RNA Structure: CASP perspective

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12 December, 2022 CASP15 conference Antalya, Turkey



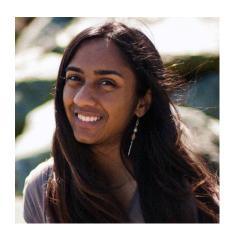












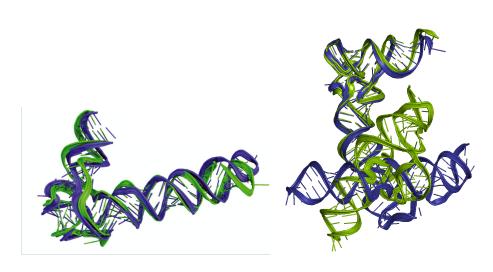
Ramya Rangan

We're predictors, experimentalists, and (now) assessors.

Thanks: Andriy Kryshtafovych, Krzystof Fidelis, John Moult

An RNA category in CASP15

3D **RNA deep learning** in its nascency



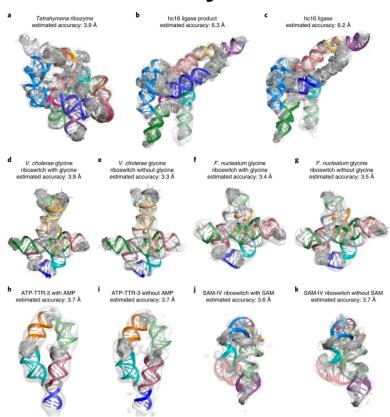
RNA Puzzle 24

RNA Puzzle 28

Townshend, Eismann, Watkins, ..., Das, Dror, Science, 2021

Thanks, Eric Westhof, Chichau Miao and RNA Puzzles community

More targets: increasing throughput of RNA Cryo-EM

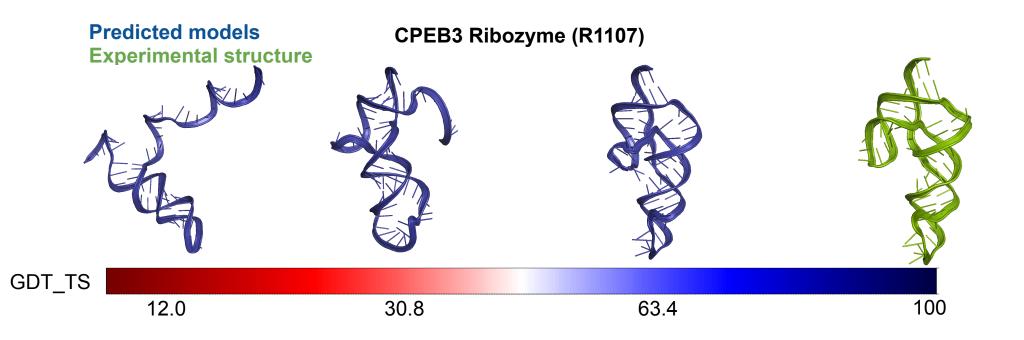


Kappel, Zhang, Su, ..., Chiu, Das, Nat. Methods, 2020

CASP15: 12 targets, excellent models from a field of 40 predictors Atomically ordered RNA's Larger, more flexible RNA's **Best predicted model Experimental structure** X-ray structures (cryo-EM) R1117TS287_5 R1107TS232 1 R1108TS232_ R1126TS232_5 R1156TS128_5 R1116TS285_5 R1149TS110_2 RNA-protein complex (cryo-EM) R1136TS232_3 R1138TS232_4 R1128TS232_1 R1190TS081_3 R1189TS444 3

What do CASP-style quantitative rankings say?

Piloting GDT for RNA



CASP15 RNA assessment metric

"Topology" "Local environment"

Stereochemical quality

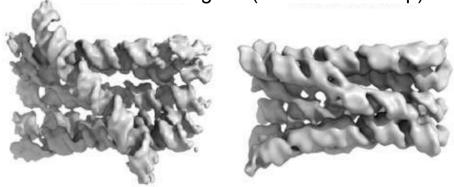
$$Z_{RNA} = \frac{1}{3}[Z_{TM} + Z_{GDT-TS}] + \frac{1}{8}[Z_{INF} + Z_{IDDT}] + \frac{1}{12}Z_{clash}$$

where Z-score = number of standard deviations from the mean (after filtering out of poor models with initial Z < -2)

Used in prior CASP topology assessment Used in (non-CASP) RNA assessment

How we assessed multi-state RNA's

R1138 RNA origami (with a kinetic trap)



R1156 Bt-CoV-HKU5 SL5 domain



Compare:

all five predictor models

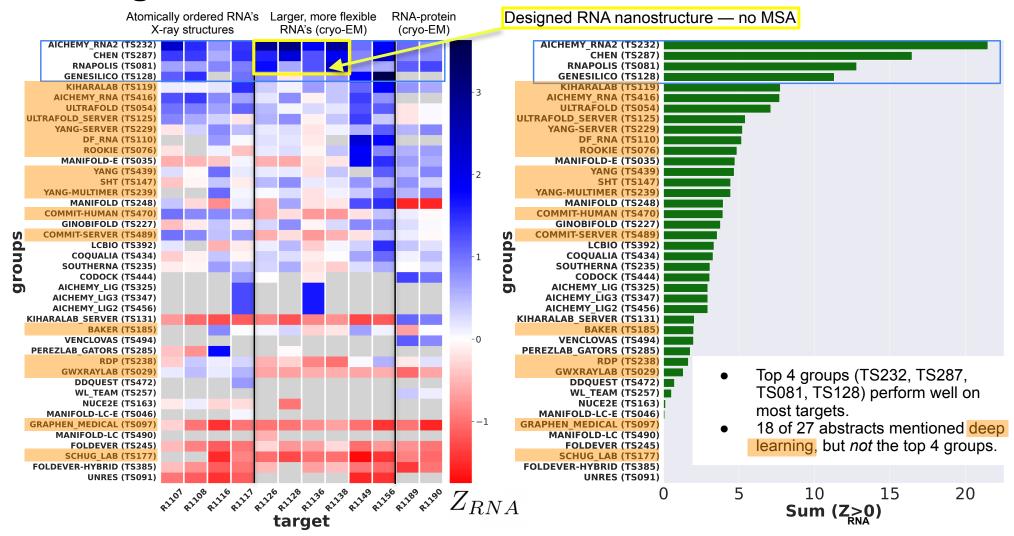
vs.

all available experimental models

Reward predictor based on *best* score.

Thanks: Lisa Kinch, Nick Grishin

Rankings from CASP15 RNA metric



Let's double-check our rankings

- Topological scores all consistent in ranking top four groups
 - In local environment scores, top 3 remain top 3; after that ranking is different from topological scores
- Some top rankers were not the cleanest in terms of clash score, but were also not the large negative outliers
- Comparisons directly to EM maps (to avoid experimental model bias) give same top 4 ranking.

$$Z_{EM} = \frac{1}{5} [Z_{CCmask} + Z_{CCpeaks} + Z_{SMOC} + Z_{MI} + Z_{AI}]$$

| | ZRNA | Ztopo | ТМ | GDT-TS | RMSD | Zlocal | INF | IDDT (| clashscore | EM |
|---|------|-------|----|--------|------|--------|-----|--------|------------|----|
| AICHEMY_RNA2 (TS232) | i | 1 | 1 | 1 | ĺ | 1 | 1 | 1 | 21 | 1 |
| CHEN (TS287) | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 1 | 3 |
| RNAPOLIS (TS081) | 3 | 3 | 4 | 3 | 3 | 3 | 3 | 3 | 16 | 4 |
| GENESILICO (TS128) | 4 | 4 | 3 | 4 | 4 | 6 | 6 | 5 | 23 | 2 |
| KIHARALAB (TS119) | 5 | 6 | 7 | 5 | 9 | 5 | 4 | 6 | 6 | 7 |
| AICHEMY_RNA (TS416) | 6 | 5 | 5 | 6 | 11 | 7 | 8 | 7 | 13 | 10 |
| ULTRAFOLD (TS054) | 7 | 7 | 6 | 7 | 7 | 4 | 5 | 4 | 8 | 11 |
| ULTRAFOLD_SERVER (TS125) | 8 | 12 | 10 | 13 | 10 | 8 | 7 | 8 | 9 | 8 |
| YANG-SERVER (TS229) | 9 | 9 | 12 | 9 | 5 | 10 | 16 | 9 | 25 | 17 |
| DF_RNA (TS110) | 10 | 8 | 13 | 8 | 23 | 17 | 19 | 13 | 28 | 6 |
| ROOKIE (TS076) | 11 | 14 | 14 | 14 | 20 | 11 | 13 | 12 | 12 | 5 |
| YANG (TS439) | 12 | 15 | 18 | 11 | 6 | 9 | 9 | 10 | 18 | 16 |
| YANG-MULTIMER (TS239) | 13 | 13 | 22 | 10 | 8 | 13 | 18 | 11 | 26 | 18 |
| MANIFOLD-E (TS035) | 14 | 10 | 15 | 12 | 21 | 20 | 23 | 17 | 11 | 9 |
| SHT (TS147) | 15 | 19 | 17 | 19 | 15 | 18 | 15 | 19 | 3 | |
| COMMIT-HUMAN (TS470) | 16 | 11 | 8 | 15 | 13 | 14 | 11 | 14 | 39 | 21 |
| MANIFOLD (TS248) | 17 | 17 | 19 | 17 | 27 | 24 | 24 | 21 | 15 | 20 |
| COMMIT-SERVER (TS489) | 18 | 16 | 11 | 16 | 14 | 15 | 12 | 16 | 41 | 22 |
| GINOBIFOLD (TS227) | 19 | 20 | 9 | 20 | 18 | 19 | 17 | 20 | 4 | |
| LCBIO (TS392) | 20 | 21 | 21 | 22 | 16 | 12 | 10 | 18 | 19 | 19 |
| COQUALIA (TS434) | 21 | 22 | 20 | 27 | 19 | 21 | 20 | 23 | 5 | 15 |
| COQUALIA (TS434) COUTHERNA (TS235) COUTHERNA (TS235) COUTHERNA (TS2347) | 22 | 18 | 16 | 18 | 12 | 16 | 14 | 15 | 22 | |
| AICHEITI_EIOS (15547) | 23 | 27 | 26 | 25 | 30 | 26 | 26 | 25 | 34 | 13 |
| AICHEMY_LIG2 (TS456) | 24 | 26 | 25 | 24 | 28 | 27 | 27 | 26 | 36 | 12 |
| AICHEMY_LIG (TS325) | 25 | 25 | 27 | 23 | 29 | 25 | 25 | 24 | 35 | 14 |
| CODOCK (TS444) | 26 | 23 | 24 | 21 | 26 | 22 | 21 | 22 | 27 | |
| BAKER (TS185) | 27 | 31 | 31 | 30 | 22 | 23 | 22 | 28 | 10 | |
| KIHARALAB_SERVER (TS131) | 28 | 28 | 29 | 28 | 33 | 29 | 28 | 29 | 33 | |
| VENCLOVAS (TS494) | 29 | 29 | 30 | 29 | 32 | 30 | 29 | 30 | 31 | |
| RDP (TS238) | 30 | 24 | 23 | 26 | 17 | 28 | 30 | 27 | 42 | |
| PEREZLAB_GATORS (TS285) | 31 | 30 | 32 | 31 | 31 | 31 | 31 | 31 | 24 | |
| GWXRAYLAB (TS029) | 32 | 32 | 28 | 32 | 35 | 35 | 42 | 33 | 7 | |
| DDQUEST (TS472) | 33 | 33 | 33 | 34 | 37 | 34 | 39 | 34 | 40 | |
| WL_TEAM (TS257) | 34 | 34 | 34 | 35 | 36 | 33 | 32 | 36 | 30 | |
| NUCE2E (TS163) | 35 | 35 | 35 | 33 | 25 | 32 | 33 | 32 | 38 | |
| MANIFOLD-LC-E (TS046) | 36 | 36 | 38 | 36 | 39 | 41 | 40 | 41 | 29 | |
| FOLDEVER-HYBRID (TS385) | 37 | 37 | 39 | 38 | 34 | 36 | 34 | 35 | 17 | |
| FOLDEVER (TS245) | 38 | 38 | 36 | 39 | 24 | 37 | 35 | 37 | 14 | |
| SCHUG_LAB (TS177) | 39 | 39 | 37 | 40 | 38 | 38 | 36 | 38 | 37 | |
| GRAPHEN_MEDICAL (TS097) | 40 | 40 | 40 | 37 | 41 | 39 | 37 | 39 | 2 | |
| UNRES (TS091) | 41 | 41 | 41 | 41 | 42 | 40 | 38 | 40 | 20 | |
| MANIFOLD-LC (TS490) | 42 | 42 | 42 | 42 | 40 | 42 | 41 | 42 | 32 | |

Thanks: Maya Topf, Tom Mulvaney, Andriy Kryshtafovych

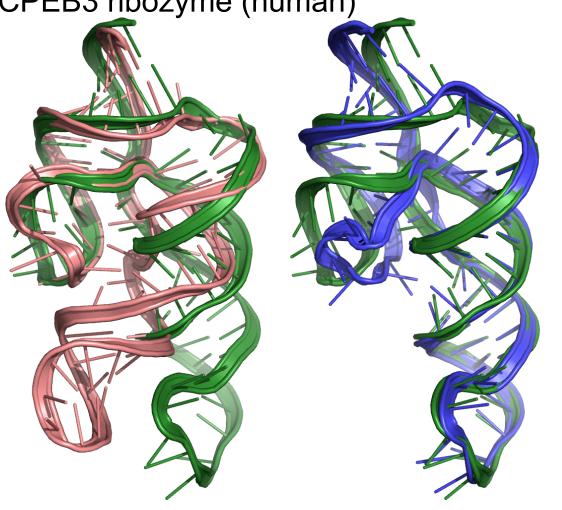


R1107 CPEB3 ribozyme (human)

Rosetta template-based models 1 (Prepared by Ramya in 2019)

Crystal structure (of dimer)

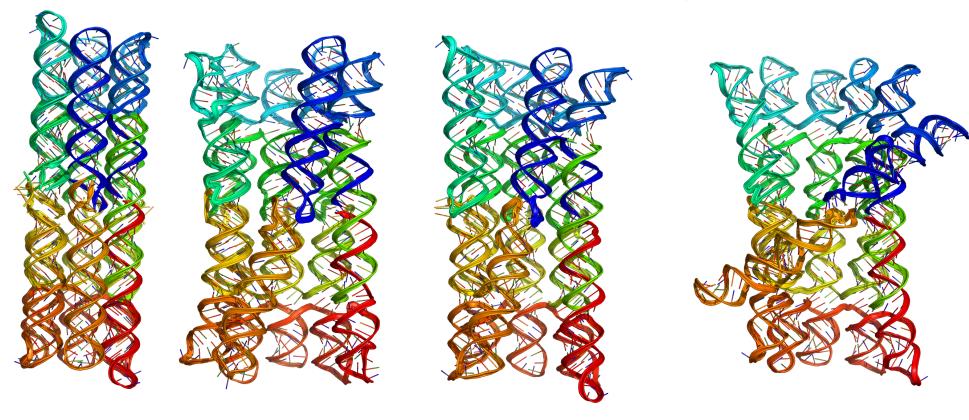
Alchemy-RNA2 (TS232) Model 1



Przytula-Mally et al., bioRxiv (2022)

Thanks: Masquida, Sigel groups; Eric Westhof

R1138 Six-helix bundle RNA origami



Design
TM-score 0.623

Cryo-EM structure

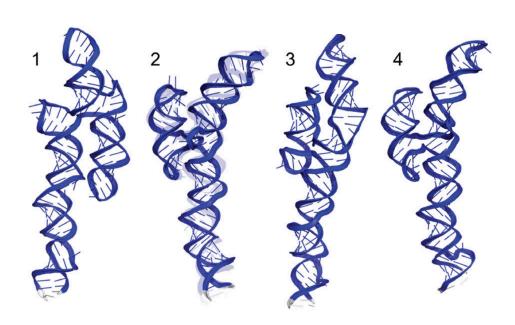
Alchemy-RNA2 (TS232) Model 4

TM-score 0.800

Another resolved Cryo-EM structure [no CASP models were within TM-score of 0.63]

Thanks: Ebbe Anderson and colleagues

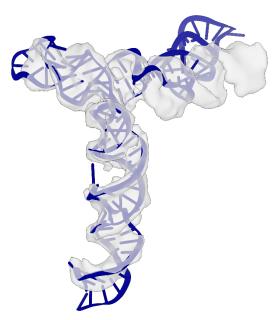
R1149 SARS-CoV-2 SL5 domain



Prior Rosetta FARFAR2 modeling suggested no well-defined 3D structure for this sequence

Rangan et al., NAR (2021)

GeneSilico (TS128) Model 1

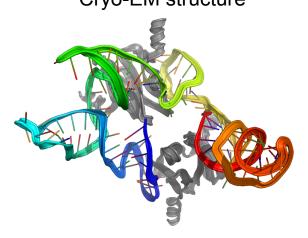


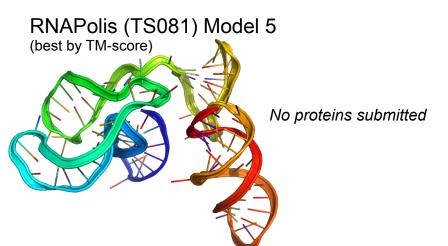
The domain *is* resolvable by cryo-EM
A "T-shape" not an "H-shape"
Fold captured by some CASP predictors
Similar for R1156 (more flexible bat CoV homolog)

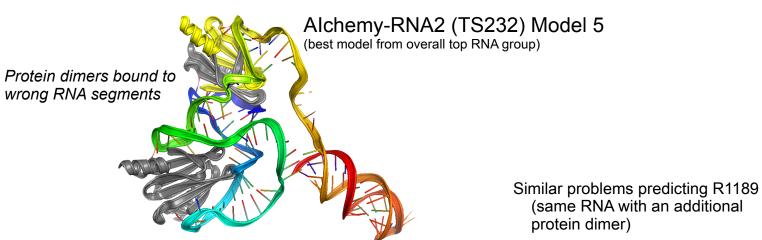
Thanks: Rachael, Wah Chiu and collaborators

R1190 CsrA RNA-protein complex

Cryo-EM structure







Thanks, Zhaoming Su

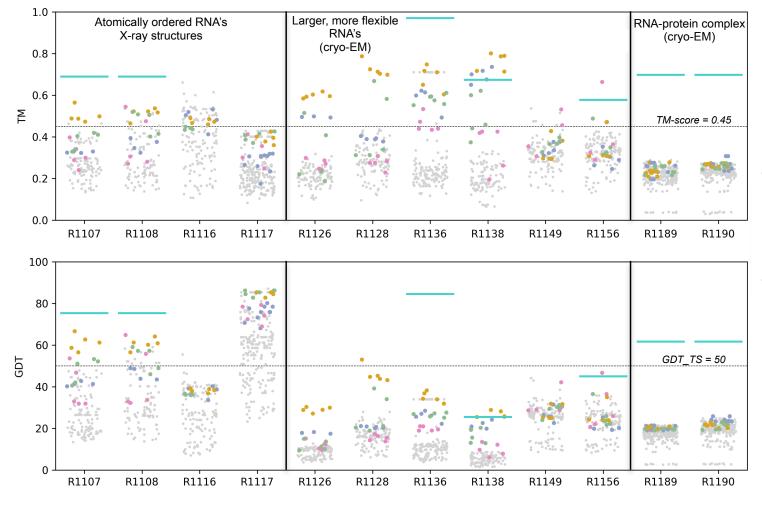
In an absolute sense, how did CASP15 RNA modelers do?

TM-score > 0.45 means 'matching template' (for RNA) GDT_TS > 50 means 'correct topology' (for proteins)

S. Gong, C. Zhang, and Y. Zhang, Bioinformatics (2019)

CASP9 (Kinch et al.), CASP10 (Tai et al.), CASP11 (Abriata et al.)... [CASP topology assessment papers]

GDT and **TM** summary



- AICHEMY_RNA2 (TS232)
- CHEN (TS287)
- RNAPOLIS (TS081)
- GENESILICO (TS128)
- Other groups
- All 10 RNA-only targets have submissions with TM-score > 0.45 and/or GDT_TS > 50
- Similarity to experimental structure typically worse than similarity between alternative experimentally captured conformations

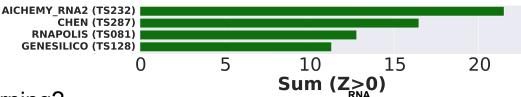
Summary

12 interesting targets (most from cryo-EM)

$$Z_{RNA} = \frac{1}{3}[Z_{TM} + Z_{GDT-TS}] + \frac{1}{8}[Z_{INF} + Z_{IDDT}] + \frac{1}{12}Z_{clash}$$

40 predictors (most new to RNA)

Classic CASP metrics are useful for RNA assessment, even flexible targets



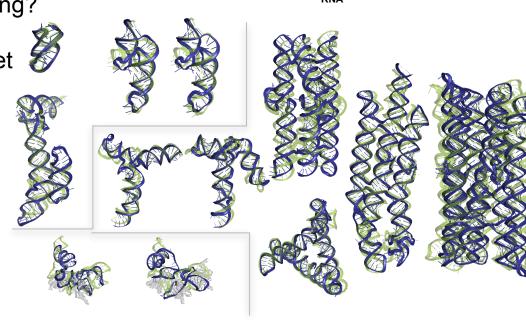
Top four predictors unambiguous. No deep learning?

At least one good model for each RNA-only target

- Clear refinement over template or designed structure
- Achieved in all cases: TM-score > 0.45 or GDT > 50
- Similarity to closest experimental structure typically worse than similarity between alternative experimental structures

RNA-protein complexes remain challenging*

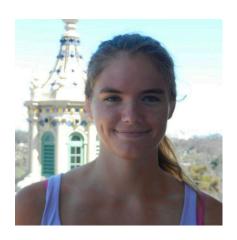
*And maybe atomically ordered new RNA folds?



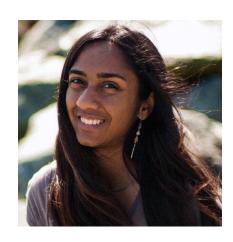
Thank you!

Eric Westhof, Chichau Miao, Marta Szachniuk, Maciej Antczak, Maya Topf, Tom Mulvaney, Gabriel Studer, Marcin Magnus, Adam Zemla, Chengxin Zhang, Nick Grishin, Lisa Kinch, Andriy Kryshtafovych, Krzystof Fidelis, John Moult









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