSPRO**, **LZERRD**, **GALAXYPPDOCK**

Venclovas has \*\*\* in top-5.

**Only Seok got \* for interface 1 (model 2)**

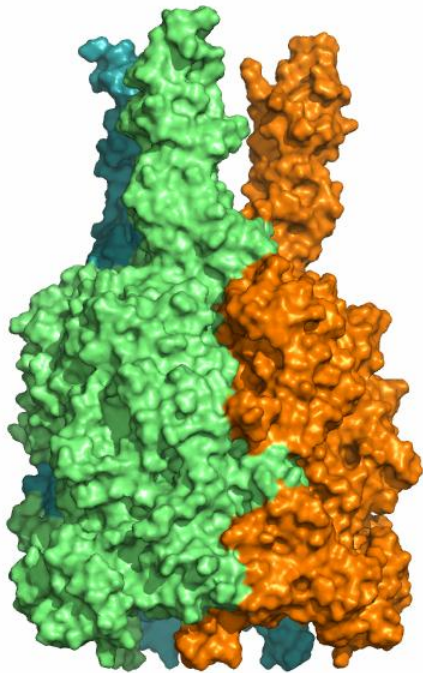


T170

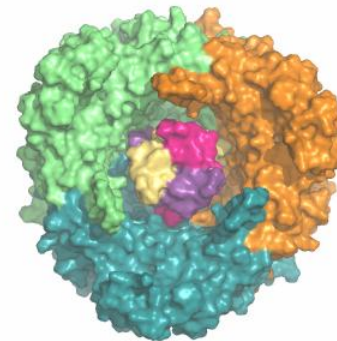
H1060

N/A

# The phage tail



E3

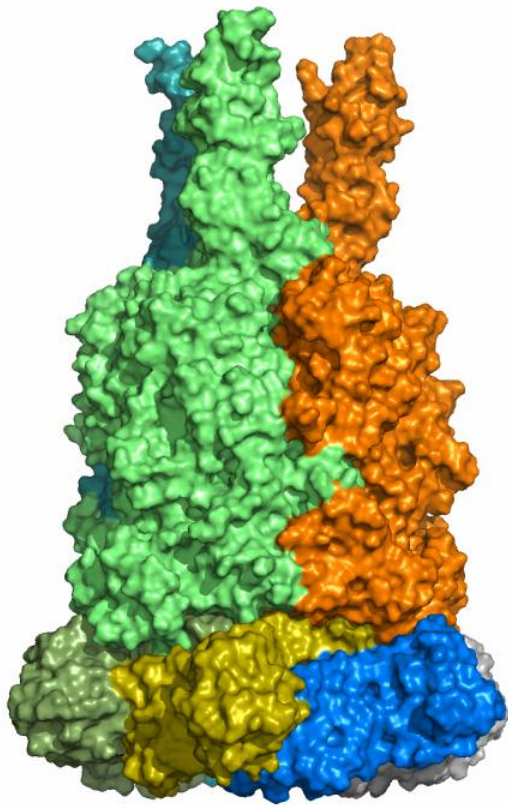


T170

H1060

N/A

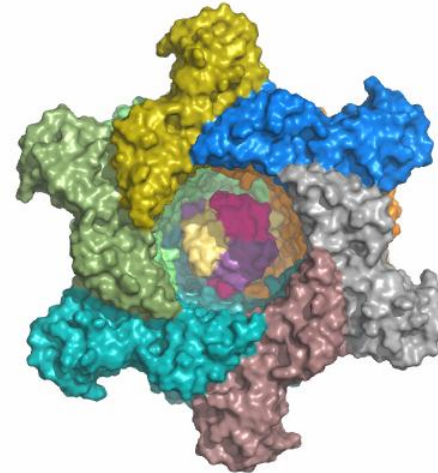
# The phage tail



E3

D6

12000 Å<sup>2</sup>

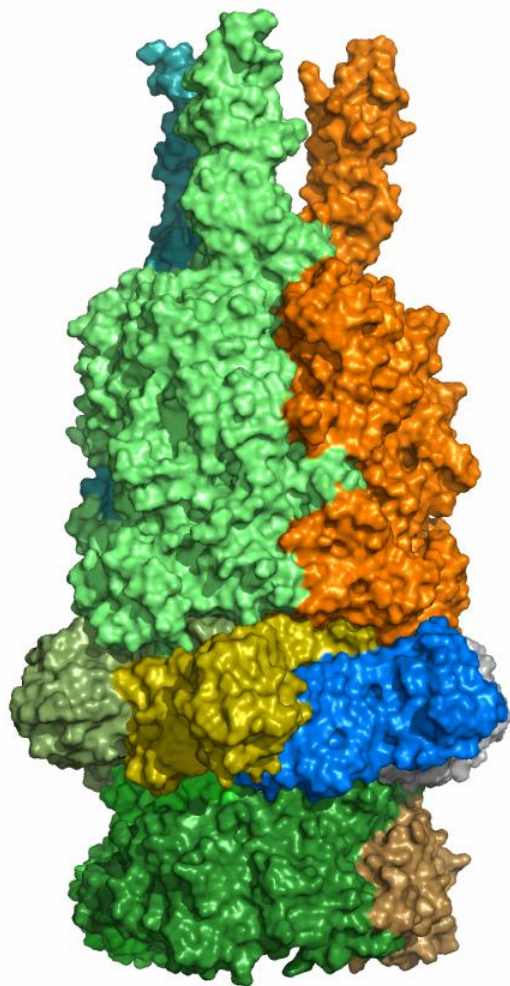


# The phage tail

T170

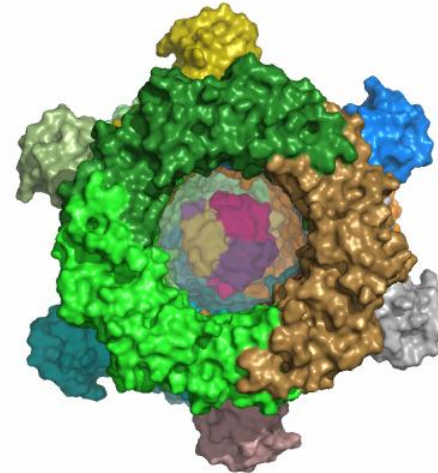
H1060

N/A



12000 Å<sup>2</sup>

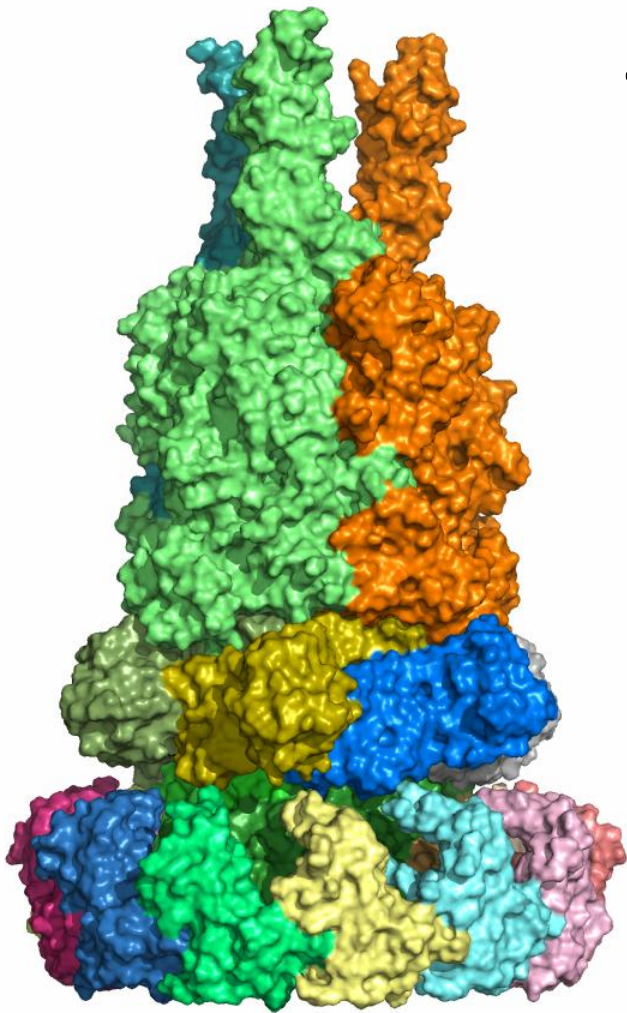
9800 Å<sup>2</sup>



T170
H1060
N/A

# The phage tail

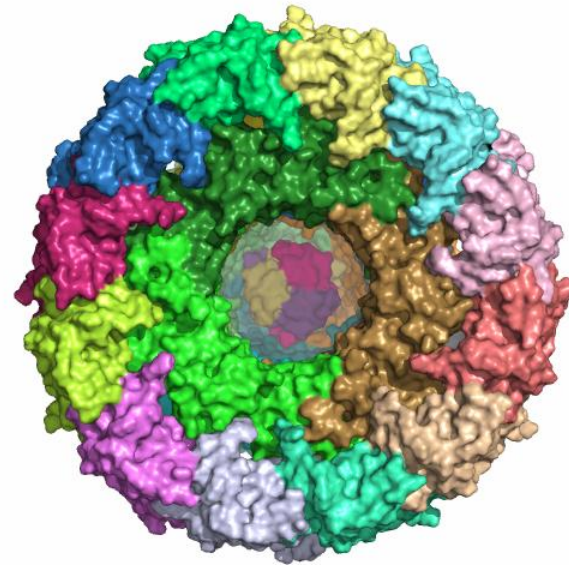
E3  
D6  
C12  
B3



12000 Å<sup>2</sup>

9800 Å<sup>2</sup>

15000 Å<sup>2</sup>



# The phage tail

T170

H1060

N/A

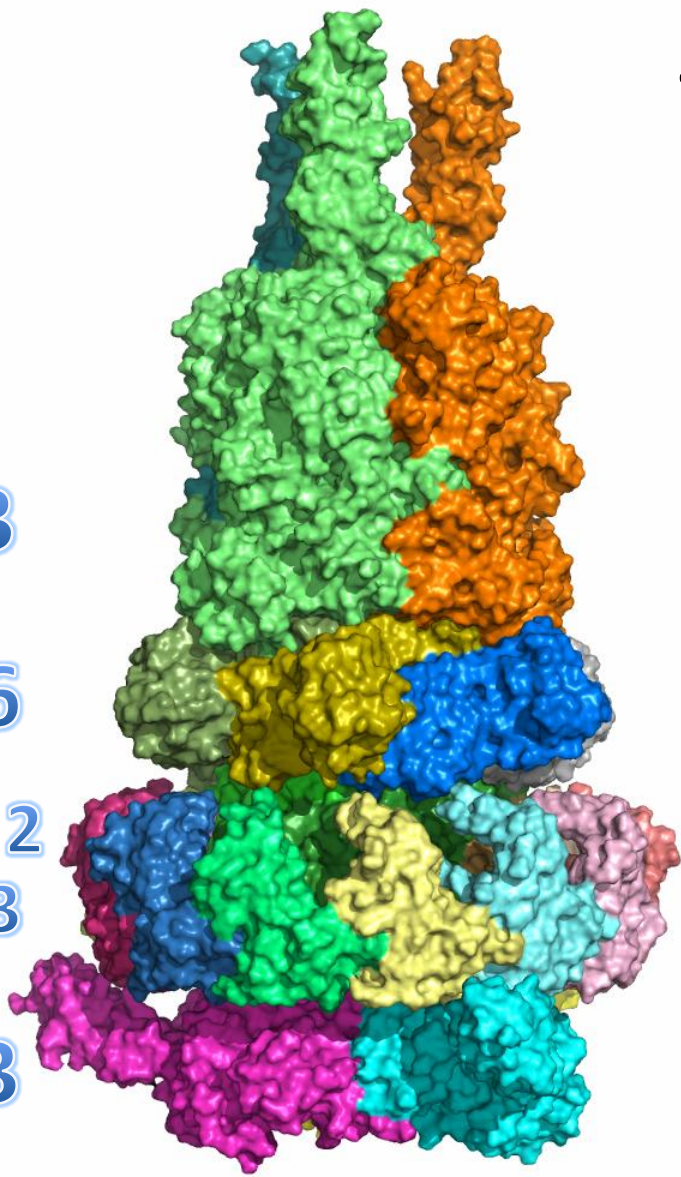
E3

D6

C12

B3

A3

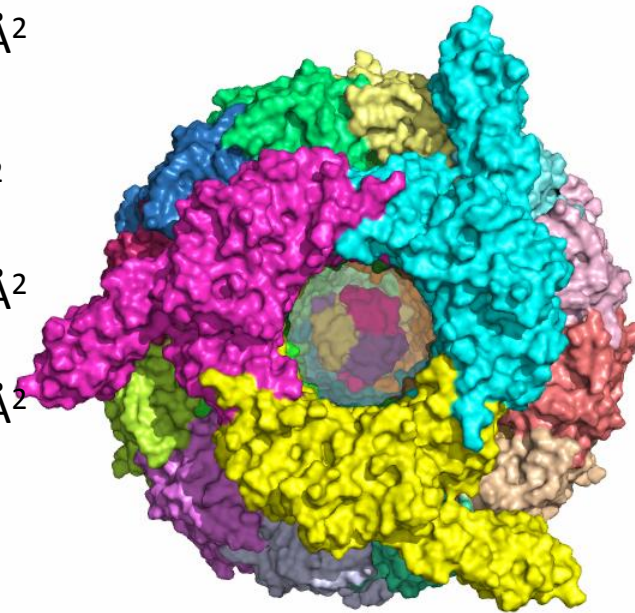


12000 Å<sup>2</sup>

9800 Å<sup>2</sup>

15000 Å<sup>2</sup>

11000 Å<sup>2</sup>

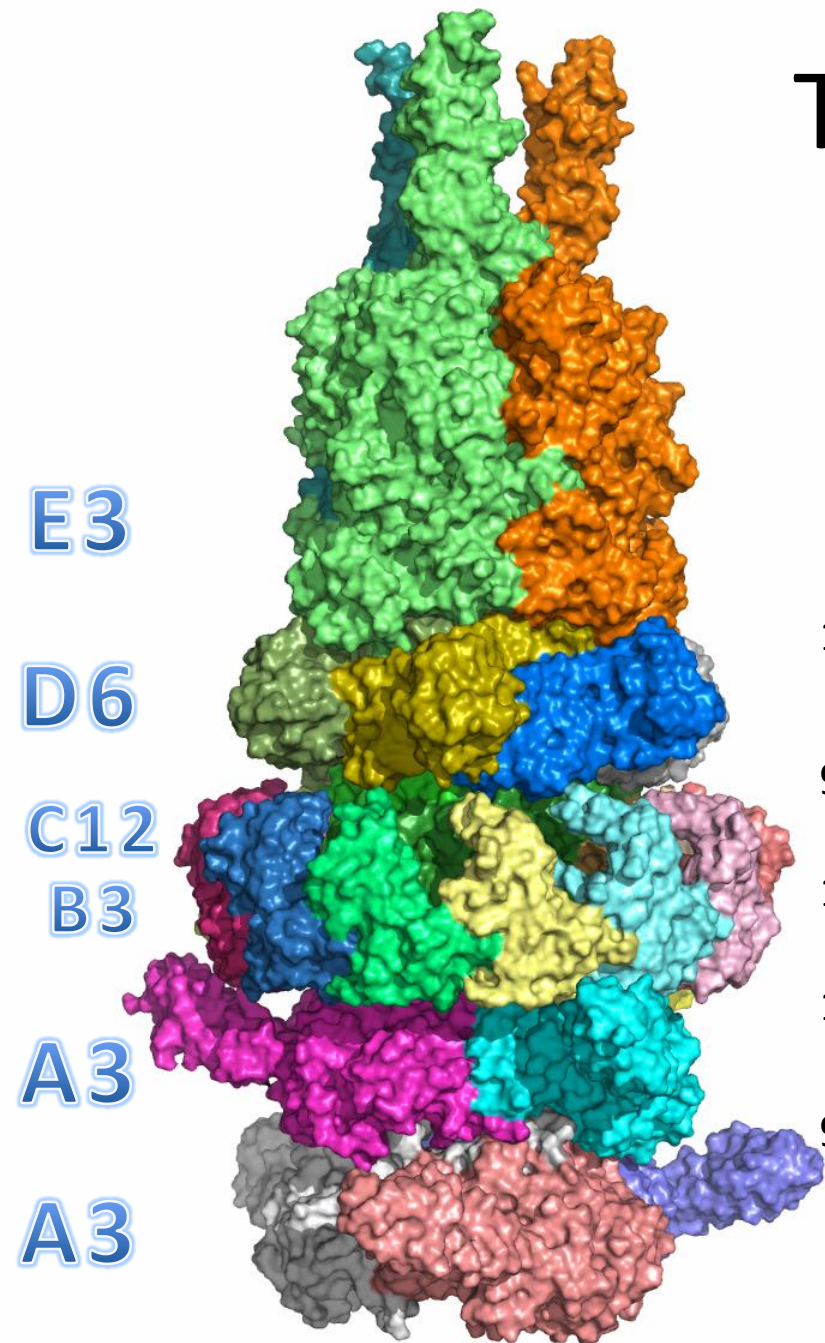


# The phage tail

T170

H1060

N/A



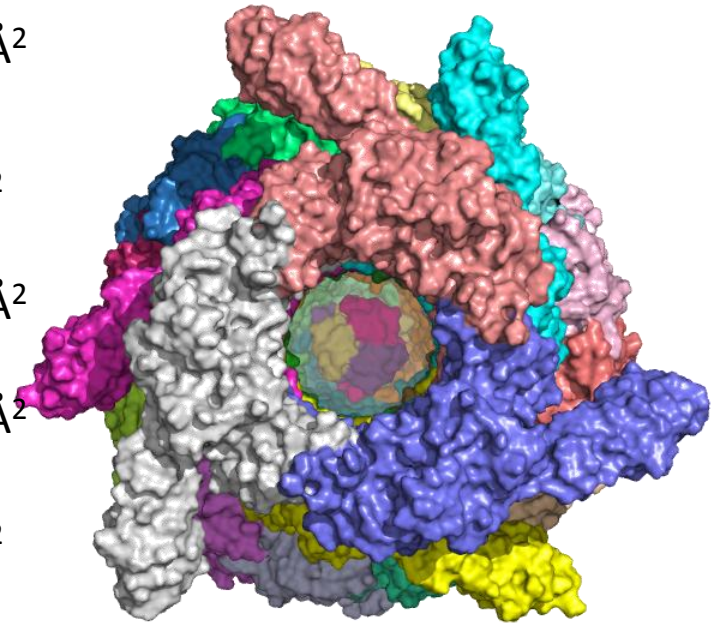
12000 Å<sup>2</sup>

9800 Å<sup>2</sup>

15000 Å<sup>2</sup>

11000 Å<sup>2</sup>

9900 Å<sup>2</sup>





T170

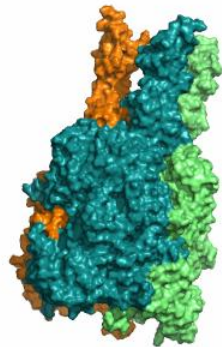
H1060

N/A

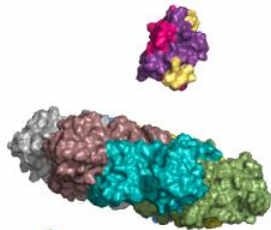
# The phage tail

Each interface between rings  $\approx 10000 \text{ \AA}^2$

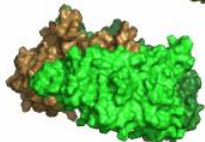
E3F3



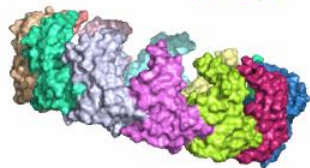
D6



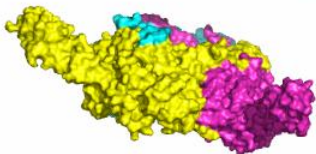
B3



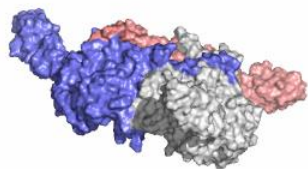
C12



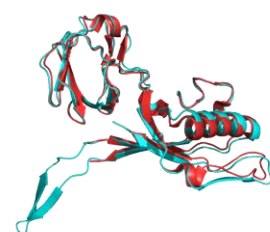
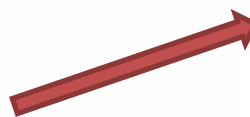
A3



A3



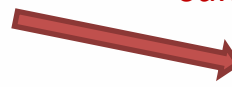
C12  $\approx 15000 \text{ \AA}^2$ , but not connected to A3 or D6



Template	6f2m
RMSD	0.9 Å
Seq.ID	94%

**EASY**

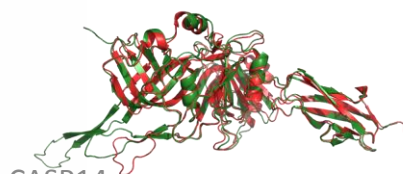
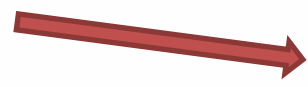
*N-terminal 370 residues of the same template*



Template	5ngj
RMSD	7.1 Å
Seq.ID	10%

**EASY**

*Template is monomeric (dimeric through crystal contacts)*



CASP14

Template	5ngj
RMSD	0.9 Å
Seq.ID	100%

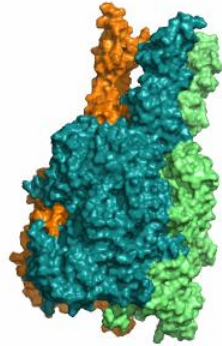
T170
H1060
N/A

# The phage tail

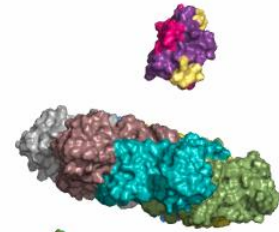
Each interface  
between rings  
 $\approx 10000 \text{ \AA}^2$

*C12  $\approx 15000 \text{ \AA}^2$ , but not  
connected to A3 or D6*

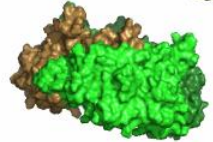
E3F3



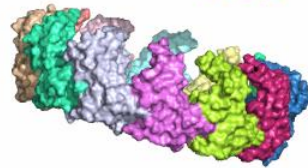
D6



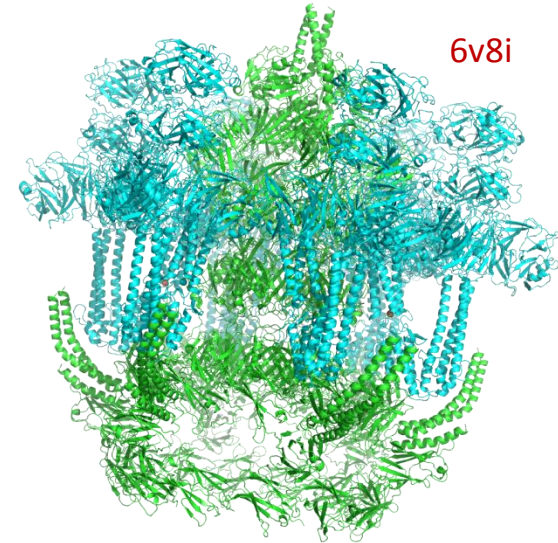
B3



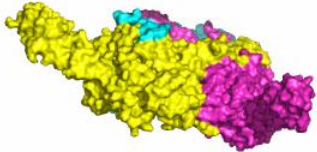
C12



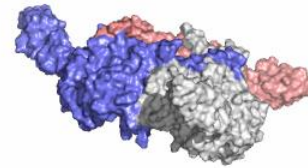
**DIFFICULT**



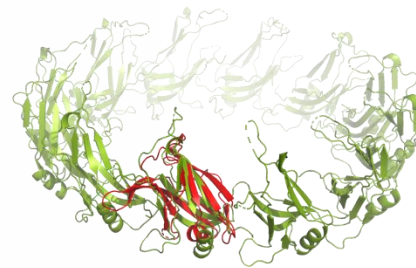
A3



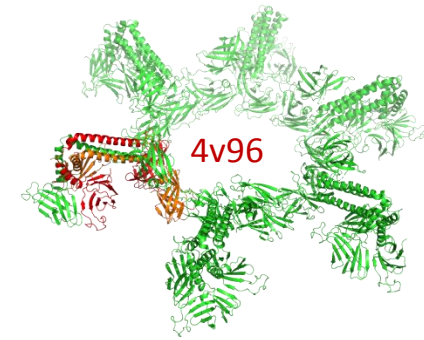
A3



Template	3uh8	4v96 (EM)	6v8i (EM)
RMSD	6.4 Å	3.8 Å	7.7 Å
Seq.ID	11%	9%	15%



3uh8  
CASP14



4v96

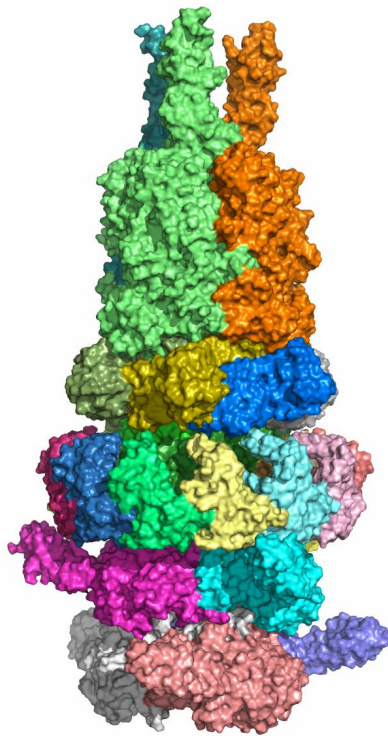
T170

H1060

N/A

# The phage tail

D  
Z C  
B  
A



1:2:3:4:5:6 (204)

J:K:L:M:N:O:P:Q:R:S:T:U (140)  
G:H:I (298)

D:E:F (464)

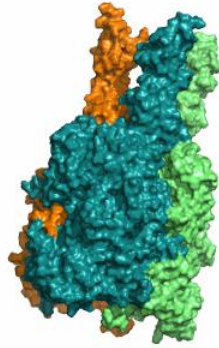
A:B:C (464)

Interface		Chains	Area/Å <sup>2</sup>	Difficulty
9	CD	G:2	750	Easy
8	D	1:2	1200	Easy
7	CZ	H:K	550	Difficult
6	CZ	H:L	680	Difficult
5	Z	K:L	680	Difficult
4	BC	E:G	980	Easy
3	C	G:H	1650	Easy
2	AB	A:E	1650	Easy
1	A	A:B	1800	Easy

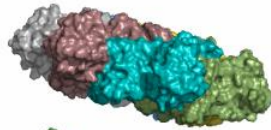
T170

H1060

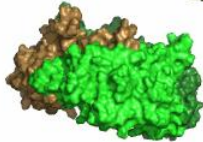
N/A



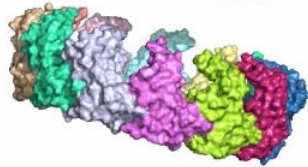
D



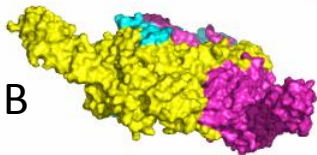
C



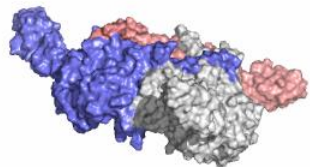
Z



B



A

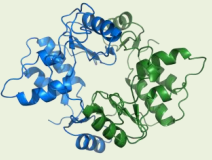
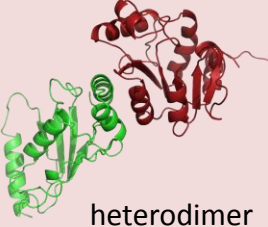


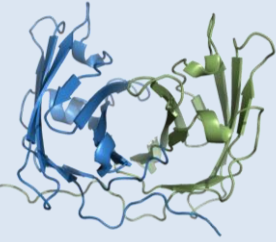
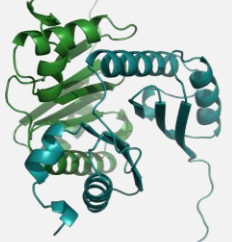
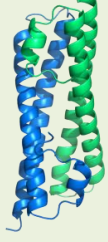
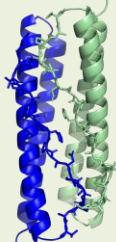
Interface	Rings	Difficulty	Performance
1	A:A	Easy	13/5**
2	A:B	Easy	1
3	C:C	Easy	5
4	B:C	Easy	4
5	Z:Z	Difficult	12/5**
6	C:Z	Difficult	1
7	C:Z	Difficult	1
8	D:D	Easy	7/1**
9	C:D	Easy	6

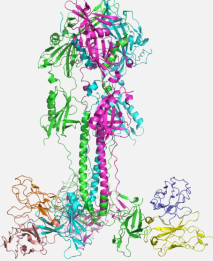
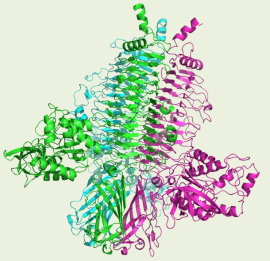
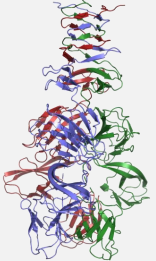
	Group
1	HDOCK, Huang, MDOCKPP, Shen, Zou, Chang, Venclovas, Seok, Fernandez-Recio, Kozakov/Vajda, Kihara, Nakamura, CLUSPRO, Takeda-Shitaka, DATE
2	Fernandez-Recio
3	Venclovas, Seok, MDOCKPP, Zou, Shen
4	HDOCK, Huang, Venclovas, Chang
5	Chang, Shen, Nakamura, Kozakov/Vajda, CLUSPRO, DATE, HDOCK, Huang, Venclovas, Kihara, Seok, Grudinin, Vakser, Baker, VoroCNN-select
6	Shen
7	Kihara
8	Venclovas, Baker, Chang, LZERD, Grudinin, Seok, Shen, Kihara
9	Huang, HDOCK, Shen, Chang, Kihara, Seok, Takeda-Shitaka

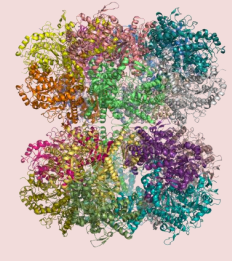
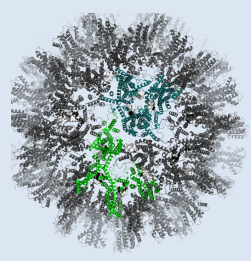

Group	1	2	3	4	5	6	7	8	9		Score
Shen	**		*		**	*		*	*	6/2**	8
Venclovas	*		*	*	*			**		5/1**	6
Chang	*			*	**			*	*	5/1**	6
<b>HDOCK</b>	**			*	*				*	4/1**	5
Huang	**			*	*				*	4/1**	5
Seok	*		*		*			*	*	5	5
Kihara	*				*		*	*	*	5	5
<b>CLUSPRO</b>	*				**					2/1**	3
<b>MDOCKPP</b>	**		*							2/1**	3
Zou	**		*							2/1**	3
Kozakov/Vajda	*				**					2/1**	3
Nakamura	*				**					2/1**	3
<i>Takeda-Shitaka</i>	*								*	2	2
Fernandez-Recio	*	*								2	2
Grudinín					*			*		2	2
<i>DATE</i>	*									1	1
Vakser					*					1	1
<b>LZERD</b>								*		1	1
	15/5**	1	5	4	12/5**	1	1	7/1**	7		
	A	A:B	C	B:C	Z	C:Z	C:Z	D	C:D		
	Easy	Easy	Easy	Easy	Diff.	Diff.	Diff.	Easy	Easy		

# Summary

	 heterodimer
19/3**	17/1***/12**
Two easy dimers	

			
4	No acceptable	19/1**	13/1**
Four difficult dimers			

antibody binding 		
No acceptable	24/21**	No acceptable
Three trimers		

		phage tail 
33/1***/22**	15	0 - 1 - 6
Three big assemblies		

Predictor performance

$$\text{Score} = \omega_1 \cdot N_{\text{ACC}} + \omega_2 \cdot N_{\text{MED}} + \omega_3 \cdot N_{\text{HIGH}}$$

$$\omega_1 = 1; \omega_2 = 2; \omega_3 = 3$$

Best top-5 prediction

Same formula applied for multi-interface targets, divided by number of interfaces

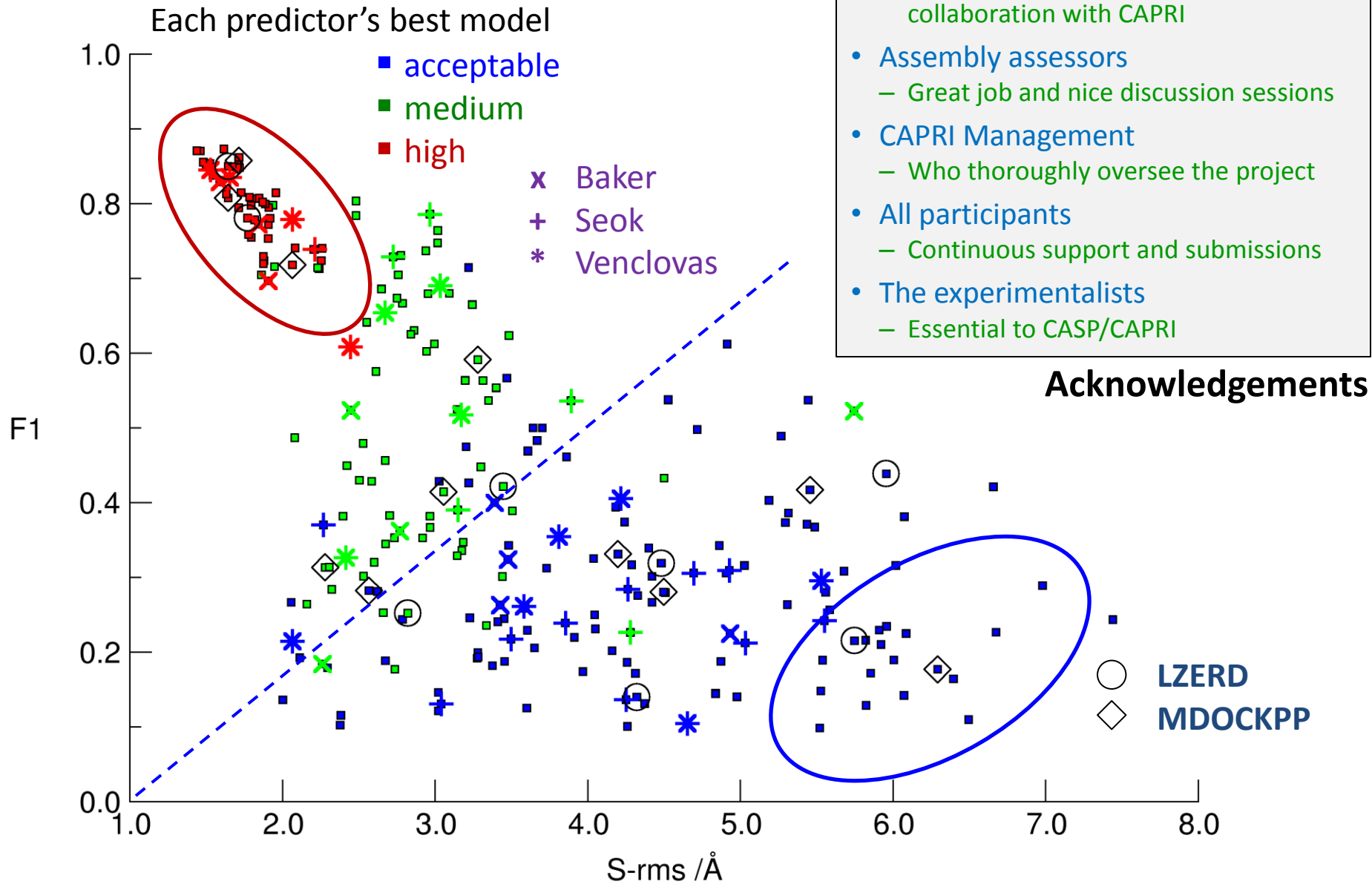
Rank	Group	Performance	Participation	Score
1	Seok	9/4**	16	13
	Venclovas, <i>Baker</i>	8/1***/3**	16	13
4	Zou, Chang	8/3**	16	11
	<b>MDOCKPP</b>	7/1***/2**	16	11
7	Pierce, Kihara	7/3**	15, 16	10
9	Huang, Bates, Kozakov/Vajda, Fernandez-Recio	5/3**	16	8
	<b>LZERD</b> , <i>CoDock</i>	6/2**	16, 12	8
15	Shen	6/1**	16	7
	Vakser	4/3**	16	7
	<i>Takeda-Shitaka</i>	4/1***/1**	16	7
18	<b>GALAXYPPDOCK</b>	5/1**	16	6
	<b>SWARMDOCK</b>	4/2**	16	6
	<i>Lamoureux</i>	3**	13	6
21	Nakamura	3/2**	13	5
22	<b>HDOCK, CLUSPRO</b>	3/1**	16	4
24	Czaplewski, Liwo, <i>DellaCorte, UNRES, DATE</i>	2/1** or 3	6 – 16	3

Ranking does not change significantly if Top-10 is considered, Top-1 gives:

- 1 *Baker*
- 2 Seok
- 3 Venclovas, Chang, Pierce
- 6 Zou, Kihara, Huang

Rank	Group	Performance	Participation	Score
	Seok	9/4**	16	13
	Venclovas, <i>Baker</i>	8/1***/3**	16	13
	Zou, Chang	8/3**	16	11
1	<b>MDOCKPP</b>	7/1***/2**	16	11
	Pierce, Kihara	7/3**	15, 16	10
	Huang, Bates, Kozakov/Vajda, Fernandez-Recio	5/3**	16	8
2	<b>LZERD, CoDock</b>	6/2**	16, 12	8
	Shen	6/1**	16	7
	Vakser	4/3**	16	7
	<i>Takeda-Shitaka</i>	4/1***/1**	16	7
3	<b>GALAXYPPDOCK</b>	5/1**	16	6
	<b>SWARMDOCK</b>	4/2**	16	6
	<i>Lamoureux</i>	3**	13	6
	Nakamura	3/2**	13	5
4	<b>HDOCK, CLUSPRO</b>	3/1**	16	4
	Czaplewski, Liwo, <i>DellaCorte, UNRES, DATE</i>	2/1** or 3	6 – 16	3





# Conclusions

- The best predictors stand out by producing acceptable models for difficult targets
  - They do a good job in side-chain modeling
  - Shared 1<sup>st</sup> place for Baker, Seok and Venclovas
    - One more acceptable target for Seok; Venclovas significantly improved scoring
  - Everybody can do Template-based “docking”, but naïve modeling did not do very well this time
  - MDOCKPP is the best-performing server, on-par with human predictors
- Human predictors do better than their server counterparts
  - Most docking servers still require a significant amount of human input
- Most targets were obligate
  - Templates work well; flexibility and conformational change less an issue

